



INSTITUT DE FRANCE
Académie des sciences

Comptes Rendus

Biologies

Isabelle K. Vila and Nadine Laguette

The unexpected role of the STING protein in lipid metabolism

Volume 346 (2023), p. 29-33

Published online: 18 April 2023

<https://doi.org/10.5802/crbio.110>



This article is licensed under the
CREATIVE COMMONS ATTRIBUTION 4.0 INTERNATIONAL LICENSE.
<http://creativecommons.org/licenses/by/4.0/>



*Les Comptes Rendus. Biologies sont membres du
Centre Mersenne pour l'édition scientifique ouverte*

www.centre-mersenne.org

e-ISSN : 1768-3238



Review Article / Article de revue

The unexpected role of the STING protein in lipid metabolism

Le rôle inattendu de la protéine STING dans le métabolisme des lipides

Isabelle K. Vila[✉]*, ^a and Nadine Laguette[✉]*, ^a

^a IGMM, Université de Montpellier, CNRS, Montpellier, France

E-mails: isabelle.vila@cnrs.fr (I. K. Vila), nadine.laguette@cnrs.fr (N. Laguette)

Abstract. Detection of cytosolic pathological nucleic acids is a key step for the initiation of innate immune responses. In the past decade, the stimulator of interferon genes (STING) adaptor protein has emerged as a central platform enabling the activation of inflammatory responses in the presence of cytosolic DNAs. This has prompted a plethora of approaches aiming at modulating STING activation in order to boost or inhibit inflammatory responses. However, recent work has revealed that STING is also a direct regulator of metabolic homeostasis. In particular, STING regulates lipid metabolism directly, a function that is conserved throughout evolution. This indicates that STING targeting strategies must take into consideration potential metabolic side effects that may alter disease course, but also suggests that targeting STING may open the route to novel treatments for metabolic disorders. Here we discuss recent work describing the metabolic function of STING and the implications of these findings.

Résumé. La détection des acides nucléiques pathologiques cytosoliques est une étape clé pour le déclenchement des réponses immunitaires innées. Au cours de la dernière décennie, la protéine adaptatrice STING (stimulator of interferon genes) est apparue comme une plateforme centrale permettant l'activation des réponses inflammatoires en présence d'ADN cytosolique. Cela a donné lieu à une multitude d'approches visant à moduler l'activation de STING afin de stimuler ou d'inhiber les réponses inflammatoires. Cependant, des travaux récents ont révélé que STING est également un régulateur direct de l'homéostasie métabolique. En particulier, STING régule directement le métabolisme des lipides, une fonction qui est conservée au cours de l'évolution. Cela indique que les stratégies de ciblage de STING doivent prendre en compte les effets secondaires métaboliques potentiels qui peuvent modifier l'évolution de la maladie, mais suggère également la possibilité que le ciblage de STING puisse ouvrir la voie à de nouvelles façons de traiter les pathologies présentant une composante métabolique. Nous discutons ici les travaux récents décrivant la fonction métabolique de STING et les implications de ces résultats.

Keywords. STING, Metabolism, Innate immunity, Cytosolic DNA, Fatty acids, Inflammation.

Mots-clés. STING, Métabolisme, Immunité innée, ADN cytosolique, Acides gras, Inflammation.

Published online: 18 April 2023

* Corresponding authors.

1. STING, a central protein of the immune response associated with cytosolic nucleic acids

The innate immune system is the first line of defence against pathogens. One of the central pathways for pathogen detection is the recognition of their nucleic acid content by intracellular receptors. A large panel of receptors involved in the initiation of the innate immune response has been described [1]. Their interaction with pathological nucleic acids leads to the activation of signalling pathways that induce the production of inflammatory cytokines. This inflammatory response mobilises effector cells of the adaptive immune system and is therefore a crucial step in the initiation, maintenance and coordination of immune responses.

Recently, a central pathway for the detection of cytosolic nucleic acids has been described (Figure 1). This pathway was originally identified in immune cells as activated by cytosolic double-stranded DNAs during viral or bacterial infections [2]. This pathway relies on the cyclic GMP-AMP synthase (cGAS) receptor and the Stimulator of Interferon Genes (STING) adaptor protein. The cGAS receptor recognises cytosolic nucleic acids, including double-stranded DNA (dsDNA), single-stranded DNA (ssDNA) or DNA:RNA hybrids [3–5]. The interaction of cGAS with cytosolic nucleic acids induces its activation, enabling its capacity to synthesize the 2'3'-cGAMP cyclic dinucleotide from ATP and GTP [6]. This secondary messenger interacts with the adaptor protein STING, promoting the assembly of a signalosome in which STING serves as an anchoring site for the various partners required for the induction of the inflammatory response. Those partners notably include the protein TANK binding kinase 1 (TBK1) and the interferon regulatory factor 3 (IRF3) and/or nuclear factor κ B (NF- κ B) transcript factors [7, 8]. Activation of the cGAS-STING signalling axis is characterised by the production of type I interferons, besides other pro-inflammatory cytokines and chemokines. Thus, this signalling pathway is particularly critical in the response to pathogens, as the production of type I interferons leads to the establishment of an antiviral state through induction of the expression of interferon response genes.

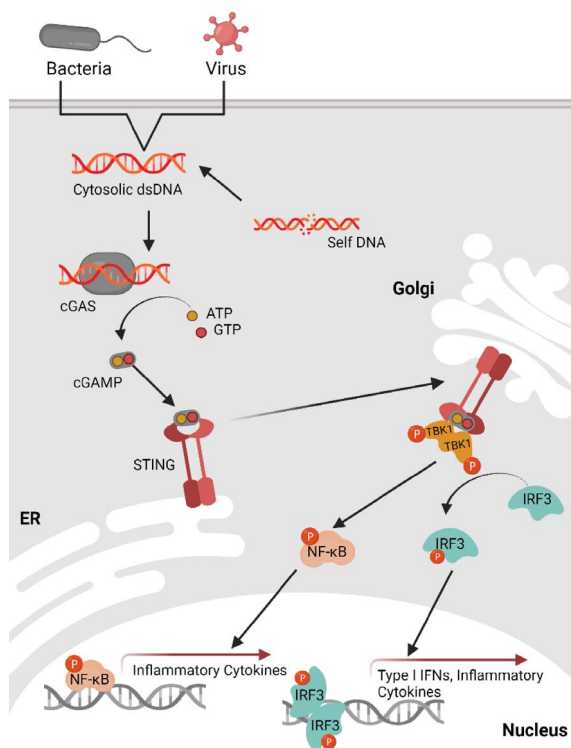


Figure 1. Overview of the cytosolic DNA sensing via the cGAS-STING pathway. dsDNA presence in the cytosol can result from pathogen infection or self-DNA after cellular stress. cGAS binds dsDNA leading to cGAS activation and synthesis of 2,3'-cGAMP from ATP and GTP. cGAMP binds to ER-resident STING, this binding will trigger STING translocation to the Golgi apparatus. In the Golgi, STING binds to and activates TBK1 (by autophosphorylation) as well as the transcription factor NF- κ B via IKK. TBK1 then phosphorylates IRF3 which undergoes dimerization and translocates to the nucleus to induce transcription of type I IFNs and several inflammatory chemokines. NF- κ B induces transcription of inflammatory cytokines. Created with Biorender.

Recent work suggests that cGAS and STING beyond their canonical function in innate immunity, are also involved in cellular processes such as the induction and maintenance of senescence or the regulation of cell proliferation [9–11].

2. STING in fatty acid metabolism: an unexpected role

In 2022, we revealed a central role of STING in energy and glucose homeostasis via the regulation of adipose tissue metabolism [12] (Figure 2). Using a mouse model deleted for the STING protein [13], this study showed that STING inhibits the fatty acid desaturase 2 (FADS2) enzyme essential for the desaturation of dietary polyunsaturated fatty acids (PUFAs). Therefore, ablation or agonist-induced degradation of STING increased FADS2 activity, leading to the accumulation of PUFA derivatives and driving thermogenic gene activation. As a result, white adipose tissue showed signs of a browning process. Notably, white adipose depots were metabolically more active, burning energy through adaptive thermogenesis. PUFAs were also shown to bind STING and inhibit its activity. Modulation of PUFA production thus regulates STING-dependent antiviral responses while contributing to the resolution of STING-associated inflammation. Finally, this study also revealed that STING agonists can bind and modulate the enzymatic activity of FADS2. Thus, this work demonstrates the existence of a negative regulatory loop between STING and FADS2, opening many questions pertaining to their respective roles in human pathologies presenting with a chronic STING-associated inflammation [14].

Interestingly, alleles of STING not competent for the induction of interferon responses have been documented in human populations. These include the STING HAQ allele that bears three amino acid substitutions: R71H-I229A-R292Q [15]. Analysis of STING haplotypes revealed a high prevalence of the HAQ allele in East Asian populations as compared to sub-Saharan African populations. Conversely, sub-Saharan African populations show a high prevalence of an AQ allele (G230A, R293Q) competent for the recognition of cyclic dinucleotides and the induction of interferon responses [16]. The authors propose that these STING alleles (HAQ and AQ) underwent natural selection during migration out of Africa 50,000–70,000 years ago [16]. Using mouse models expressing the HAQ or AQ human STING alleles, they showed that AQ mice store less lipid than HAQ mice, while displaying improved fatty acid oxidation. Furthermore, the visceral white adipose tissue of HAQ mice was infiltrated by regulatory T cells and

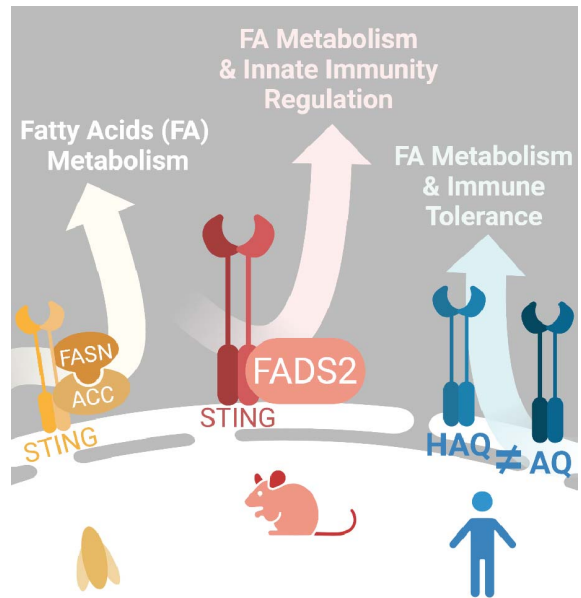


Figure 2. STING plays a role in lipid metabolism. Convergent data in 2022 showed that STING regulates lipid metabolism. In *Drosophila*, STING form a multi-protein complex with two enzymes involved in long-chain fatty acid synthesis FASN and ACC. In mouse STING regulates the activity of the lipid desaturase FADS2 modulating the synthesis of long-chain polyunsaturated fatty acid. The production of those lipid species will in turn modulate STING activity and narrow its activation in presence of its agonist cGAMP. In human populations several alleles exist and were already known to be more or less potent in the induction of IFN response. The study of AQ and HAQ alleles showed that they also differ in lipid metabolism modulation as HAQ promotes more fat storage than AQ. Created with Biorender.

anti-inflammatory macrophages, unlike that of AQ mice. Thus, the HAQ haplotype promotes fat storage, while the AQ haplotype promotes immune tolerance. As the HAQ allele confers better energy storage capacity than the AQ allele, they conclude that this ability to store energy could have been critical during ice age for the survival of early humans [17]. The study by Vila *et al.* showed that the HAQ allele of STING strongly interacts with FASD2 [12], which is consistent with the description of a prominent role of this

allele in fatty acid metabolism [16]. Thus, the study of the evolution of STING haplotypes suggests that the function of STING in modulating fatty acid metabolism may have been critical during human migration out of Africa.

In support of this hypothesis, work in *Drosophila* [18] confirmed the conservation of the functional link between STING and fatty acid metabolism. The authors showed that the *Drosophila* STING orthologue interacts with enzymes involved in the biosynthesis of long-chain fatty acids such as acetyl-CoA carboxylase (ACC) and fatty acid synthetase (FASN) within a multiprotein complex. Furthermore, the absence of STING in *Drosophila* led to decreased lipid storage and decreased expression of genes related to lipid metabolism. Thus, the absence of STING increased sensitivity to food deprivation and oxidative stress in *Drosophila*. These results showing profound lipid metabolism alteration in *Drosophila* support a conserved role for STING in the regulation of lipid metabolism.

Finally the FADS (FADS1, FADS2 and FADS3) genomic region was found to be under selection in Greenlandic Inuit, probably driven by a diet high in PUFAs [19]. Those results confirming FADS genetic selection among populations [20] suggest that these genes play an important role in human adaptation to diets. Thus the study of STING and FADS allele co-evolution could bring new insight into their synergistic role in inflammation and metabolic functions.

3. Regulation of lipid metabolism by STING: a link to innate immune regulation?

Although the role of STING in host defence mechanisms against pathogens is well established, these recent findings on the link between STING and fatty acid metabolism demonstrate that this protein has important functions beyond the detection of cytosolic nucleic acids. Evolutionary analysis of STING suggests that the modulation of lipid metabolism is ancient, conserved and may be the primordial function of STING.

Metabolic pathways that interact with STING, such as those leading to the production of PUFAs, are also involved in the control of STING-independent inflammatory responses. Indeed, while PUFAs can

directly inhibit STING [12], they can also be processed into lipid mediators (oxylipins) that possess, among others, immunomodulatory properties [21]. Thus, it can be hypothesised that STING alleles expressed in organisms lacking an interferon signalling pathway could participate to the control of inflammatory responses by regulating the generation of these lipid metabolites.

Altogether, these recent studies [12, 16, 18] highlight a tight link between innate immunity and lipid metabolism, paving the way for research into the role of metabolic alterations in human pathologies associated with aberrant STING activation [22]. Those notably include cancer, where a crucial role of a functional cGAS-STING axis has been recently shown to be essential for driving tumour immunogenicity [23, 24]. The role of STING-dependent metabolic regulation remains to be investigated in this context.

Conflicts of interest

Authors have no conflict of interest to declare.

References

- [1] O. Takeuchi, S. Akira, "Pattern recognition receptors and inflammation", *Cell* **140** (2010), no. 6, p. 805-820.
- [2] J. Tao, X. Zhou, Z. Jiang, "cGAS-cGAMP-STING: The three musketeers of cytosolic DNA sensing and signaling", *IUBMB Life* **68** (2016), no. 11, p. 858-870.
- [3] A. K. Mankan, T. Schmidt, D. Chauhan, M. Goldeck, K. Höning, M. Gaidt, A. V. Kubarenko, L. Andreeva, K. P. Hopfner, V. Hornung, "Cytosolic RNA:DNA hybrids activate the cGAS-STING axis", *EMBO J.* **33** (2014), no. 24, p. 2937-2946.
- [4] P. J. Kranzusch, A. S. Lee, J. M. Berger, J. A. Doudna, "Structure of human cGAS reveals a conserved family of second-messenger enzymes in innate immunity", *Cell Rep.* **3** (2013), no. 5, p. 1362-1368.
- [5] L. Sun, J. Wu, F. Du, X. Chen, Z. J. Chen, "Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway", *Science* **339** (2013), no. 6121, p. 786-791.
- [6] A. Ablasser, M. Goldeck, T. Cavlar, T. Deimling, G. Witte, I. Röhl, K. P. Hopfner, J. Ludwig, V. Hornung, "cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates STING", *Nature* **498** (2013), no. 7454, p. 380-384.
- [7] S. Liu, X. Cai, J. Wu, Q. Cong, X. Chen, T. Li, F. Du, J. Ren, Y. T. Wu, N. V. Grishin, Z. J. Chen, "Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3 activation", *Science* **347** (2015), no. 6227, article no. aaa2630.
- [8] T. Abe, G. N. Barber, "Cytosolic-DNA-mediated, STING-dependent proinflammatory gene induction necessitates canonical NF- κ B activation through TBK1", *J. Virol.* **88** (2014), no. 10, p. 5328-5341.

- [9] H. Jiang, X. Xue, S. Panda, A. Kawale, R. M. Hooy, F. Liang, J. Sohn, P. Sung, N. O. Gekara, "Chromatin-bound cGAS is an inhibitor of DNA repair and hence accelerates genome destabilization and cell death", *EMBO J.* **38** (2019), no. 21, article no. e102718.
- [10] S. Glück, B. Guey, M. F. Gulen, K. Wolter, T. W. Kang, N. A. Schmacke, A. Bridgeman, J. Rehwinkel, L. Zender, A. Ablasser, "Innate immune sensing of cytosolic chromatin fragments through cGAS promotes senescence", *Nat. Cell Biol.* **19** (2017), no. 9, p. 1061-1070.
- [11] S. Cerboni, N. Jeremiah, M. Gentili, U. Gehrman, C. Conrad, M. C. Stolzenberg, C. Picard, B. Neven, A. Fischer, S. Amigorena, F. Rieux-Laucat, N. Manel, "Intrinsic antiproliferative activity of the innate sensor STING in T lymphocytes", *J. Exp. Med.* **214** (2017), no. 6, p. 1769-1785.
- [12] I. K. Vila, H. Chamma, A. Steer, M. Saccas, C. Taffoni, E. Turtoi, L. S. Reinert, S. Hussain, J. Marines, L. Jin, X. Bonnefont, M. Hubert, O. Schwartz, S. R. Paludan, G. Van Simaey, G. Doumont, B. Sobhian, D. Vlachakis, A. Turtoi, N. Laguette, "STING orchestrates the crosstalk between polyunsaturated fatty acid metabolism and inflammatory responses", *Cell Metab.* **34** (2022), no. 1, p. 125-139.e8.
- [13] L. Jin, A. Getahun, H. M. Knowles, J. Mogan, L. J. Akerlund, T. A. Packard, A. L. Perraud, J. C. Cambier, "STING/MPYS mediates host defense against *Listeria monocytogenes* infection by regulating Ly6C(hi) monocyte migration", *J. Immunol.* **190** (2013), no. 6, p. 2835-2843.
- [14] I. K. Vila, S. Guha, J. Kalucka, D. Olagnier, N. Laguette, "Alternative pathways driven by STING: From innate immunity to lipid metabolism", *Cytokine Growth Factor Rev.* **68** (2022), p. 54-68.
- [15] L. Jin, L. G. Xu, I. V. Yang, E. J. Davidson, D. A. Schwartz, M. M. Wurfel, J. C. Cambier, "Identification and characterization of a loss-of-function human MPYS variant", *Genes Immun.* **12** (2011), no. 4, p. 263-269.
- [16] S. Mansouri, H. Gogoi, S. Patel, D. S. Katikaneni, A. Singh, A. Aybar-Torres, G. de Lartigue, L. Jin, "MPYS modulates fatty acid metabolism and immune tolerance at homeostasis independent of type I IFNs", *J. Immunol.* **209** (2022), no. 11, p. 2114-2132.
- [17] R. J. Johnson, M. A. Lanaspá, J. W. Fox, "Upper paleolithic figurines showing women with obesity may represent survival symbols of climatic change", *Obesity* **29** (2021), no. 1, p. 11-15.
- [18] K. Akhmetova, M. Balasov, I. Chesnokov, "*Drosophila* STING protein has a role in lipid metabolism", *eLife* **10** (2021), article no. e67358.
- [19] M. Fumagalli, I. Moltke, N. Grarup, F. Racimo, P. Bjerregaard, M. E. Jørgensen, T. S. Korneliusen, P. Gerbault, L. Skotte, A. Linneberg, C. Christensen, I. Brandslund, T. Jørgensen, E. Huerta-Sánchez, E. B. Schmidt, O. Pedersen, T. Hansen, A. Albrechtsen, R. Nielsen, "Greenlandic Inuit show genetic signatures of diet and climate adaptation", *Science* **349** (2015), no. 6254, p. 1343-1347.
- [20] A. Ameur, S. Enroth, A. Johansson, G. Zaboli, W. Igl, A. C. Johansson, M. A. Rivas, M. J. Daly, G. Schmitz, A. A. Hicks, T. Meitinger, L. Feuk, C. van Duijn, B. Oostra, P. P. Pramstaller, I. Rudan, A. F. Wright, J. F. Wilson, H. Campbell, U. Gyllenstein, "Genetic adaptation of fatty-acid metabolism: a human-specific haplotype increasing the biosynthesis of long-chain omega-3 and omega-6 fatty acids", *Am. J. Hum. Genet.* **90** (2012), no. 5, p. 809-820.
- [21] M. Gabbs, S. Leng, J. G. Devassy, M. Monirujaman, H. M. Aukema, "Advances in our understanding of oxylipins derived from dietary PUFAs", *Adv. Nutr.* **6** (2015), no. 5, p. 513-540.
- [22] I. K. Vila, M. Fretaud, D. Vlachakis, N. Laguette, C. Langevin, "Animal models for the study of nucleic acid immunity: novel tools and new perspectives", *J. Mol. Biol.* **432** (2020), no. 20, p. 5529-5543.
- [23] C. Taffoni, J. Marines, H. Chamma, S. Guha, M. Saccas, A. Bouzid, A. C. Valadao, C. Maghe, J. Jardine, M. K. Park, K. Polak, M. De Martino, C. Vanpouille-Box, M. Del Rio, C. Gongora, J. Gavard, N. Bidère, M. S. Song, D. Pineau, J. P. Hugnot, K. Kissa, L. Fontenille, F. P. Blanchet, I. K. Vila, N. Laguette, "DNA damage repair kinase DNA-PK and cGAS synergize to induce cancer-related inflammation in glioblastoma", *EMBO J.* **42** (2023), no. 7, article no. e111961.
- [24] C. Hong, M. Schubert, A. E. Tijhuis, M. Requesens, M. Roroda, A. van den Brink, L. A. Ruiz, P. L. Bakker, T. van der Sluis, W. Pieters, M. Chen, R. Wardenaar, B. van der Vegt, D. C. J. Spierings, M. de Bruyn, M. van Vugt, F. Foijer, "cGAS-STING drives the IL-6-dependent survival of chromosomally unstable cancers", *Nature* **607** (2022), no. 7918, p. 366-373.