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Infestation by *Ergasilus coatiarus* (Copepoda: Ergasilidae) in two Amazonian cichlids with new host record from Peru: An ectoparasites natural control approach



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ABSTRACT

The occurrence of copepods ergasilid was investigated in two species of cichlids of economic importance for aquaculture in the Amazon region: *Cichla monoculus* and *Chaetobranchus semifasciatus*. The fish were collected from a semi-intensive fish farm, near the city of Nauta, Loreto State, Peru. Copepods were found in the gill filaments of 44 of 85 specimens (51.7%) of *C. monoculus* and in eight of 30 (26.6%) specimens of *C. semifasciatus*. The parasite was identified as *Ergasilus coatiarus* based on its morphological features. The occurrence did not vary significantly with host size ($P \ge 0.05$) in both species. This is the first report of *E. coatiarus* parasitizing *C. semifasciatus* in the Amazon basin and the first report in *C. monoculus* from Peru. The high occurrence of these copepods in the present study points out the need of improving the strategies of parasitic prevention and control in order to better prevent future disease outbreaks.

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1. Introduction

In the last years, interest in parasitological studies on wild fish populations is increasing with the growing importance of fish cultures as a source of protein to human nutrition [1,2]. In the Amazon region of Peru, fish farming represents an activity that is developing rapidly and great interest has focused on several members of the family Cichlidae Heckel, 1840. The interest is attributable primarily to the economic importance of these species, its adaptability to diverse culture systems, good resistance to diseases, excellent feed conversion ratio, and high

* Corresponding author. *E-mail address:* patrickmathews83@gmail.com (P.D. Mathews). reproductive capacity [3]. Indeed, cichlids are commercially important for both wild fisheries and aquaculture in Amazonian countries, with the *Cichla monoculus* Agassiz, 1831 and *Chaetobranchus semifasciatus* Steindachner, 1875, being a significant example.

Copepods have a wide distribution around the World playing diverse roles in the aquatic environment and infecting farmed and wild fishes of commercial importance [1–6]. In aquaculture, some are beneficial and others are extremely adverse, which may result in entire production losses [1]. Among the species of copepods parasites of fish, Ergasilidae family includes species with high pathogenic potential, some of them responsible for great mortality among cultured fishes in freshwater and brackish environments [1,7]. However, little is known about of the ergasilids which infect cichlids in culturing

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systems and beyond in natural environment from Peruvian Amazon. In this sense, the present study aimed to evaluate the occurrence of copepods ergasilid in specimens of *C. monoculus and C. semifasciatus*, two South American freshwater fish of considerable economic importance.

2. Materials and methods

Eighty-five specimens of C. monoculus (8.4 to 22.3 cm in length) and thirty specimens of C. semifasciatus (6.5 to 12.3 cm in length) were collected from of a semi-intensive fish farm, near the city of Nauta (4°30'30"S, 73°35'00"W), Loreto State, Peru. The fish were monitored for three months (June to August 2016), during the dry season in the Amazon region. Immediately after collection, the fish were placed in plastic bags containing water, under conditions of artificial aeration, and transported alive to the field laboratory, where they were measured and euthanized by neural pithing, this method was approved by Ethics Committee of the Federal University of São Paulo-UNIFESP (CEUA No. 9209080214), in accordance with Brazilian law (Federal law No. 11.794, dated 8 October 2008 and Federal Decree No. 6899, dated 15 July 2009). All organs were examined using a stereoscopic microscope for parasites infestation. The parasites were removed with dissecting needles from the gill filaments and fixed in 70% ethanol. Small samples of gill tissue infected with adult copepods were removed and examined by means of differential interference contrast (DIC) microscopy at Department of Biophysics, Federal University of São Paulo.

The identification of the parasites was based on the methodology of Araujo and Varella [8] and Thatcher [2]. Holotype and paratypes from Collection of Invertebrates of Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil (INPA-CR No. 529, female; INPA-CR No. 530a-e, 5 females) were employed in order to support identification. The prevalence and mean intensity of the parasites were calculated according to Bush et al. [9]. The effect of host size on occurrence of copepods was analyzed by Chi² test. All results were considered significant for P < 0.05.

The physicochemical parameters of the water were measured two times daily (at 8 am and 4 pm) with daily checks of dissolved oxygen, pH, temperature and conductivity by means of a YSI multiparameter meter (Model MPS 556). Ammonium values, hardness, carbon dioxide and total alkalinity were monitored weekly and in the morning (8 am), using a complete package for analysis of freshwater (LaMotte AQ-2).

3. Results

In the present study, 44 (51.7%) *C. monoculus* and eight (26.6%) *C. semifasciatus* had adult parasites of ergasilid in their gill filaments. The mean intensity was three and two adult copepods per fish to *C. monoculus* and *C. semifasciatus*, respectively. These were not found in any other organs and no clinical signs were observed in the parasitized organ.

Based on the morphology of adult parasites observed via light microscopy, showing body with sub-triangular cephalon, smooth carapace, abdomen with three segments, uropod with two elongated caudal filaments, antennule with six articles, one segment in the exopod of the fourth leg and one serrate spine at the union of the third and fourth segments of the antenna (Fig. 1A-E), the analysis evidenced that these parasites belong to the genus *Ergasilus* Nordmann, 1832 and the same were identified as *Ergasilus coatiarus* Araujo and Varella, 1998. From the examined samples, it was evidenced that the host size of the animals did not affect the occurrence of *E. coatiarus* ($\chi^2 = 0.59$, df = 2, P = 0.744). However, to *C. monoculus* higher occurrence was found in fishes from 18.1 to 22.3 cm long.

The values of physicochemical parameters of water in culture pond were: dissolved the oxygen $(5.64 \pm 0.4 \text{ mgL}^{-1}),$ pН $(4.83 \pm 0.10),$ temperature $(27.23 \pm 0.50 \text{ °C})$ and conductivity $(106.1 \pm 14.0 \,\mu\text{Scm}^{-1})$. $(0.02 \pm 0.10 \text{ mgL}^{-1})$, Ammonium values hardness $(21.40 \pm 1.80 \text{ mgL}^{-1})$, carbon dioxide $(3.2 \pm 0.9 \text{ mgL}^{-1})$, and total alkalinity ($16.14 \pm 0.80 \text{ mgL}^{-1}$).

4. Discussion

Previous studies in Peru have shown widespread distribution of ectoparasites in many wild and cultivated fishes [14], bearing high economic importance in the Amazon region [7,10–13,15]. However, to our knowledge, this study reports for the first time the presence of *E. coatiarus* in *C. semifasciatus* in the Amazon basin and is besides the first report of *C. monoculus* from Peru. The information on morphological data in the present study corroborates the characteristics defined by Araujo and Varella [8] for this species. In the same manner, the site of infection where these parasites were found is in accordance with Araujo and Varella [8], Araujo et al. [16] and Thatcher [2], who reported that specimens of *E. coatiarus* preferably infect gills of their hosts.

The high occurrence of adult copepods of *E. coatiarus* in C. monoculus indicates that the infestation by this pathogen is probably frequent during the dry season in the earth ponds. Surely, seasonal occurrence survey is necessary in order to establish the pattern of infection for this parasite, taking in account that seasonal changes represent a combination of many factors influencing the success of a parasite to penetrate a host [17]. In the same way, according Tavares et al. [18], seasonal variation in rainfall levels is a major environmental factor affecting the dynamics of parasite communities in the Amazon region. Nevertheless, there are yet limited number of studies about the effects of the season in infestation patterns of copepods parasite in fish species in the Amazon region, making it difficult to confirm the influence of seasonality on prevalence of these ectoparasites. In this context, future studies to determine the infestation patters by parasites in several wild and cultured fish from Amazon region would be necessary to establish prophylactic strategies to decrease the infestation by ectoparasites, especially when present in farmed fish.

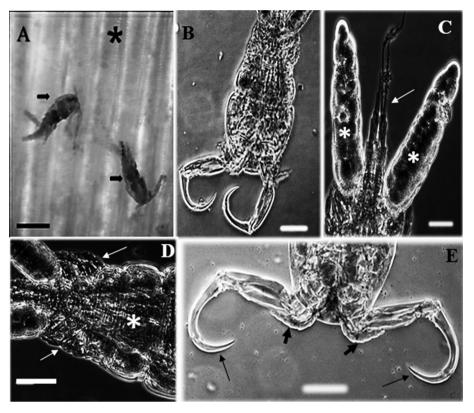


Fig. 1. (A-E) Light photomicrographs of *Ergasilus coatiarus* parasitizing gill filaments of the fish *Cichla monoculus* and *Chaetobranchus semifasciatus*. (A) Gills (asterisk) showing adult female of *E. coatiarus* (arrows). Scale bar = 20 μ m. (B) Ventral view of *E. coatiarus* total. Scale bar = 10 μ m. (C) Egg sac (white asterisks) and caudal rami (thin arrow). Scale bar = 10 μ m. (D) Ventral view of urosome (asterisk) showing the legs (thin arrows). Scale bar = 10 μ m. (E) Ventral view of antennule (large arrows) and antenna (thin arrows). Scale bar = 10 μ m.

Regarding host specificity, the major ergasilids species show no specificity to parasitize a single host. Indeed, *E. coatiarus* was also reported parasiting *Cichla orinocensis* Humboldt, 1821 and *Cichla temensis* Humboldt, 1821 [16,18], evidencing a low host-specificity of this parasite. In the present study, we reported *C. semifasciatus* with a new host record to *E. coatiarus*, despite that low prevalence was observed in the specimens that were examined. However, according to Tavares-Dias et al. [18], *E. coatiarus* has preference to parasitize fish of the family Cichlidae. This remark may be more related to the consideration that some parasites species prefer hosts with a similar behavior, habitats, additionally to ecological factors [19,20].

In our study, despite the high occurrence of *E. coatiarus*, no apparent lesions were observed in the fishes. A possible reason may be due to the fact that fishes have shown low level of parasitism. These findings are similar to those of Araujo and Varella [8] and Araujo et al. [16] who reported no disease symptoms in *C. monoculus* parasitized by *E. coatiarus*. However, it is well-known that when ectoparasites, in particular copepods and monogeneans species, are present in sufficient number in the gill, they may cause tissue damage and obstruct blood flow, thereby compromising respiratory capacity [1,7,21], taking in account that the gill is the major respiratory organ and

plays an important role in nitrogenous waste excretion and in the ionic balance [22].

The results of the present study show that there is not a relationship between the prevalence of parasites and size of examined fish. This finding is in agreement with those described by Ferrari-Hoeinghaus et al. [23], who reported no influence of host size on the prevalence and intensity of parasitism by *Amphithecium* sp., and *Notozothecium* sp. in *Astyanax altiparanae* Garutti and Britski, 2000 and *Urocleidoides mastigatus* Suriano, 1986 and *Scleroductus* sp. in *Rhamdia quelen* Quoy and Gaimard, 1824, but contrast with Siddiqui et al. [24] and Saha et al. [25], who reported a correlation between host size with prevalence and intensity of protozoan parasites in ornamental fish from India.

Although, in our study all water parameters remained within acceptable values for cultivation of tropical fish [26], a moderate prevalence of *E. coatiarus* was observed in the fish that were examined. In this context, using natural controls to prevent the spread of ectoparasites in fish culture become important, in order to promote avoidance of highly toxic products that could kill or leave the host fish unfeasible for human consumption [7,27]. For instance, Barker and Cone [27] suggest that flow rates above 5 cm/s should impair the transmission of *Ergasilus celestis* Mueller, 1937 and *Pseudodactylogryrus anguillae* Yin and Sproston,

1948 in eel aquaculture. Hence, the first recommendation should be the establishment of a water circulation system and manipulation of flow rates in the ponds culture of *C. monoculus* and *C. semifasciatus*, preventing the transmission of free-swimming stage of ergasilids and other ectoparasites.

Likewise, the use of temperature and pH manipulation in order to reduce ergasilids infestation could be another recommendation to control these parasites in this aquaculture facility, considering that rates of oviposition and egg hatching of Ergasilus spp. are greater above 23 °C [28]. However, despite that the use of temperature to control fish parasites has had much success with controlling of infections for some taxa of parasites [28,29], manipulation of these abiotic parameters in fish farms from the Amazon region, must take into account the tolerance and sensitivity of fish species that are cultivated. Another recommendation would be continuous exchange of the water to promote elimination of large accumulation of organic matter on the pond bottom, preventing hypoxia and anoxia, which lead the fish more susceptible not only to ectoparasites but also to opportunistic bacteria [30]. Nevertheless, the manipulation of physical parameters in the environment on fish farms of other geographic regions had shown to be effective in order to minimize the transmission of parasites. In the Amazon region, information about this method is still scarce. In this context, future studies about the influence of some abiotic factors on levels of parasitism of gill parasites are highly recommended to prevent and control spreading of ergasilids and other ectoparasites.

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References

- W. Piasecki, A.E. Goodwin, J.C. Eiras, B.F. Nowak, Importance of copepoda in freshwater aquaculture, Zool. Stud. 43 (2004) 193–205.
- [2] V.E. Thatcher, Amazon fish parasites, Sofia, Bulgaria, 2006, [Second ed.].
 [3] S.O. Kullander, E.J.G. Ferreira, A review of the South American cichlid
- [5] So. Kunanter, E.J.G. Friena, A review of the order South Anterical Central genus Cichla, with descriptions of nine new species (Teleostei: Cichlidae), Ichthyol. Explor. Freshwaters 17 (2006) 289–398.
- [4] E.M. Bayoumy, H.B. Baghdadi, M.E.A. Hassanain, Light and scan electron microscopes on *Caligus kuwaitensis* (Copepoda: Siphonostomatoida), from Arabian Gulf Doubleur Bream, *Acanthopagrus bifasciatus*, off Dammam, Saudi Arabia, Aust. J. Basic Appl. Sci. 7 (2013) 974–978.
- [5] A.T. Yuniar, H.W. Palm, T. Walter, Crustacean fish parasites from Segara Anakan Lagoon, Java, Indonesia, Parasitol. Res. 100 (2007) 1193–1204.
- [6] C. Boualleg, H. Ferhati, N. Kaouachi, M. Bensouilahand, S. Ternengo, The Copepod parasite of the gills of four teleost fishes caught from the gulf of Annaba (Algeria), Afr. J. Microbiol. Res. 4 (2010) 801–807.
- [7] P.D. Mathews, D.J.P. Mathews, A.J. Vega, O.R. Ismiño, Massive infestation by *Perulernaea gamitanae* (Crustacea: Cyclopoida: Lernaidae) in juvenile gamitana, cultured in the Peruvian Amazon, Vet, Mex 42 (2011) 59–64.

- [8] C.S.A. Araujo, A. Varella, Ergasilus coatiarus sp. n. (Copepoda, Poecilostomatoida, Ergasilidae) parasite from the gills of Cichla monoculus Spix, 1831 (Perciformes: Cichlidae) from Brazilian Amazon, Acta Amaz. 28 (1998) 417–424.
- [9] A.O. Bush, K.D. Lafferty, J.M. Lotz, A.W. Shostak, Parasitology meets ecology on its own terms: Margolis et al. revisited, J. Parasitol. 3 (1997) 575–583.
- [10] P.D. Mathews, D.J.P. Mathews, O.R. Ismiño, Massive infestation by *Gussevia undulata* (Platyhelminthes: Monogenea: Dactylogyridae) in fingerlings of *Cichla monoculus* cultured in the Peruvian Amazon, Neotrop. Helminthol. 6 (2012) 231–237.
- [11] P.D. Mathews, J.P.D. Mathews, R.O. Ismiño, Parasitic infections in juveniles of *Prochilodus nigricans* ket in a semi-intensive fish farm in the Peruvian Amazon, Bull. Eur. Ass. Fish. Pathol. 33 (2013) 28–32.
- [12] P.D. Mathews, A.F. Malheiros, O.R. Ismiño, N.D. Vasquez, Jainus amazonensis (Monogenea: Dactylogyridae) parasites of Brycon cephalus (Günther, 1869) cultured in the lowland of the Peruvian Amazon, Croat. J. Fisheries 72 (2014) 83–86.
- [13] L. Soberon, P. Mathews, A. Malheiros, Hematological parameters of Colossoma macropomum naturally parasitized by Anacanthorus spathulatus (Monogenea: Dactylogiridae) in fish farm in the Peruvian Amazon, Int. Aquat. Res. 6 (2014) 251–255.
- [14] J.L. Luque, C. Cruces, J. Chero, F. Paschoal, P.V. Alves, A.C. Da Silva, et al., Checklist of metazoan parasites of fishes from Peru, Neotrop. Helminthol. 10 (2016) 301–375.
- [15] A.F. Gonzales, P.D. Mathews, L.E. Luna, J.D. Mathews, Outbreak of Notozothecium bethae (Monogenea: Dactylogyridae) in Myleus schomburgkii (Actinopterygii: Characiformes) cultured in the Peruvian Amazon, J. Parasit. Dis. 40 (2016) 1631–1635.
- [16] C.S.O. Araujo, M.C. Barros, A.L.S. Gomes, A.M.B. Varella, G.M. Viana, N.P. Silva, et al., Parasites of natural and artificial populations of *Cichla* spp., Rev. Bras. Parasitol. Vet. 18 (2009) 34–38.
- [17] R. Abdel-Gaber, F. Abdel-Ghaffar, S. Maher, A. El-Mallah, S. Al Quraishy, H. Mehlhorn, Morphological re-description and phylogenetic relationship of five myxosporean species of family Myxobolidae infecting the Nile tilapia Oreochromis niloticus (Perciformes: cichlidae), Dis. Aquat. Org. 124 (2017) 201–214.
- [18] M. Tavares-Dias, M.B.F. Dias-Júnior, A.C. Florentino, L.M.A. Silva, A.C. Cunha, Distribution pattern of crustacean ectoparasites of freshwater fish from Brazil, Rev. Bras. Parasitol. Vet. 24 (2015) 136–147.
- [19] V.E. Thatcher, Copepods and fishes in the Brazilian Amazon, J. Mar. Syst. 15 (1998) 97–112.
- [20] M. Mamani, C. Hamel, P.A. Van Damme, Ectoparasites (Crustacea: Branchiura) of *Pseudoplatystoma fasciatum* (surubí) and *P. tigrinum* (chuncuina) in Bolivian whitewater floodplains, Ecol. Boliv. 39 (2004) 9–20.
- [21] I. Paperna, Parasites and diseases of the grey mullet (Mugilidae) with special reference to the seas of the Near East, Aquaculture 5 (1975) 65– 80.
- [22] E.J. Noga, Fish disease: diagnosis and treatment, Blackwell, Ames, 2000, [Second ed.].
- [23] A.P. Ferrari-Hoeinghaus, R.M. Takemoto, L.C. Oliveira, M.C. Makrakis, G. Baumgartner, Host-parasite relationships of monogeneans in gills of *Astyanax altiparanae* and *Rhamdia quelen* of the São Francisco Verdadeiro River, Brazil, Parasite 13 (2006) 315–320.
- [24] A.A. Siddiqui, Effects of seasons, host age, size and sex on monogenetic trematode, *Hamatopeduncularia indicus* of host fish, *Arius jella*, J. Chem. Biol. Phys. Sci. 4 (2014) 1146–1151.
- [25] M. Saha, P.K. Bandyopadhyay, A. Roy, S. Ghosh, Impact of seasons, host age, size and sex on the prevalence of protozoan parasites in ornamental fish, J. Agric. Vet. Sci. 8 (2015) 54–59.
- [26] L.H. Sipaúba-Tavares, J.G. Durigan, S.R. Ligeiro, Characterization of some limnological variables in a nursery for fish farming in two periods of the day, Rev. UNIMAR 16 (1994) 217–227.
- [27] D.E. Barker, D.K. Cone, Occurrence of Ergasilus celestis (Copepoda) and Pseudodactylogryrus anguilae (Monogenea) among wild eels (Anguilla rostrata) in relation to stream flow, pH and temperature and recommendations for controlling their transmission among captive eels, Aquaculture 187 (2000) 261–274.
- [28] N.C. Marchioria, E.L.T. Gonçalves, K.R. Tancredo, J. Pereira-Junior, J.R.E. Garcia, M.L. Martins, Effect of water temperature and salinity in oviposition, hatching success and infestation of *Aphanoblastella mastigatus* (Monogenea, Dactylogyridae) on *Rhamdia quelen*, Braz. J. Biol. 75 (2015) 245–252.
- [29] P.T.K. Woo, S.L. Poynton, Diplomonadida, kinetoplastida and amoebida Phylum Sarcomastigophora, in: P.T.K. Woo (Ed.), Fish diseases and disorders, CAB International, UK, 1995, pp. 27–96.
- [30] F. Garcia, F. Pilarskia, E.M. Onakac, F.R. Moraes, M.L. Martins, Hematology of *Piaractus mesopotamicus* fed diets supplemented with vitamins C and E, challenged by *Aeromonas hydrophila*, Aquaculture 271 (2007) 39–46.