

Available online at www.sciencedirect.com



C. R. Biologies 332 (2009) 480-488

COMPTES RENDUS BIOLOGIES

http://france.elsevier.com/direct/CRASS3/

Evolution / Évolution

# Morphological identification of sibling species: the case of West African *Mastomys* (Rodentia: Muridae) in sympatry

Aude Lalis<sup>a,b,\*</sup>, Allowen Evin<sup>a,b</sup>, Christiane Denys<sup>a</sup>

<sup>a</sup> UMR CNRS 5202 – USM 601, Origine, structure et évolution de la biodiversité, Département systématique et évolution, Muséum national d'histoire naturelle, CP 51, 55, rue Buffon, 75005 Paris, France

<sup>b</sup> Plate-forme Morphométrie MNHN – CNRS IFR 101, 45, rue Buffon, 75005 Paris, France

Received 9 May 2008; accepted after revision 20 November 2008

Available online 9 January 2009

Presented by Pierre Buser

# Abstract

In this study, we investigate skull size and shape differentiation between sibling species of *Mastomys* with the aim to characterize and discriminate three sympatric species found in West Africa: *M. huberti, M. erythroleucus* and *M. natalensis*. A total of 133 genetically determined specimens were used for the morphometric analyses. Statistical analyses clearly demonstrated that the three species largely overlapped in centroid size (*M. erythroleucus* tends to be larger on average than the *M. huberti* and *M. natalensis*) but they exhibited large differences in skull shape. The current study focused on skull shape, and allowed us to discriminate three morphological groups that are congruent with the three species suggested by molecular identification (90% of the individuals are correctly assigned by cross-validated classifications). In the *Mastomys*, the evolution of cranial length and shape may be influenced by competitive pressure between closely related species separated by ecological segregation. This source of variability could possibly induce character displacement between species of *Mastomys*. *To cite this article: A. Lalis et al., C. R. Biologies 332 (2009).* 

© 2009 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

# Résumé

Identification morphologique d'espèces jumelles : le cas de *Mastomys* (Rodentia : Muridae) en sympatrie. La différenciation morphométrique entre trois espèces jumelles de *Mastomys* d'Afrique de l'Ouest a été étudiée à partir de 133 spécimens, en recherchant les caractères diagnostiques crâniens chez *M. huberti, M. erythroleucus* et *M. natalensis* en sympatrie. L'étude de morphométrie géométrique a permis de mettre en évidence des différenciations associées à des phénomènes d'adaptation à différents biotopes. La légère différence observée dans la taille du crâne entre les espèces (*M. erythroleucus* présentant une taille crânienne supérieure à celle observée chez *M. natalensis* ou *M. huberti*) ne constitue pas un critère fiable d'identification. En revanche, les principales différences de formes observées entre les espèces révèlent une variabilité dans le processus de mastication, qui pourrait résulter d'une compétition dans la disponibilité des ressources et qui aurait favorisé l'émergence de traits différents par déplacement de caractères et spécialisation pour un nouveau type d'habitat. L'analyse des composantes de formes a donc permis de confirmer la présence d'unités morphologiques partiellement distinctes pour les trois espèces retrouvées en sympatrie. *Pour citer cet article : A. Lalis et al., C. R. Biologies 332 (2009).* 

<sup>\*</sup> Corresponding author at: UMR CNRS 5202 – USM 601, Origine, structure et évolution de la biodiversité, Département systématique et évolution, Muséum national d'histoire naturelle, CP 51, 55, rue Buffon, 75005 Paris, France.

E-mail address: lalis@mnhn.fr (A. Lalis).

© 2009 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

Keywords: Sibling species; Interspecific variability; Skull shape; Geometric morphometrics; Mastomys

Mots-clés: Espèces jumelles; Variabilité interspécifique; Conformation crânienne; Morphométrie géométrique; Mastomys

# 1. Introduction

In taxonomy, species identification may rely on the morphospecies, or typological, concept which has been used since the beginning of systematics [1]. This concept has been commonly used in palaeontology and zoology for a lot of species described only on the basis of diagnostical morpho-anatomical characters. However, the recent development of various other systematic techniques (DNA molecular sequencing and morphometrics) and new diagnostic characters like karyotypes or chromosome banding patterns have allowed the description of new species [2–4]. It has been reported that karyotyped data can provide good evidence of probable reproductive isolation at specific level, without any sign of morphological discrimination even by more sophisticated methods of morphometrics [5,6]. In other cases, analyses of size and shape by conventional or geometric morphometrics have suggested specific separation [7-10]. In West Africa, the multimammate rat Mastomys is the dominant genus from the rodent communities [11–13]. Using its exceptional ability to adapt and reproduce, this rodent is generally found in all habitats from houses to forests. The taxonomy of the genus has been problematic due to the high morphological convergence between the species which prevents correct assignation to a specific level by traditional systematic analyses [14]. The determination key of *Mastomys* is based on various morphological features like the mammae number (between 5 to 12 pairs); sperm and penis morphology and dimensions [15,16]; karyotype characters such as the diploid number (2N); and the fundamental number (FN). The use of cytogenetic and molecular tools has recently highlighted presence of many sibling species within the genus, especially in West Africa [14]. There is now a consensus to consider M. natalensis (2n = 32, NFa = 52-54), M. huberti (2n = 32, NFa = 52-54), M.NFa = 44) and *M. erythroleucus* (2n = 38, NFa = 54-56). Few phylogenetic analyses have been devoted to this genus. Karyological analyses by Britton-Davidian et al. [17] found M. natalensis to be closer M. huberti, while M. erythroleucus was the basal taxon of this clade. Protein electrophoresis data placed M. erythroleucus closer to M. huberti and both well separated from M. natalensis, while immunological data and DNA/DNA hybridization did not provide species discrimination [18].

A cytochrome-b phylogeny found Mastomys genus paraphyletic with a well supported monophyletic group including M. huberti, M. erythroleucus and M. natalensis [19]. All these results are incongruent, emphasizing the difficulty of classification among these taxa. Duplantier [20] tried to find other morphometric and genetic characteristics, but was unable to completely separate the three different species in Senegal in his analyses of mandibular and craniometrical distances. For instance, in a discriminant analysis of 15 cranial distance measurements [20], three species groups were shown with percentages of well classified individuals ranging from 75 to 92%. Said study was not able to provide significant criteria to allow for reliable identification. At lower taxonomic levels, morphological divergence can be subtle and traditional morphometrics is sometimes insufficient to differentiate groups, while geometric morphometrics performs better [21,22]. Thus, geometric morphometrics is especially well suited when investigating the evolution of forms among and within species [e.g. [23-26]] as well as to address problems of functional morphology and ecological divergence [27,28]. In this study, we investigate skull size and shape differentiation between closely related species of Mastomys. The goal of the present geometric morphometrics study is to characterize and discriminate three sympatric species of West African Mastomys: M. huberti, M. erythroleucus and M. natalensis.

#### 2. Material and methods

#### 2.1. Specimens

A total of 133 genetically determined (DNA sequencing for cytochrome b) specimens were used for the morphometric analyses. They were classified into three taxa: *M. huberti* (N = 43), *M. erythroleucus* (N = 60), and *M. natalensis* (N = 30). They were trapped at 2 sites in Guinea (Mankountan and Bantou) separated by 426 km. *M. huberti* and *M. natalensis* were only trapped in Mankountan and Bantou respectively whereas *M. erythroleucus* was trapped in both sites (Mankountan N = 30; Bantou N = 30). As in preliminary analyses, it was found that the two populations of *M. erythroleucus* present a significant difference in size (student's *t*-test, F = 4.21, p < 0.01) and in shape

(MANOVA, dorsal side: F = 1.97, p < 0.05; ventral side: F = 1.51, p < 0.05; lateral side: F = 1.28, p < 0.05), they were considered distinct populations in further analyses (ERY 1, Mankountar; ERY 2, Bantou).

After capture, each individual was measured, weighed and sexed. Age class was determined on the basis of body weight following Leirs [29], Granjon et al. [30] and Lalis et al. [31]. The samples were deposited in the collection of the National Museum of Natural History (France NMNH). As we did not find either significant sexual dimorphism differences between groups (2 way ANOVA,  $p_{(dorsal side)} = 0.756$ ;  $p_{(ventral side)} = 0.698$ ;  $p_{(lateral side)} = 0.589$ ) nor differences between groups for the 'age' factor (2 way ANOVA,  $p_{(dorsal side)} = 0.565$ ;  $p_{(ventral side)} = 0.578$ ;  $p_{(lateral side)} = 0.521$ ), all of the groups were pooled together for further analyses.

# 2.2. Geometric morphometrics analyses of size and shape

Three-dimensional coordinates of 25, 35 and 21 landmarks on the dorsal, ventral and lateral sides of skull, respectively, were acquired using a Reflex Microscope (Reflex Measurements Ltd.) (Fig. 1). Coordinates were superimposed using a generalized Procrustes analysis algorithm (GPA) [32,33]. We investigated size and shape parameters separately and pooled both to analyse the form of skull. Size was eliminated by dividing coordinates by the centroid size, which corresponds to the square root of the sum of the squared distances between the centre of the object and each landmark [34]. Residuals of superpositions (= Procrustes residuals) together with centroid size constitute the shape and size variables respectively used in the univariate and multivariate statistical analyses. Possible effects of sex and age were investigated and possible errors of measurement were detected by a preliminary Principal Component Analvsis (PCA) of Procrustes residuals. The analyses were done by pooling the sides.

#### 2.3. Statistical analyses

Differences in the log of centroid size of taxa were depicted by boxplots and their significance tested with analyses of variance (student *t*-tests and ANOVAs) and pairwise comparisons using Bonferroni tests. Canonical Variate Analyses (CVA) has been used to characterize and analyse the shape variability among taxa; Multivariate Analyses of Variance (MANOVAs) to test the significance of differences between taxa. To visualize the shape differences, deformations along factorial axes were calculated by multivariate regressions [35]. In order to facilitate the visualization of shape differences, deformations were amplified by a factor of three. Classification percentages were estimated by multiple discriminant functions using shape parameters and leaveone-out cross validations [36] to test the validity of the a priori taxonomic assignments. Due to the relatively small sample sizes and the large number of variables (81 three-dimension landmarks), statistical analyses of shape were done using the dimension reduction approach advocated in Baylac and Friess [37]. We tested and removed allometries in shapes by a regression between the CVA projections and the log centroid size. The overall phenotypic similarities between taxa ware depicted using a neighbour-joining tree (NJ) computed from the matrix of Mahalanobis's  $D^2$  distances. All calculations were done using with the "R" language version 2.5.0. Morphometric procedures were carried out with the 'Rmorph' library for R [38].

#### 3. Results

#### 3.1. Size

Log centroid-size of the three taxa differed significantly for all views (dorsal side: F = 1.87, p < 0.05; ventral side: F = 1.15, p < 0.05; lateral side: F =1.49, p < 0.05). Indeed, the log centroid size of skulls overlapped largely (Fig. 2) except for the population 'ERY 2' which presents a larger skull size than the others. The intraspecific variability of *M. erythroleucus* was very large.

The link between size and shape is not significant  $(p_{(\text{dorsal})} = 0.187, p_{(\text{ventral})} = 0.288, p_{(\text{lateral})} = 0.886)$ . As the allometries do not disturb the taxa identification, we removed allometries from shape of skull using a multivariate regression.

# 3.2. Shape

Shape analyses were realized using the 15, 17 and 12 first principal components (PCs) extracted from Procrustes residuals which explained 78.4, 74.2 and 68.4% of the total variance for the dorsal, ventral and lateral sides.

MANOVA results revealed highly significant differences between taxa for the three pooled sides (F = 16.28, p < 0.001). The first two axes of the CVA (Fig. 3) explained 53.22% and 13.22% respectively of the shape variance and practically separated all three taxa (Fig. 3). Furthermore, the first axis further partially distinguished *M. huberti* from the two other species,



Fig. 1. Landmarks location for (a) the dorsal, (b) the ventral and (c) the lateral views of the skull. Dorsal view: tip of the nasals (1), most anterior points at nasal-premaxillary suture (2, 3), anterior projection of zygomatic (4, 5), intersection of premaxilla and frontal (6, 7), nasal-frontal suture (8, 9, 25), suture of premaxilla and maxilla over lachrymal capsule (10, 11), frontal-parietal suture (12, 13, 24), most medial point at interorbital constriction (14, 15), back of zygomatic notch (16, 17), intersection of parietal-interparietal and supraoccipital sutures (18, 19), back of the lateroccipital protuberances (20, 21, 22), midpoint of parietal interparietal suture (23). Ventral view: tip of the nasals (1), antero-lateral extremity of left or right incisive alveolus (2, 3), anterior palatine foramen (4, 5), posterior palatine foramen (6, 7), back of zygomatic plate (8, 9), intersection between left or right anterior end of molar and mandible (10, 11), intersection between left or right posterior end of molar and mandible (12, 13), suture between jugal and squamosal in the left or right zygomatic arch (14, 15), front of glenoid fossa on squamosal root of zygomatic arch (16, 17), suture between left or right parietal and squamosal (18, 19), back of external opening of auditory bullae (20, 21), lateral points of sphenoccipital suture (22, 23), styloinastoid foramina at the posterior border of external auditory meatus (24, 25), anterior extremity of occipital (26, 27), posterior intersection between foramen magnum and occipital condyle (28, 29), posterior extremity of foramen magnum (30), anterior extremity of foramen magnum (31), anterior extremity of occipital (32), cranio-pharyngeal canal (33), contact point between palatine and presphenoid (34), contact point between maxilla and palatine (35). Lateral view: tip of nasal (1), upper point of incisors (2), margin of the alveolus at the back of incisors (3), tip of incisors (4), inferior margin of the infraorbital foramen (5), front of the zygomatic plate (6), anterior point at maxillary root of zygomatic (7), front of first molar (8), back of toothrow (9), dorsal intersection of alysphenoid and maxillary (10), lateral point at parietal-frontal suture (11), front of the squamosal root of the zygomatic (12), tip of pterygoid process (13), upper and inferior point at the posterior margin of the hamular process squamosal (14, 15), anterior and posterior extremity of occipital (16, 17), point above occipital condyle (18), interparietal-occipital-squamosal interaction (19), front of interparietal (20), back of interparietal (21).

and tended to separate the two populations of *M. erythroleucus*. Deformations further suggested that *M. huberti* differed from other species by a shorter cranium, but a larger braincase on a level with tympanic bulla and zygomatic arches (Fig. 3). The second axis separated *M. erythroleucus* and *M. natalensis*. The visualizations of Procrustes superimpositions showed that *M. erythroleucus* is characterized by a proportionally



Fig. 2. Skull size variability among taxa. Boxplots of the sum of dorsal, ventral and lateral log centroid sizes. The analyses were done by pooling the sides. Boxplots present medians and 25 and 75 percentiles; limits are the 95 confidence intervals. ERY1: Mankountan; ERY2: Bantou.

Table 1Classification results of the discriminant analysis by species.

| Actual group | Ν  | Predicted group membership |          |          |
|--------------|----|----------------------------|----------|----------|
|              |    | HUB                        | ERY      | NAT      |
| HUB          | 43 | 36 (83.7%)                 | 3 (6.9%) | 4 (9.3%) |
| ERY          | 60 | 1 (1.7%)                   | 57 (95%) | 2 (3.3%) |
| NAT          | 30 | 2 (6.7%)                   | 1 (3.3%) | 27 (90%) |

broader and higher cranium (Fig. 3) than *M. natalensis*. *M. natalensis* is characterized by an inter-orbital constriction (Fig. 3). Cross-validated classifications correctly assigned 90% of the 133 individuals analysed (Table 1) to the species level. Within *M. erythroleucus*, a cross-validated classification correctly assigned 68% of the 60 individuals.

According to the NJ tree based on Mahalanobis distances, *M. natalensis* and *M. huberti* seemed to be morphologically more similar to each other than they are to *M. erythroleucus* (Fig. 4).

# 4. Discussion

Geometric morphometrics analyses clearly demonstrated that the three species largely overlapped in centroid size but exhibited large differences in skull shape. The results of skull shape analyses showed also that *M. natalensis* and *M. huberti* fit the classic definition of sibling species because they seem to be morphologically more similar to each other than they are to *M. erythroleucus*. According to Duplantier [20] has a significantly smaller mandible than the two other species while the dental rows of *M. erythroleucus* were higher than either M. natalensis or M. huberti. Three other works have also been more or less successful in attempting to discriminate between sympatric species of Mastomys. First, Dippenaar et al. [39] in South Africa found similar arithmetic means of skull measurements between M. natalensis and M. coucha. In the Ethiopian rift valley, Lavrenchenko et al. [40] studied three sympatric species of Mastomys using classical PCA methods on skull distances. The authors showed that M. natalensis was separated from a group made by M. erythroleucus and M. awashensis representatives. However their study was limited by the number of specimens used (only 4 M. natalensis). Finally, Bronner et al. [41] showed that the inclusion of subadult specimens reduced a posteriori classification accuracy below 95% confidence levels between M. natalensis and M. coucha in South Africa, implying that the age-related variation is sufficiently pronounced to obscure interspecific craniometric differences.

Our results suggest that the three species can be differentiated owing to their skull shapes with a high probability (90%). However, we did not analyze the overall intra-specific variability, as only one site of trapping was used for *M. huberti* and *M. natalensis*. Indeed, our results within *M. erythroleucus* samples suggest potential high intra-species variability.

The main shape changes between species concerned the height of braincase, the length of the rostrum and the larger size of the zygomatic bar, possibly implying a modification in the attachment of masseter muscles. These anatomical elements are important for the biomechanics of mastication [42,43]. It has been shown that diet is an important selective factor acting upon the evolution of skull morphology [35,44,45]. Hence, a source of variability could result from competition for food. This competition could act as a prime selective pressure among sympatric species and could possibly allow for character displacement between and within Mastomys species. Very few data concerning Mastomys feeding behaviour are available. In general, the three species are characterised by their opportunism from their omnivorous-granivorous feeding habits [46]. The feeding habits of the three species are very similar but they distinguish themselves in the details, each species consuming more particularly the food available in its micro-habitat [47]. In Mankountan, M. huberti is primarily observed in swamps (95.1%); M. natalensis occupies houses in Bantou (64.1%); M. erythroleucus is primarily found in houses at Mankountan (84.3%) and in fields at Bantou (89.6%). Smithers [47] observed in the field that *M. natalensis* primarily eats grass



Fig. 3. Shape variability among taxa: first two axes of the CVA, computed for the three pooled sides of skull (for 21 PCs included). Light grey: *M. huberti*; dark grey: *M. erythroleucus*; black: *M. natalensis*. Shape differences along the CVA axes one and two. Solid lines depict shapes on the positive side of the CVA axis, dotted lines on the negative one.

and grains. Kingdon [48] showed that this species can feed itself with animals (insects) or vegetation. Jackson and Van Aarde [49] showed evidence in which M. erythroleucus was able to adapt itself to the less demanding feeding habits of the two other cogeneric species. M. erythroleucus would then present an adaptive capacity higher than the two other species. M. erythroleucus shows a broader and higher skull shape with a length and strength greater than that of M. huberti or *M. natalensis*. It is possible that this more resistant and adaptive feeding habit would require more from the chewing muscles, leading to a more developed insertion zone. Furthermore, the competition among available resources could explain the high variability in skull size and shape for M. erythroleucus in Bantou. Similar observations have already been made for populations of the European wood mice Apodemus sylvaticus and flavicollis [50], in Murinae [51], and in Tanzanian M. natalensis populations [52]. Furthermore, it is interesting to note that a study has compared several samples of *M. natalensis* issued from all Africa and has showed that in Tanzania, when not in sympatry, the *M. natalensis* are larger (Denys, pers. com.). These results clearly confirm that there is a high intraspecific variability at a local scale in *Mastomys*.

In conclusion, the present geometric morphometrics study based on skull shape allowed us to partially discriminate three morphological groups that are congruent with the three species suggested by molecular identification. In *Mastomys*, the evolution of cranial length and shape could be interpreted as a link to the competitive pressure between close species by ecological segregation. We confirm that selection may play a crucial role in the evolution of skull shape. Future studies on *Mastomys* species should be focused on ecological partitioning of the habitats and diets in the three sibling species of Guinean *Mastomys*. The cranial characteristics using the statistical formalism of geometric



Fig. 4. Overall shape differences among taxa: Neighbour-joining tree of the Mahalanobis  $D^2$  distance.

morphometrics are sufficient to be a valid identification criterion in such a case of sibling species, a condition previously reached by Dobigny et al. [19] and Cordeiro et al. [21]. It is only a combined set of characters (external, skull, dental morphology) that allows determination of the species with 100% confidence. The integration of all these techniques (molecular, cladistic, phylogenetic and cytogenetic) together provides a powerful tool in allowing identification of sibling species. In the future, the Barcode of life initiative may also represent a good alternative to solving the problems of sibling species identification [53,54].

#### Acknowledgements

We would like to thank the field staff: Project PFHG Guinea. We are grateful to C. Houssin, A. Delapré, F. Kourouma and P. Guilavogui for the preparation of specimens. We thank M. Baylac for the use of morphometric platform. This work was supported by the European Community (INCO-DEV grant ICA4-CT2002-10050).

#### References

A.J. Cain, Animal Species and Their Evolution, Hutchinson University Library, London/New York, 190 p.

- [2] T.R.O. De Freitas, Tuco-tucos (Rodentia, Octodontidae) in Southern Brazil: *Ctenomys lami* spec. nov. Separated from *C. minutus* Nehring 1887, Studies on Neotropical Fauna and Environment 36 (1) (2001) 1–8.
- [3] E. Van der Straeten, E. Lecompte, C. Denys, *Praomys petteri* : une nouvelle espèce de Muridae africains (Mammalia, Rodentia), Bonner Zoologische Beitrage 4 (2003) 329–345.
- [4] L.P. Costa, S.E. Pavan, Y.L.R. Leite, V. Fagundes, A new species of *Juliomys* (Mammalia: Rodentia: Cricrtidae) from the Atlantic forest of southeastern Brazil, Zootaxa 1463 (2007) 21–37.
- [5] G. Dobigny, M. Baylac, C. Denys, Geometric morphometrics, neural networks and diagnosis of sibling *Taterillus* (Rodentia, Gerbillinae) species, Bio. J. Linn. Soc. 77 (2002) 319–327.
- [6] J. Hausser, D. Jammot, Etude biométrique des mâchoires chez les Sorex du groupe *araneus* en Europe continentale (Mammalia, Insectivora), Mammalia 38 (1974) 324–343.
- [7] P.J. Taylor, I.L. Rautenbach, D. Gordon, K. Sink, P. Lotter, Diagnostic morphometrics and southern African distribution of two sibling species of the rat, *Thallomys paedulcus* and *Thallomys nigricauda* (Rodentia: Muridae), Durban. Mus. Novit. 20 (1995) 49–62.
- [8] E. Capanna, A. Bekele, M. Capula, R. Castiglia, M.V. Civitelli, J.T. Codjia, M. Corti, C. Fadda, A multidisciplinary approach to the systematics if the genus *Arvicanthis* Lesson, 1842 (Rodentia, Murinae), Mammalia 60 (4) (1996) 677–696.
- [9] E. Van der Straeten, C.B. Robbins, Further studies on *Masto-mys* (Rodentia: Muridae) types and generic distinction among African Muridae, Mitt. Zool. Mus. Berl. 73 (1) (1997) 153–163.
- [10] P. Taylor, Patterns of chromosomal variation in southern African rodents, J. Mammalogy 81 (2) (2000) 317–331.
- [11] L. Granjon, J.M. Duplantier, J. Catalan, J. Britton-Davidian, G.N. Bronner, Conspecificity of *Mastomys natalensis* (Roden-

tia: Muridae) from Senegal and South Africa: evidence from experimental crosses, karyology and biometry, Mammalia 60 (4) (1996) 697–706.

- [12] L. Granjon, J.M. Duplantier, J. Catalan, J. Britton-Davidian, Systematics of the genus *Mastomys* (Thomas, 1915) (Rodentia: Muridae). A review, Belg. J. Zool. 127 (1) (1997) 7–18.
- [13] G.G. Musser, M.D. Carleton, Superfamily Muroidea, in: D.E. Wilson, D.M. Reeder (Eds.), Mammal Species of the World. A Taxonomic and Geographic Reference, 3rd edition, The John Hopkins University, Pr Baltimore, 2005, pp. 894–1531.
- [14] E. Lecompte, K. Aplin, C. Denys, F. Catzeflis, M. Chades, P. Chevret, Phylogeny and biogeography of African Murinae based on mitochondrial and nuclear gene sequences with a new tribal classification of the subfamily, BMC Evolutionnary Biology 8 (2008) 199–220.
- [15] M.I. Baskevitch, L.A. Lavrenchenko, On the morphology of spermatozoa in some African Murinae (Rodentia, Muridae): the taxonomic and phylogenetic aspects, J. Zool. Syst. Evol. Res. 33 (1995) 9–16.
- [16] L.A. Lavrenchenko, M.I. Baskevitch, Variation of generative system structure in some species of the genus *Mastomys* Thomas, 1915 (Rodentia, Muridae), Mammalia 60 (1996) 277–288.
- [17] J. Britton-Davidian, J. Catalan, L. Granjon, J.M. Duplantier, Chromosomal phylogeny and evolution in the genus *Mastomys* (Mammalia, Rodentia), J. Mammalogy 76 (1) (1995) 248–262.
- [18] P. Chevret, L. Granjon, J.M. Duplantier, C. Denys, F.M. Catzeflis, Molecular phylogeny of the *Praomys* complex (Rodent, Murinae): a study based on DNA/DNA hybridization experiments, Zool. J. Linn. Soc. 112 (1994) 425–442.
- [19] E. Lecompte, L. Granjon, J.C. Kerbis Peterhans, C. Denys, Cytochrome b-based phylogeny of the *Praomys* group (Rodentia, Murinae): a new African radiation? C. R. Biologies 325 (2002) 827–840.
- [20] J.M. Duplantier, Biologie évolutive de populations du genre Mastomys (Rongeur, Muridés) au Sénégal, Thèse Doc. Etat, Univ. Montpellier II, pp. 1–215.
- [21] P. Cordeiro-Estrela, M. Baylac, C. Denys, J. Polop, Combining geometric morphometrics and pattern recognition to identify interspecific patterns of skull variation: case study in sympatric Argentinian species of the genus *Calomys* (Rodentia: Cricetidae: Sigmodontinae), Bio. J. Linn. Soc. 94 (2) (2008) 365–378.
- [22] A. Evin, M. Baylac, M. Ruedi, J.M. Pons, Taxonomy, skull diversity and evolution in a species complex of *Myotis* (Chiroptera: Vespertilionidae): a Geometric morphometric appraisal, Bio. J. Linn. Soc. 95 (3) (2008) 529–538.
- [23] A.S. Loy, S. Di Martino, D. Capolongo, Patterns of geographic variation of *Talpa romana* Thomas (Insectivora: Talpidae). Preliminary results derived from a geometric morphometrics approach, Mammalia 60 (1996) 77–89.
- [24] L.R. Monteiro, Multivariate regression models and geometric morphometrics: the search for casual factors in the analysis of shape, Syst. Biol. 84 (1999) 192–199.
- [25] P. Cordeiro-Estrela, M. Baylac, C. Denys, J. Marinho-Filho, Interspecific patterns of skull variation between sympatric Brazilian vesper mice: Geometric morphometrics assessment, J. Mammalogy 87 (6) (2006) 1270–1279.
- [26] W.L. Gannon, G.R. Racz, Character displacement and ecomorphological analysis of twolong-eared *Myotis* (*M. auriculus* and *M. evotis*), J. Mammalogy 87 (2006) 171–179.
- [27] D.C. Adams, F.J. Rohlf, Ecological character displacement in Plethodon: Biomechanical differences found from a geometric morphometric study, Proc. Nat. Acad. Sci. 97 (8) (2000) 4106– 4111.

- [28] J.S. McKinnon, S. Mori, B.K. Blackman, L. David, D.M. Kingsley, L. Jamieson, J. Chou, D. Schluter, Evidence for ecology's role in speciation, Nature 429 (2004) 294–298.
- [29] H. Leirs, Population Ecology of *Mastomys natalensis* (Smith, 1834). Implications for rodent control in Africa, PhD, University of Antwerp, Brussels, 1995.
- [30] L. Granjon, J.F. Cosson, E. Quesseveur, B. Sicard, Population dynamics of the multimammate rat *Mastomys huberti* in an annually flooded agricultural region of Central Mali, J. Mammalogy 86 (5) (2005) 997–1008.
- [31] A. Lalis, E. Lecompte, R. Cornette, S. Moulin, R.S. Machangu, R. Makundi, V. Aniskin, C. Denys, Polymorphism of the age population structure of two wild *Mastomys natalensis* (Rodentia: Muridae) Tanzanian habitat samples: a multicriteria comparison, Mammalia 70 (3–4) (2006) 293–299.
- [32] C.R. Goodall, Procrustes methods in the statistical analysis of shape revisited, in: K.V. Mardia, C.A. Gill (Eds.), Current Issues in Statistical Shape Analysis, Leeds University Press, 1995, pp. 18–33.
- [33] F.J. Rohlf, D. Slice, Extensions of the Procrustes method for the optimal superimposition of landmarks, Syst. Zool. 39 (1990) 40– 59.
- [34] F.L. Bookstein, Morphometric Tools for Landmark Data: Geometry and Biology, Cambridge University Press, New York, 1991.
- [35] L.R. Monteiro, J.A.F. Diniz-Filho, S.F. DosReis, E. D'Araujo, Shape distances in general linear models: are they really at odds with the goals of morphometrics? A reply to Klingenberg, Evolution 57 (2003) 196–199.
- [36] B.D. Ripley, Pattern Recognition and Neural Networks, Cambridge University Press, Cambridge, 1996.
- [37] M. Baylac, M. Friess, Fourier descriptors, Procrustes superimposition and data dimensionality: an example of cranial shape analysis in modern human population, in: E. Dennis, D. Slice (Eds.), Modern Morphometrics in Physical Anthropology, Part 1, Theory and Methods, Kluwer Academic/Plenum Publishers, New York, 2005, pp. 142–165.
- [38] M. Baylac, Rmorph: a R geometric ad multivariate morphometrics library, 2008, baylac@mnhn.fr.
- [39] N.J. Dippenaar, P. Swanepoel, D.H. Gordon, Diagnostic morphometrics of two medically important southern African rodents, *Mastomys natalensis* and *M. coucha* (Rodentia, Muridae), S. Afr. J. Sci. 89 (1993) 300–303.
- [40] L.A. Lavrenchenko, O. Likhnova, M.I. Baskevich, A. Bekele, Systematics and distribution of *Mastomys* (Muridae, Rodentia) from Ethiopia, with the description of a new species, Z. Säugetierk. 63 (1998) 37–51.
- [41] G.N. Bronner, M. Van der Merwe, K. Njobe, Nongeographic cranial variation in two medically important rodents from south Africa, *Mastomys natalensis* and *Mastomys coucha*, J. Mammalogy 88 (5) (2007) 1179–1194.
- [42] W.R. Atchley, B.K. Hall, A model for development and evolution of complex morphological structures, Biol. Rev. 66 (1991) 101– 157.
- [43] K.K. Smith, The form of the feeding apparatus in terrestrial vertebrates: studies of adaptation and constraint, in: J. Hanken, B.K. Hall (Eds.), The Skull, University of Chicago Press, Chicago, 1993, pp. 150–196.
- [44] L.F. Aguirre, A. Herrel, R. van Damme, E. Matthysen, The implications of food hardness for diet in bats, Functional Ecol. 17 (2003) 201–212.
- [45] E.R. Dumont, A. Herrel, The effects of gape angle and bite point on bite force in bats, J. Exp. Bio. 206 (2003) 2117–2123.

- [46] B. Hubert, G. Couturier, A. Poulet, A. Adam, F. Adam, Les conséquences d'un supplément alimentaire sur la dynamique des populations de rongeurs au Sénégal. I. Cas de *Mastomys erythroleucus* en zone sahélo-soudanienne, Rev. Ecol. (Terre & Vie) 35 (1981) 73–95.
- [47] R.H.N. Smithers, The Mammals of Botswana, Trustees of the National Museum of Rhodesia, Salisbury, 1971.
- [48] J. Kingdon, East African Mammals, An Atlas of Evolution in Africa, London, 1974.
- [49] T.P. Jackson, R.J. Van Aarde, Sex- and species-specific growth patterns in cryptic African rodents, *Mastomys natalensis* and *Mastomys coucha*, J. Mammalogy 84 (3) (2003) 851–860.
- [50] C. Panzironi, G. Cerone, M. Cristaldi, G. Amori, A method for the morphometric identification of southern Italian populations of *Apodemus* (*Sylvaemus*), Hystix 5 (1–2) (1994) 1–16.

- [51] S. Renaud, J. Michaux, P. Mein, J.P. Aguilar, J.C. Auffray, Morphometric analysis of the evolutionary of the European Miocene murine rodents, Lethaia 32 (1999) 61–71.
- [52] A. Lalis, M. Baylac, J.F. Cosson, R.H. Makundi, R.S. Machang'u, C. Denys, Cranial morphometric and fine scale genetic variability of two adjacent *Mastomys natalensis* (Rodentia: Muridae) populations, Acta Theriol. (2008), in press.
- [53] S. Ball, K.F. Armstrong, DNA barcodes for insect pest identification: a test case with tussock moths (Lepidoptera: Lymantriidae), The ecology of forest insect invasions and advances in their management, Canadian Journal of Forest Research 36 (2) (2006) 337–350.
- [54] S. Ratnasingham, P.D.N. Hebert, Bold: the Barcode of Life Data system, Molecular Ecology Notes 7 (2007) 355–364.