



ELSEVIER

Contents lists available at ScienceDirect

Comptes Rendus Biologies

www.sciencedirect.com



Molecular biology and genetics/Biologie et génétique moléculaires

Integrating marker-assisted background analysis with foreground selection for pyramiding bacterial blight resistance genes into Basmati rice

Nikita Baliyan^{a,*}, Rekha Malik^a, Reema Rani^{a,b}, Kirti Mehta^a,
Urvashi Vashisth^a, Santosh Dhillon^a, Khazan Singh Boora^a^a CCS Haryana Agricultural University, Hisar 125004, Haryana, India^b ICAR-Directorate of Rapeseed-Mustard Research, Sewar, Bharatpur, Rajasthan 321303, India

ARTICLE INFO

Article history:

Received 22 August 2017

Accepted after revision 13 November 2017

Available online 17 December 2017

Keywords:

Basmati rice

Bacterial leaf blight

Gene pyramiding

Molecular breeding

Marker assisted selection

Xanthomonas oryzae pv. *oryzae*

ABSTRACT

Bacterial leaf blight (BB), caused by the bacterium *Xanthomonas oryzae* pv. *Oryzae* (*Xoo*), is the major constraint amongst rice diseases in India. CSR-30 is a very popular high-yielding, salt-tolerant Basmati variety widely grown in Haryana, India, but highly susceptible to BB. In the present study, we have successfully introgressed three BB resistance genes (*Xa21*, *xa13* and *xa5*) from BB-resistant donor variety IRBB-60 into the BB-susceptible Basmati variety CSR-30 through marker-assisted selection (MAS) exercised with stringent phenotypic selection without compromising the Basmati traits. Background analysis using 131 polymorphic SSR markers revealed that recurrent parent genome (RPG) recovery ranged up to 97.1% among 15 BC₃F₁ three-gene-pyramided genotypes. Based on agronomic evaluation, BB reaction, aroma, percentage recovery of RPG, and grain quality evaluation, four genotypes, viz., IC-R28, IC-R68, IC-R32, and IC-R42, were found promising and advanced to BC₃F₂ generation.

© 2017 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

1. Introduction

Rice is an important cereal crop in the world and provides more than 21% of the food for the world population and up to 76% of the caloric intake in Southeast Asia [1,2]. Basmati is one of the premium rice groups, cultivated in the northwestern plains of India and eastern Pakistan. There is a marked increase in demand for Basmati rice worldwide for its unique aroma, cooking, and eating qualities [3]. In India, more than 60% of the total Basmati rice is grown in Haryana, and thus the latter is the major Basmati rice cultivating state. India is the leading exporter of Basmati rice and exports US\$ 4.86 billion of Basmati rice

annually [4]. Aroma, non-stickiness of cooked rice and elongation upon cooking are some of the important traits of Basmati rice. Bradbury et al. [5] showed that a functional BADH2 enzyme inhibits the biosynthesis of 2-acetyl-1-pyrroline, which is a major component of aroma. A fully functional copy of the gene encoding BAD2 is present in non-fragrant varieties, whereas fragrant varieties possess a copy of the gene containing an eight-base pair deletion resulting in a frame shift mutation disabling the BADH2 enzyme. Studies by Kovach et al. [6] also confirmed that BADH2 is the major genetic determinant of fragrance in rice. Amylose content and alkali spread value (ASV) are the most important constitutional indices related to cooking and processing quality of rice [7,8].

BB disease is one of the most devastating diseases effecting Basmati rice acreage in India. BB is caused by the

* Corresponding author.

E-mail address: nikita.biotech@gmail.com (N. Baliyan).

Gram-negative proteobacterium, *Xanthomonas oryzae* pv. *Oryzae*, and causes yield losses ranging from 74% to 81% in severe conditions, depending on the stage of the crop, the cultivar's susceptibility, and the environmental conditions [9,10]. Effective control of disease is limited by the use of chemical control measures and the health issues posed by them [11,12]. The development of varieties carrying resistance genes has been considered to be the most effective way to control the BB disease [13–16]. To date, at least 38 BB resistance genes [27 dominant (*Xa1*, *Xa2*, *Xa3*, *Xa4*, *Xa7*, *Xa10*, *Xa11*, *X12*, *Xa14*, *Xa16*, *Xa17*, *Xa18*, *Xa21*, *Xa22* (t), *Xa23*, *Xa25* (t), *Xa26*, *Xa27*, *Xa29*, *Xa30*, *Xa30* (t), *Xa31* (t), *Xa32* (t), *Xa34*, *Xa35* (t), *Xa36* (t), *Xa38*), and 11 recessive (*xa5*, *xa5*(t), *xa8*, *xa13*, *xa15*, *xa19*, *xa20*, *xa24*, *xa28* (t), *xa31* and *xa32*)] conferring host resistance against various strains of *Xoo* have been identified [17,18].

Molecular breeding involving MAS addresses the limitations of conventional breeding and allows pyramiding of multiple resistance genes into a single cultivar, thus providing an effective solution to resistance breakdown. Using the gene pyramiding approach, improved rice cultivars with broad spectrum and durable BB resistance have been developed by combining multiple resistance genes [13–16,19–22]. A marker-assisted background selection approach can recover up to 99% of the recurrent-parent genome in just three backcross cycles, whereas conventional breeding takes up to six backcrosses to recover 99% of the recurrent parent genome [23]. BB resistance genes *xa13* and *Xa21* have been introgressed into 'Pusa Basmati 1' through MAS coupled with phenotypic selection for agronomic, grain, and cooking quality traits [22]. One of the improved genotypes was released as 'Improved Pusa Basmati 1' for commercial cultivation in 2007, and is one of the first products of MAS to be released in India [24]. In the present study, we employed MAS along with stringent phenotypic selection for the introgression of three BB resistance genes, viz., *Xa21*, *xa13*, and *xa5* into Basmati variety CSR-30. This work will facilitate future efforts to transfer combinations of BB resistance genes into other preferred Basmati rice cultivars from non-Basmati rice cultivars without compromising the Basmati quality characters.

2. Material and methods

2.1. Plant material

The plant material consisted of the Basmati rice (susceptible) variety CSR-30 as a recurrent parent, the

(resistant) IRBB-60 variety as a donor parent having BB resistance genes *xa5*, *xa4*, *xa13*, and *Xa21*. Crosses were made between CSR-30 and IRBB-60, and F₁ plants thus obtained were backcrossed with CSR-30. Among the BC₁F₁ population, the foreground selection was performed using polymerase chain reaction (PCR)-based sequence tagged sites (STS) markers linked to the three BB resistance genes *xa5*, *xa13*, and *Xa21*. The BC₁F₁ genotypes were advanced to BC₁F₂ and subjected to foreground selection followed by BB incidence analysis on artificial inoculation and background analysis. A similar strategy was used in the BC₂F₁ to obtain BC₃F₁ populations from which three-gene positive genotypes were selected based on molecular marker analysis and assessed for agronomic performance and disease response at the tillering stage. The BC₃F₁ plants having the three BB resistance genes were selfed and BC₃F₂ seeds thus obtained were used for grain quality evaluation.

2.2. DNA marker analysis and PCR amplification

Mini-scale genomic DNA isolation was carried out using the CTAB extraction method of Murray and Thompson [25] as modified by Saghai-Marooof et al. [26] and Xu et al. [27]. Three STS markers, viz., pTA248, RG136 and RG556 linked to the BB resistance genes, *Xa21*, *xa13* and *xa5*, respectively, were used to confirm the presence of these resistance genes at each backcross generation. The pTA248 marker is 0.2 cM from *Xa21* [28], the RG136 marker is 3.8 cM from *xa13* [29], and the RG556 marker is 1.7 cM from *xa5* [30] (Table 1). PCR was carried out in 20- μ l reaction mixtures containing 50 ng of genomic DNA, 2 units of Taq DNA polymerase, 1X PCR Buffer (10 mM Tris HCL, 1.5 mM MgCl₂), 100 μ M each of dNTPs, and 10 μ M of each primer. The template DNA was initially denatured at 94 °C for 5 min, followed by 30 cycles of PCR amplification with the following conditions: 30 s of denaturation at 94 °C, 1 min of annealing at 55 °C, and 1 min of primer extension at 72 °C followed by final extension at 72 °C for 10 min. The amplified product of pTA248 was electrophoretically resolved on 1.5% agarose gel containing 0.5 μ g/ml of ethidium bromide in a 1.0 X TBE buffer and visualized under UV light. For the amplified products of RG136 and RG556, 5 μ l of the PCR product were used for gel electrophoresis to check DNA amplification. The remaining PCR product was used for restriction digestion. The reaction mixture used for the digestion of the PCR product with the respective restriction enzyme consisted of 0.3 μ l (10 U/ μ l) of restriction enzyme *Hinf*1 for

Table 1
Molecular markers used for marker-assisted selection of BB resistance genes and aroma gene.

Molecular marker	R genes for BB/ <i>fgr</i> for aroma	Chromosome	primer sequences (5'–3')	Reference
pTA248 (0.2cM)	<i>Xa21</i>	5	F:5'AGACGCGGAAGGGTGGTTCCCGGA3' R:5'AGACCGGTAATCGAAAGATGAAAA3'	Yoshimura et al. [30]
RG136 (3.8cM)	<i>xa13</i>	8	F:5'TCCCAGAAAGCTACTACAGC3' R:5'GCAGACTCCAGTTTGACTTC3'	Zhang et al. [29]
RG556 (1.7cM)	<i>xa5</i>	11	F:5'TAGCTGCTGCCGTGCTGTC3' R:5'AATATTTTCAGTGTGCACTCTC3'	Ronald et al. [28]
BAD2	<i>fgr</i>	8	F:5'TTGTTGGAGCTTGCTGATG3' R:5'CTGGTAAAAAGATTATGGCTTCA3' R:5'CATAGGAGCAGCTGAAATAATAC3' R:5'AGTGCTTTACAAAGTCCCGC3'	Bradbury et al. [5]

RG136 amplicons and *Dra*I for RG556 amplicons, 2.5 μ l of 10 X restriction buffer, 7.2 μ l of sterile distilled water and 15 μ l PCR product to make a final volume of 25 μ l [31]. The reaction mixture was incubated for 5 h at 37 °C, and the products of restriction digestion were separated by gel electrophoresis (2.0% agarose) and visualized under UV light. MAS was also carried out for *fgr* gene for aroma located on the short arm of rice chromosome 8 using primer BAD2 [5] (Table 1).

For background analysis, 131 polymorphic SSR markers out of the 428 tested were used for the assessment of the relative contribution of the two parental genomes to the pyramided genotypes. The computer package NTSYS-PC [32] was used for cluster analysis. Graphical genotypes (GGT) Version 2.0 [33] software programme was used for the assessment of the genomic contribution of the parent in the selected genotypes based on SSR marker data.

2.3. BB inoculation and resistance evaluation

Plants selected based on molecular marker analysis from the BC₁F₂ and BC₃F₁ generation carrying resistance genes (*Xa21*, *xa13* and *xa5*) individually and in combinations, along with the control, were inoculated with the predominant *Xoo* isolate prevalent in Kaul, Haryana, using a bacterial suspension of 10⁹ cells/ml [34]. The plants were clip inoculated at the maximum tillering stage. Inoculation of the leaf blades was done by clipping with scissors at 3 cm below the leaf tips. On average, five leaves per plant were inoculated, and the disease incidence using the 0–5 scale was measured 14 days after inoculation.

2.4. Evaluation of quality traits

Plant height, effective tillers per plant, panicle length, filled grains per panicle, 1,000 grain weight and grain yield were recorded in the rice genotypes pyramided with three resistance genes in BC₁F₂ and BC₃F₁ generation. The

pyramided BC₁F₂ and BC₃F₁ genotypes were grown during the kharif season of 2012 and 2014 in a net house at CCSHAU, Hisar, and in field at the Regional Rice Research Station, Kaul. Harvested seeds from BC₃F₁ three-gene-pyramided genotypes were further analysed for physicochemical characteristics like kernel dimensions of uncooked and cooked rice, aroma, ASV and amylose content. For testing aroma, one gram milled rice kernels were soaked in 10 ml of 1.7% KOH at room temperature in covered Petri plates for 10 min [45]. The amylose content of the pyramided genotypes was estimated according to Perez and Juliano [35]. ASV was assayed according to the method of Little et al. [36].

3. Results

3.1. Foreground and background selection for BB resistance genes

In BC₁F₂ population, plants with single and with three resistance genes were identified. The presence of BB resistance genes, viz., *Xa21*, *xa13*, and *xa5* in the BC₁F₂ genotypes were determined by respective linked marker and 10/230 genotypes had all the three BB resistance genes. Among resistant genotypes with different gene combinations, 44 were homozygous for *Xa21* (i.e. *Xa21Xa21*), six genotypes were homozygous for *xa13* (i.e. *xa13xa13*), nine genotypes were homozygous for *xa5* (i.e. *xa5xa5*) and the remaining plants were heterozygous, either for *Xa21* or *xa13* or *xa5*. A total of 104 polymorphic SSR markers spanning uniformly across all the rice chromosomes were used for background selection among the ten three-gene-pyramided genotypes. RPG recovery ranged from 44.1% to 78.9%. Three out of ten plants that showed RPG recovery above 70% were backcrossed with CSR-30, and BC₂F₁ generation was obtained, which was further advanced to BC₃F₁. Of the 112 BC₃F₁ plants, 15 genotypes were positive for all the three BB resistance genes using the linked markers (Fig. 1). In BC₃F₁ genera-

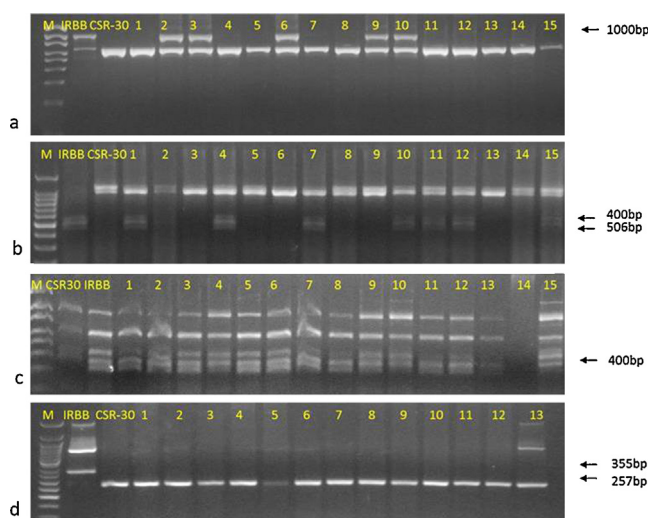


Fig. 1. Marker-assisted selection in BC₃F₁ genotypes from cross CSR-30 × IRBB-60 for (a) *Xa21* using pTA248 (b) *xa13* using RG136 (c) *xa5* using RG556, and (d) *fgr* using BAD2 markers.

tion, background selection was performed using a total of 131 polymorphic SSR markers. The similarity indices between different genotypes ranged from 0.02 to 0.97 across all the genotypes. The donor parent (IRBB-60) and recurrent parent (CSR-30) had low genetic similarity coefficient and bifurcated at a coefficient value of 0.09. Basmati and non-Basmati genotypes are highly differentiated genetically, as evident from the cluster pattern. The maximum genetic similarity coefficient for the three-gene-pyramided BC₃F₁ rice genotypes with respect to recurrent parent CSR-30 was observed in IC-R28 (0.97) followed by IC-R68 (0.94), IC-R32 (0.93) and IC-R42 (0.93). UPGMA cluster tree was divided into two groups (Fig. 2). The recipient parent CSR-30 and all the BC₃F₁ rice plants positive for *Xa21*, *xa13* and *xa5* BB resistance genes fell in one group with two major sub-groups. Sub-group I consisted of recipient parent CSR-30 and two genotypes, IC-R28 and IC-R32 with maximum similarity to CSR-30. Sub-group II consisted of IC-R42 and IC-R68 with close similarity to CSR-30 and the remaining eleven genotypes. In BC₃F₁, RPG recovery ranged from 72.9% to 97.1%. On BB resistance genes carrier chromosomes, 6 polymorphic markers were found on chromosome 5, 8 on chromosome 8 and 5 on chromosome 11, respectively. Maximum donor parent genomic regions among the three carrier chromosomes were present on chromosome 8 and chromosome 11 flanking the genes *xa13* and *Xa21*, as a result of linkage drag (Fig. 3). Background analysis conferred that IC-R28, IC-R68, IC-R32 and IC-R42 had the least introgression of donor segments. These four three-gene-pyramided genotypes with RPG above 89% were selfed and BC₃F₂ seeds were harvested and analysed for grain and cooking quality traits. Screening for BB resistance, and agronomic performance resulted in a cost-effective selection process due to a reduction in the number of plants subjected to foreground

selection. The molecular analysis with BAD2 primer combination specific for aroma (*fgr*) produced a 257 bp fragrance specific allele in all 15 recombinants and CSR-30, whereas IRBB-60 produced a 355 bp non-fragrance allele (Fig. 1d).

3.2. Evaluation of BB resistance

The BC₃F₁ pyramided genotypes were evaluated for their resistance to BB under glass house conditions using the *Xanthomonas oryzae* strain isolated from the BB infected fields of RRS, Kaul, CCSHAU, Hisar. The three-gene-pyramided BC₃F₁ genotypes with mean lesion length of 0.4 cm, were found to be equally effective against the virulent *Xoo* strain as the donor parent IRBB-60 with a mean lesion length of 0.5 cm. Also, the genotypes having either *Xa21* (mean lesion length of 1.2 cm) or *xa5* (mean lesion length of 1.1 cm) resistance genes alone were found to be resistant or moderately resistant to the BB disease. However, genotypes with *xa13* gene (mean lesion length of 4.8 cm) alone were found to be susceptible to BB disease. The pyramided genotypes with three genes had a higher level of resistance and a broader resistance spectrum than parental genotypes or genotypes with a single gene. The results indicated that the genes in combinations were more effective against the pathogen than a single gene (Fig. 4).

3.3. Agronomic performance and quality traits

Significant variation was observed among the pyramided lines and parental rice genotypes for plant height, tillers per plant, panicle length, filled grains/panicle, and grain yield per plant (Table 2). Most of the three gene-pyramided BC₃F₁ genotypes were similar or superior to the recurrent parent CSR-30 for the agronomic traits. In the pyramided BC₃F₁ genotypes, the plant height ranged from

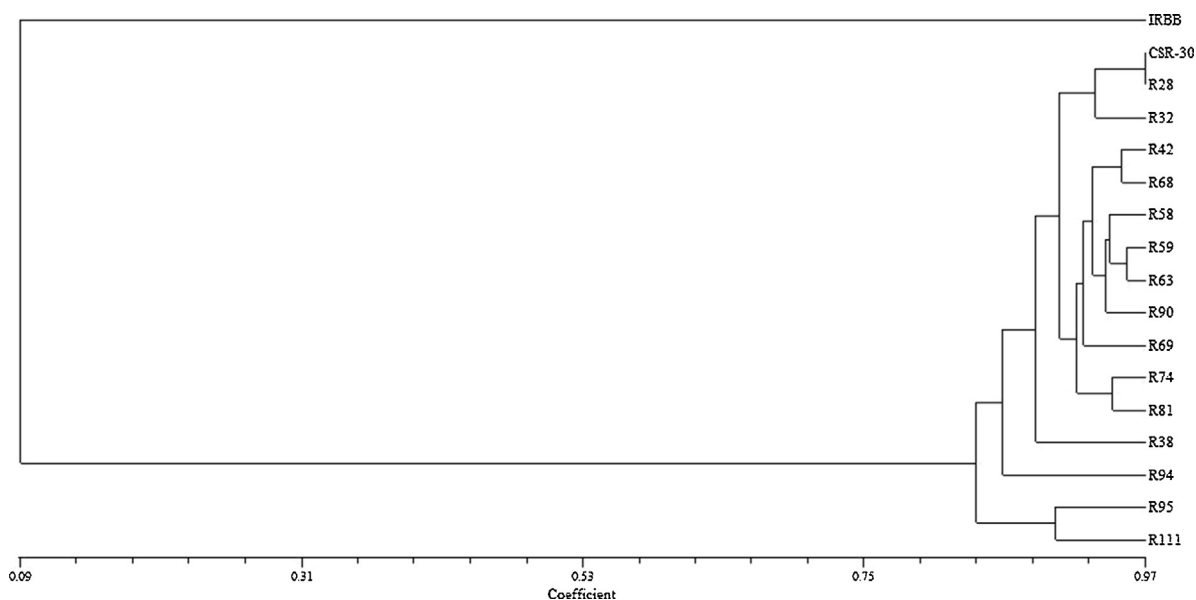


Fig. 2. Dendrogram (NTSYS-pc, UPGMA) showing the genetic similarity among the BC₃F₁ three-gene-pyramided genotypes and the parental rice genotypes based on SSR diversity data at 428 loci.

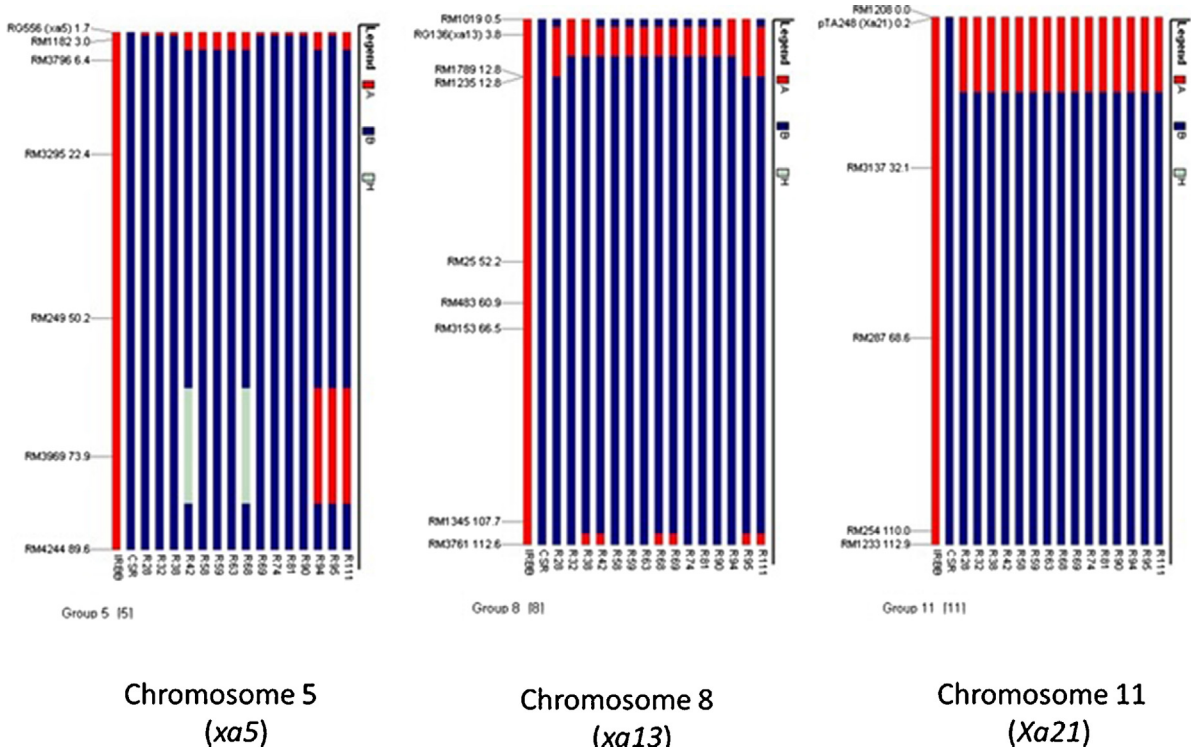


Fig. 3. Demonstration of the donor parent (IRBB-60) and recurrent parent (CSR-30) genome on chromosomes 5, 8 and 11 (having BLB resistance genes *xa5*, *xa13* and *Xa21*) in the 15 three-gene-pyramided BC₃F₁ genotypes.

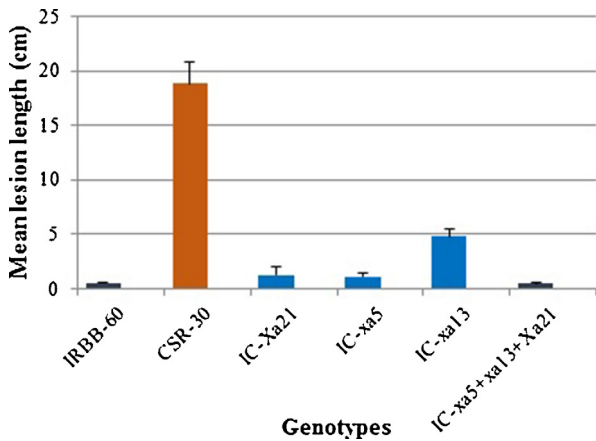


Fig. 4. Disease reaction of the donor parent (IRBB-60), the recurrent parent (CSR-30), and the pyramided BC₃F₁ genotypes (carrying one to three BB resistance genes).

130.5 to 154 cm, whereas the plant height of the recurrent parent CSR-30 was 164.5 cm. The three gene-pyramided BC₃F₁ genotypes had plant height intermediate between the those of the two parents. The fifteen pyramided BC₃F₁ genotypes had 1000 grain weight greater than the recurrent parent CSR-30 (21.2 g). The genotypes with maximum recovery of RPG, IC-R28 (31.11 g/plant), IC-R32 (33.22 g/plant), IC-R42 (36.78 g/plant) and IC-R68 (33.38 g/plant) showed grain yield/plant higher than that of the recurrent parent CSR-30 (31.08 g/plant).

A significant variation was observed among the pyramided genotypes and parental rice genotypes for grain quality traits (Table 3). Kernel length/breadth ratio after cooking among pyramided BC₃F₁ genotypes varied between 4.2 (IC-R94) to 5.8 (IC-R90), respectively. The elongation ratio for CSR-30 was recorded as 1.93. The elongation ratio among pyramided BC₃F₁ genotypes varied between 1.37 (IC-R38) to 1.64 (IC-R28), respectively. The BC₃F₁ three-gene-pyramided genotypes except IC-R59 and IC-R94 (ASV-2, high gelatinization, 75–70°C) showed an ASV of 4 (intermediate gelatinization, 70–74°C) or 6 (low gelatinization, 55–60°C). The percent amylose for the pyramided genotypes varied from 21.4% to 35.0%. The four three-gene-pyramided genotypes IC-R28, IC-R68, IC-R32 and IC-R42 were found to have amylose percentages of 24.9%, 24.3%, 25.3% and 22.2%, within an intermediate range similar to that of CSR-30 (23.3%).

4. Discussion

BB causes severe yield loss and is the major constraint for Basmati rice production. Several BB resistance genes have been identified and characterized in non-Basmati rice, and incorporated and pyramided through MAS into aromatic rice [14,22,31]. Often the transfer of resistance genes from non-Basmati rice to Basmati rice impairs its grain and cooking quality characteristics. In this study, we report the successful introgression of BB resistance genes into Basmati variety CSR-30 without yield penalty or impairment of its culinary qualities by adopting a

Table 2Agronomic traits, BB reaction and % recurrent parent genome (RPG) recovery in the three-gene-pyramided BC₃F₁ genotypes.

S. No.	Plant No.	PH (cm)	NT	1000 GW (g)	PL(cm)	FG/P	Awns	Y/P (g)	BB lesion length (cm)	RPG (%)
1	IRBB-60	109	18	22.7	26.60 ± 0.53	131.00 ± 1.96	Absent	39.66	0.5	0
2	CSR-30	164.5	16	21.2	31.12 ± 0.57	89.3 ± 1.20	Small	31.08	18.8	100
3	IC-R28^a	152	14	26.8	27.32 ± 0.34	95.25 ± 0.42	Small	31.11	0.34	97.1
4	IC-R32^a	152.5	9	25	22.62 ± 0.69	81.50 ± 0.73	Small	33.22	0.31	91.2
5	IC-R38	144	12	22.7	26.1 ± 0.58	103.75 ± 0.53	Small	29.12	0.20	89
6	IC-R42^a	151.5	11	25.5	26.05 ± 0.26	74.75 ± 0.70	Absent	36.78	0.42	89.2
7	IC-R58	152.5	9	26	29.87 ± 0.52	86.75 ± 0.82	Small	31.36	0.49	81.7
8	IC-R59	151	8	27.8	24.57 ± 0.42	78.87 ± 0.52	Small	32.89	0.25	85.8
9	IC-R63	153.5	10	26.6	26.42 ± 0.32	89.00 ± 1.28	Small	29.80	0.33	82.2
10	IC-R68^a	143	11	27.7	30.20 ± 0.40	91.25 ± 0.51	Small	33.38	0.30	94.3
11	IC-R69	141	10	22.9	25.37 ± 0.61	87.75 ± 0.85	Small	29.36	0.46	83
12	IC-R74	154	10	25.2	26.20 ± 0.48	105.3 ± 0.51	Small	33.63	0.29	87.4
13	IC-R81	151	9	25.7	27.30 ± 0.46	71.75 ± 0.73	Small	31.34	0.36	82.3
14	IC-R90	138	10	24.9	26.85 ± 0.37	34.00 ± 1.04	Small	34.15	0.36	78.2
15	IC-R94	130.5	9	29.8	24.97 ± 0.36	53.75 ± 0.70	Absent	34.21	0.31	81.8
16	IC-R95	149.5	10	24.4	32.02 ± 0.20	58.75 ± 1.03	Small	34.26	0.46	72.9
17	IC-R111	142	8	23.4	21.12 ± 0.24	69.00 ± 1.03	Absent	32.67	0.42	75.9

PH: plant height; NT: No. of tillers; 1000 GW: 1000 grain weight; PL: panicle length; Y/P: yield per plant; RPG(%): recurrent parent genome (%).

^a Indicates three-gene-pyramided genotypes with maximum RPG.**Table 3**Physical grain quality and cooking quality traits of parental and three-gene-pyramided BC₃F₁ genotypes.

S. No.	Plant No.	GL (mm)	GB (mm)	KLBC (mm)	KBBC (mm)	KLAC (mm)	KBAC (mm)	KER	ASV	AC (%)	AR
1	IRBB-60	9.72 ± 0.13	1.82 ± 0.05	7.80 ± 0.26	1.89 ± 0.07	10.80 ± 0.08	2.17 ± 0.02	1.38	6	28.1	NS
2	CSR-30	8.01 ± 0.09	1.80 ± 0.03	6.10 ± 0.21	1.60 ± 0.04	11.77 ± 0.06	2.20 ± 0.04	1.93	4	23.3	SS
3	IC-R-28^a	10.91 ± 0.37	1.90 ± 0.03	7.26 ± 0.07	1.66 ± 0.03	11.91 ± 0.16	2.37 ± 0.08	1.64	4	24.9	SS
4	IC-R-32^a	9.72 ± 0.09	1.87 ± 0.03	7.14 ± 0.12	1.58 ± 0.02	11.65 ± 0.11	2.23 ± 0.00	1.63	4	25.3	SS
5	IC-R-38	10.62 ± 0.14	1.79 ± 0.06	8.44 ± 0.07	1.67 ± 0.02	11.56 ± 0.03	2.12 ± 0.04	1.37	4	26.7	MS
6	IC-R-42^a	10.79 ± 0.21	1.85 ± 0.02	7.22 ± 0.15	1.66 ± 0.02	11.02 ± 0.17	2.00 ± 0.01	1.53	4	22.2	SS
7	IC-R-58	10.54 ± 0.15	1.87 ± 0.06	7.48 ± 0.21	1.63 ± 0.04	10.88 ± 0.06	2.08 ± 0.03	1.45	4	26.6	MS
8	IC-R-59	10.34 ± 0.15	1.90 ± 0.00	7.18 ± 0.09	1.64 ± 0.03	10.59 ± 0.09	1.96 ± 0.03	1.48	2	31.7	SS
9	IC-R-63	9.88 ± 0.22	1.86 ± 0.06	7.91 ± 0.16	1.65 ± 0.02	11.63 ± 0.14	2.55 ± 0.02	1.47	6	21.4	SS
10	IC-R-68^a	10.02 ± 0.09	1.97 ± 0.03	7.53 ± 0.05	1.67 ± 0.02	11.72 ± 0.21	2.68 ± 0.02	1.56	4	24.3	SS
11	IC-R-69	10.49 ± 0.12	1.76 ± 0.13	7.28 ± 0.05	1.64 ± 0.04	11.18 ± 0.23	2.29 ± 0.04	1.54	4	26.7	SS
12	IC-R-74	10.91 ± 0.02	1.66 ± 0.04	7.48 ± 0.03	1.62 ± 0.06	11.73 ± 0.06	2.45 ± 0.06	1.57	6	24.8	SS
13	IC-R-81	9.91 ± 0.21	1.90 ± 0.02	7.72 ± 0.09	1.65 ± 0.02	10.67 ± 0.07	1.88 ± 0.06	1.38	4	25.6	SS
14	IC-R-90	10.05 ± 0.15	1.92 ± 0.03	7.64 ± 0.17	1.64 ± 0.04	10.81 ± 0.11	1.87 ± 0.06	1.42	6	25.0	SS
15	IC-R-94	10.43 ± 0.07	1.96 ± 0.03	8.16 ± 0.12	1.75 ± 0.03	11.82 ± 0.31	2.79 ± 0.09	1.45	2	35.0	SS
16	IC-R-95	9.77 ± 0.04	1.85 ± 0.03	7.60 ± 0.14	1.62 ± 0.02	11.24 ± 0.04	2.01 ± 0.04	1.48	4	29.2	MS
17	IC-R-111	10.22 ± 0.15	1.85 ± 0.03	7.79 ± 0.03	1.67 ± 0.03	10.78 ± 0.08	2.00 ± 0.00	1.39	4	25.5	MS

GL: grain length; GB: grain breadth; KLBC: kernel length before cooking; KBBC: kernel breadth before cooking; KLAC: kernel length after cooking; KBAC: kernel breadth after cooking; KER: kernel elongation ratio; ASV: alkali spread value; AC(%): amylose content (%); AR: aroma; NS: non scented; SS: strong scented; MS: mild scented.

^a Indicates three-gene-pyramided genotypes with maximum RPG.

molecular breeding approach involving foreground selection along with phenotypic selection for grain and quality characters ([37], [16]).

The three-gene-pyramided BC₃F₁ genotypes derived in this study from the cross CSR-30 × IRBB-60, were as effective against the virulent *Xoo* strain as the donor line IRBB-60. The studies suggested that *xa5* was most effective, with shorter lesion lengths, followed by *Xa21*, while lines with *xa13* were susceptible to BB. The results indicated that the genes in combination were more effective against the pathogen than a single gene. Evidence suggested that the synergistic action or quantitative complementation between the resistance genes might result in increasing the level of resistance against the *Xoo* strain [20,24,30].

Molecular-marker-assisted background selection of recombinants was used to determine the parental genome

contribution. A total of 131/428 SSR markers produced polymorphism between the parental genotypes IRBB-60 and CSR-30, and were used to estimate the recovery of the recurrent parental genome in the pyramided BC₃F₁ genotypes. The percentage recovery of RPG in the pyramided BC₃F₁ genotypes ranged from 72.9% to 97.1%, as revealed by the global statistics using the software Graphical Geno Types (GGT) version 2.0. The low background recovery observed in some genotypes can be due to linkage drag. In the present study, 72.9–97.1% RPG recovery was achieved at the BC₃F₁ generation, compared to the theoretical expected value of approximately 93.0%. Combining the marker-based foreground selection with the phenotypic selection under the target environment has resulted in more than 90% recovery of the RPG in just three backcrosses, which is similar to what had

been observed in most of the studies, where both the foreground and background selections were performed using markers. Therefore, a phenotypic selection combined with marker-assisted background selection can make the breeding protocol economic and effective [24]. Chen et al. [38] reported 98.8% recovery after three backcrosses while selecting for recurrent-parent genome. Neeraja et al. [39] developed the submergence-tolerant rice cultivar Swarna and were able to recover 96.2% RPG in BC₃F₁ generation. Joseph et al. [22] suggested that the addition of one round of background selection in BC₁F₁ along with phenotypic selection may greatly increase the efficiency of the breeding program. Most of the three-gene-pyramided BC₃F₁ genotypes were similar or superior to the recurrent parent CSR-30 for the agronomic traits, grain, and cooking quality traits. The pyramided BC₃F₁ genotypes were shorter than the recurrent parent CSR-30 and had heavier grain weight, greater than that of the recurrent parent. The genotypes with maximum recovery of RPG, IC-R28, IC-R32, IC-R42, and IC-R68 showed grain yield higher than that of the recurrent parent CSR-30. The recombinants, IC-R42 and IC-R111 also showed the absence of awns as a desirable feature. Aroma, gelatinization temperature (GT) (scored by ASV), and amylose content are the most important characteristics with respect to rice cooking qualities. The three-gene-pyramided BC₃F₁ genotypes exhibited physical and cooking grain quality characteristics similar to those of CSR-30 (kernel length, kernel breadth, aroma, GT, amylose content). Aroma ranged from mild to strong scented for various recombinants, which may be due to temperature in the grain filling period [40]. All the pyramided genotypes except IC-R59 and IC-R94 (ASV-2, high gelatinization, 75–70°C) showed an ASV of 4 (intermediate gelatinization, 70–74°C) or 6 (low gelatinization, 55–60°C). The cooking time of the rice depends on the coarseness of the grain. The intermediate ASV indicated medium disintegration and is also classified as intermediate GT, which is highly desirable for grain quality [41]. The percent amylose for the pyramided genotypes varied from 21.4% to 35.0%. The four three-gene-pyramided genotypes IC-R28, IC-R68, IC-R32, and IC-R42 had amylose percentage similar to that of CSR-30. Rice with intermediate amylose content (20–25%) has been reported to cook moist and remain soft when cooled, and is widely preferred over rice with high (25–35%) or low amylose contents (10–20%) [42]. The amylose content of the rice influences the glycaemic index (GI) value: rice that is high in amylose has usually a lower GI value [43]. Rice with more amylose has reduced glycaemic responses and lower GI values than a rice variety with less amylose (1–20%) [44]. Thus, rice with intermediate amylose content (20–25%) should have low GI and may be good for consumption by diabetic people. Therefore, all the selected three-gene-pyramided BC₃F₁ genotypes were found to meet the Basmati grain quality standards.

5. Conclusion

CSR-30 possesses excellent grain, cooking, and eating quality features. Pyramided lines selected from cross CSR-30 IRBB-60 showed grain quality features similar to those

of the recurrent parent, CSR-30. In cross CSR-30 × IRBB-60, IRBB-60 as the donor parent for BB resistance facilitated the recovery of backcross-derived plants possessing the desirable grain quality features of CSR-30, despite the donor parent is not a Basmati. Based on agronomic evaluation, BB reaction, percentage recovery of recurrent parent genome, grain quality evaluation and aroma, four genotypes, viz., IC-R28, IC-R68, IC-R32 and IC-R42, were found promising.

Traditional Basmati varieties are highly susceptible to BB disease, for which many effective genes have been tagged and cloned. More resistance genes need to be identified and pyramided together into the elite cultivars to ensure the durability of BB resistance. Most of the major genes have been overcome by new or unrecognized pathogen races. Fortunately, this can be prevented by combining them with other resistance genes. In this study, genes conferring broad spectrum resistance were successfully introgressed through MAS. The pyramided genotypes performing better or equivalently to CSR-30 in grain yield and quality in addition to resistance against BB should be further evaluated at different locations. The pyramided genotypes obtained in our study can be used as genetic resources for BB resistance in breeding programs that will help to achieve better disease management. Marker-assisted selection thus can be successfully utilized to transfer combinations of BB resistance genes into other preferred rice cultivars without compromising yield and quality traits. Advanced basmati breeding lines, which will be derived through MAS and phenotypic selection, will, therefore, be of practical value in providing durable BB resistance in the Basmati growing region and are expected to have a high impact on the yield stability and sustainability of Basmati rice production.

References

- [1] M.A. Fitzgerald, Not just a grain of rice: the quest for quality, *Trends Plant Sci* 14 (2009) 133–139.
- [2] K. Miura, The role of QTLs in the breeding of high-yielding rice, *Trends Plant Sci.* 16 (2011) 319–326.
- [3] V.P. Singh, The Basmati rice of India, in: R.K. Singh, U.S. Singh, G.S. Khush (Eds.), *Aromatic rices*, Oxford & BH Publishing Co. Pvt. Ltd, New Delhi, India, 2000, pp. 136–153.
- [4] APEDA, 2014 <http://www.apeda.gov.in>.
- [5] L.M.T. Bradbury, R.J. Henry, Q. Jin, F. Reinke, P. Waters, A perfect marker for fragrance genotyping in rice, *Molec. Breed.* 16 (2005) 279–283.
- [6] M.J. Kovach, M.N. Calingacion, M.A. Fitzgerald, S.R. McCouch, The origin and evolution of fragrance in rice (*Oryza sativa* L.), *Proc. Nat. Acad. Sci. U S A* 106 (2009) 14444–14449.
- [7] I. Kumar, G.S. Khush, Inheritance of amylose content in rice (*Oryza sativa* L.), *Euphytica* 38 (1998) 261–269.
- [8] K. Denyer, P. Johnson, S. Zeeman, A.M. Smith, The control of amylose synthesis, *J. Plant Physiol.* 158 (2001) 479–487.
- [9] B. Srinivasan, S. Gnanamanickam, Identification of a new source of resistance in wild rice, *Oryzaerufipogon* to bacterial blight of rice caused by Indian strains of *Xanthomonas oryzae* pv. *Oryzae*, *Curr. Sci.* 88 (2005) 25.
- [10] T.H. Noh, D.K. Lee, J.C. Park, H.K. Shim, M.Y. Choi, M.H. Kang, J.D. Kim, Effect of bacterial leaf blight occurrence on rice yield and grain quality in different rice growth stage, *Plant Dis.* 13 (2007) 20–23.
- [11] S. Devadath, Chemical control of bacterial blight of rice, in: *Proceedings of the International Workshop on bacterial blight of rice*, International Rice Research Institute IRRI, Manila, Philippines, 1989, pp. 89–98.
- [12] P. Guillebeau, What to do about the food quality protection act. Or how can we protect the pesticides we need, *Proc. S.E. Pecan Growers Assoc.* 91 (1998) 65–69.
- [13] R.M. Sundaram, M.R. Vishnupriya, S.K. Biradar, G.S. Laha, G.A. Reddy, N.S. Rani, N.P. Sharma, R.V. Sonti, Marker assisted introgression of

- bacterial blight resistance in Samba Mahsuri, an elite indica rice variety, *Euphytica* 160 (2008) 411–422.
- [14] D. Rajpurohit, R. Kumar, M. Kumar, P. Paul, A.A. Awasthi, P.O. Basha, A. Puri, T. Jhang, K. Singh, H.S. Dhaliwal, Pyramiding of two bacterial blight resistance and a semi dwarfing gene in Type 3 Basmati using marker-assisted selection, *Euphytica* 178 (2010) 111–126.
- [15] P. Dokku, K.M. Das, G.J.N. Rao, Pyramiding of four resistance genes of bacterial blight in Tapaswini, an elite rice cultivar, through marker-assisted selection, *Euphytica* 192 (2013) 87–96.
- [16] J.P. Suh, J.U. Jeung, T.H. Noh, Y.C. Cho, S.H. Park, H.S. Park, M.S. Shin, C.K. Kim, K.K. Jena, Development of breeding lines with three pyramided resistance genes that confer broad-spectrum bacterial blight resistance and their molecular analysis in rice, *Rice* 6 (2013) 5.
- [17] H. Bhasin, D. Bhatia, S. Raghuvanshi, J.S. Lore, G.K. Sahi, B. Kaur, Y. Vikal, K. Singh, New PCR-based sequence-tagged site marker for bacterial blight resistance gene *Xa38* of rice, *Molec. Breed.* 30 (2012) 607–611.
- [18] P. Natraj Kumar, K. Sujatha, G.S. Laha, K. Srinivasarao, B. Mishra, B.C. Viraktamath, Y. Hari, C.S. Reddy, S.M. Balachandran, T. Ram, M. Sheshumadhav, N. Shobharani, C.N. Neeraja, G. Ashokreddy, H. Shaik, R.M. Sundaram, Identification and fine-mapping of *Xa33*, a novel gene for resistance to *Xanthomonas oryzae* pv. *oryzae*, *Phytopathology* 102 (2012) 222–228.
- [19] N. Huang, E.R. Angeles, J. Domingo, G. Magpantay, S. Singh, Q. Zhang, N. Kumar avadivel, J. Bennett, G.S. Khush, Pyramiding of bacterial resistance genes in rice: marker aided selection using RFLP and PCR, *Theor. Appl. Genet.* 95 (1997) 313–320.
- [20] A.C. Sanchez, D.S. Brar, N. Huang, Z. Li, G.S. Khush, Sequence tagged site marker-assisted selection for three bacterial blight resistance genes in rice, *Crop Sci.* 40 (2000) 792–797.
- [21] S. Singh, J.S. Sindhu, N. Huang, Y. Vikal, Z. Li, D.S. Brar, H.S. Dhaliwal, G.S. Khush, Pyramiding three bacterial blight resistance genes (*xa5*, *xa13* and *Xa21*) using marker-assisted selection into indica rice cultivar PR106, *Theor. Appl. Genet.* 102 (2001) 1011–1015.
- [22] M. Joseph, S. Gopalakrishnan, R.K. Sharma, V.P. Singh, A.K. Singh, N.K. Singh, T. Mohapatra, Combining bacterial blight resistance and basmati quality characteristics by phenotypic and molecular marker assisted selection in rice, *Molec. Breed.* 13 (2004) 377–387.
- [23] S.D. Tanksley, N.D. Young, A.H. Paterson, M.W. Bonierbale, RFLP mapping in plant breeding: new tools for an old science, *Bio-Technol.* 7 (1989) 257–264.
- [24] S. Gopalakrishnan, R.K. Sharma, K. Anand Rajkumar, M. Joseph, V.P. Singh, K.V. Bhat, N.K. Singh, T. Mohapatra, Integrating marker assisted background analysis with foreground selection for identification of superior bacterial blight resistant recombinants in Basmati rice, *Plant Breed.* 127 (2008) 131–139.
- [25] M.G. Murray, W.F. Thompson, Rapid isolation of high molecular weight plant, *Nucleic Acids Res.* 8 (1980) 4321–4325.
- [26] M.A. Saghai-Marouf, K.M. Soliman, R.A. Jorgensen, R.W. Allard, Ribosomal DNA spacer-length polymorphism in barley: mendelian inheritance, chromosomal-location and population dynamics, *Proc. Nat. Acad. Sci. U S A* 81 (1984) 8014–8018.
- [27] G.W. Xu, C.W. Magill, K.F. Schertz, G.E. Hart, An RFLP linkage map of Sorghum bicolor (L.) Moench, *Theor. Appl. Genet.* 89 (1994) 139–145.
- [28] P.C. Ronald, B. Albano, R. Tabien, L. Abenes, K.S. Wu, S.R. McCouch, S.D. Tanksley, Genetic and physical analysis of the rice bacterial blight disease resistance locus, *Xa21*, *Molec. Genome Genet.* 236 (1992) 113–120.
- [29] G. Zhang, E.R. Angeles, M.L.P. Abenes, G.S. Khush, N. Huang, RAPD and RFLP mapping of the bacterial blight resistance gene *xa13* in rice, *Theor. Appl. Genet.* 93 (1996) 65–70.
- [30] S.A. Yoshimura, N. Yishimura, S.R. Iwata, M.L. McCouch, M.R. Abenes, T.W. Baraoidian, R.J. Nelson, Tagging and combining bacterial blight resistance genes in rice using RAPD and RFLP markers, *Molec. Breed.* 1 (1995) 375–387.
- [31] S. Perumalsamy, M. Bharani, M. Sudha, P. Nagarajan, Functional marker-assisted selection for bacterial leaf blight resistance genes in rice (*Oryza sativa* L.), *Plant Breed.* 129 (2010) 400–406.
- [32] F.J. Rohlf, NTSYS-PC, in: numerical taxonomy and multivariate analysis system, Applied Biostatistical Inc, New York, 2000, [Version 2.02].
- [33] R. Van Berloo, GGT: software for display of graphical genotypes, *J. Hered.* 90 (1999) 328–329.
- [34] H.E. Kauffman, A.P.K. Reddy, S.P.Y. Hsien, S.D. Merca, An improved technique for evaluating resistance of rice varieties to *Xanthomonas oryzae*, *Plant Dis. Response* 57 (1973) 537–541.
- [35] C.M. Perez, B.O. Juliano, Modification of the simplified amylose test for milled rice, *Starch* 30 (1978) 424–426.
- [36] R.R. Little, G.B. Hilder, E.H. Dawson, Differential effect of dilute alkali on 25 varieties of milled white rice, *Cereal Chem.* 35 (1958) 111–126.
- [37] A.K. Singh, S. Gopalakrishnan, V.P. Singh, K.V. Prabhu, T. Mohapatra, N.K. Singh, T.R. Sharma, et al., Marker assisted selection: a paradigm shift in Basmati breeding, *Indian J. Genet. Plant Breed.* 71 (2011) 1–9.
- [38] S.C. Chen, G. Xu, H. Lin, Q. Zhang, Improving bacterial blight resistance of 6078, an elite restorer line of hybrid rice, by molecular marker-aided selection, *Plant Breed.* 120 (2001) 133–137.
- [39] C.N. Neeraja, R. Maghirang-Rodriguez, A. Pamplona, S. Heuer, B.C. Colard, E.M. Septingsih, G. Vergara, D. Sanchez, K. Xu, A.M. Ismail, D.J. Mackill, A marker-assisted backcross approach for developing submergence-tolerant rice cultivars, *Theor. Appl. Genet.* 11 (2007) 5767–5776.
- [40] T. Itani, M. Tamaki, Y. Hayata, T. Fushimi, K. Hashizume, Variation of 2-acetyl-1-pyrroline concentration in aromatic rice grains collected in the same region in Japan and factors affecting its concentration, *Plant Prod. Sci.* 7 (2004) 178–183.
- [41] U.K. Bansal, H. Kaur, R.G. Saini, Donors for quality characteristics in aromatic rice, *Oryza* 43 (3) (2006) 197–202.
- [42] IRRI, Rice quality training manual, 1985 [International Rice Research Institute].
- [43] J. Brand-Miller, E. Pang, L. Bramall, Rice: a high or low glycemic index food, *Am. J. Clin. Nutr.* 56 (1992) 1034–1036.
- [44] S. Nik-Shanita, H. Hasnah, C.W. Khoo, Amylose and amylopectin in selected Malaysian foods and its relationship to glycemic index, *Sains Malaysiana* 40 (2011) 865–870.
- [45] B.C. Sood, E.A. Siddiq, F.U. Zaman, Genetic analysis of kernel elongation in rice, *Indian J. Genet.* 43 (1983) 40–43.