



Supplementary material: Synthesis and crystal structures of palladium complexes based on α -amino-oximes derived from (*R*)-limonene and their application in allylic alkylation of 1,3-dioxo compounds

Yasmina Homrani^{a, b}, Mohamed Amin El Amrani^{*, a}, Pauline Loxq^b, Frédéric Capet^{® b}, Isabelle Suisse^{® b} and Mathieu Sauthier^{® *, b}

^a Laboratoire de Chimie Organique Appliquée, Faculté des Sciences, BP 2121, Université Abdelmalek Essaadi, Tétouan, Morocco

^b Univ. Lille, CNRS, Centrale Lille, Univ. Artois, UMR 8181, UCCS, Unité de Catalyse et Chimie du Solide, F-59000, Lille, France

E-mails: homrani.y@gmail.com (Y. Homrani), maelamrani@uae.ac.ma (M. A. El Amrani), pauline.loxq@gmail.com (P. Loxq), frederic.capet@univ-lille.fr (F. Capet), isabelle.suisse@univ-lille.fr (I. Suisse), mathieu.sauthier@univ-lille.fr (M. Sauthier)

1. General informations

The reactions were performed under nitrogen atmosphere using standard Schlenk line techniques. All reagents were commercial grade materials and were used without any further purification. Previously the solvents were dried and distilled under N₂ and deoxygenated through N₂ bubbling for 20 min.

Conversions were determined by gas chromatography (GC) on Shimadzu 2010 equipped with a HT-5 column (30 m, i.d. = 0.32 mm). The ee's (enantiomeric excess) were determined by HPLC using a column AD-H (hexane/*i*PrOH: 95/5, 1.0 mL/min, 25 °C).

The ¹H and ¹³C NMR spectra were recorded on a Bruker AC 300 spectrometer and referenced to TMS with eventually the presence of 1,3,5-trimethoxybenzene (TMB) as internal reference standard for quantitative NMR.

2. NMR spectra of products of the new compounds (Table 3)

The physical state of the following products is liquid after purification by silica gel column chromatography using petroleum ether/ethyl acetate (90/10) as eluent.

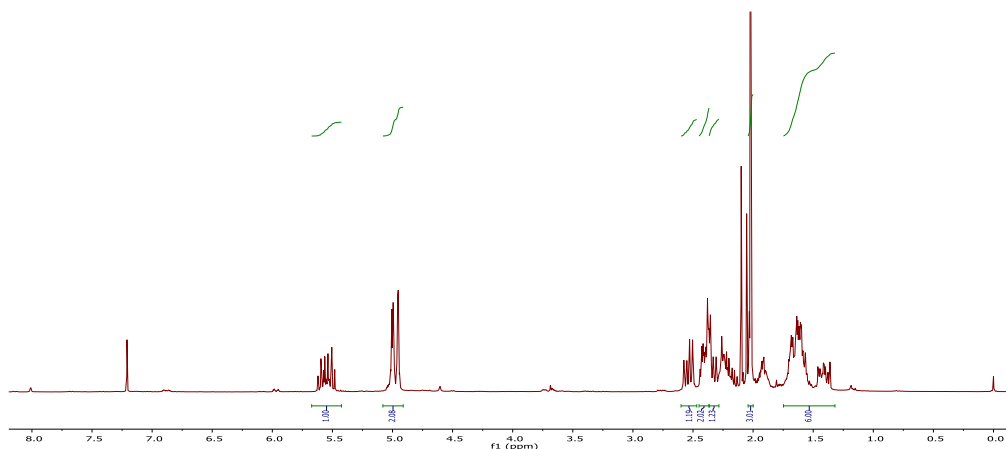
* Corresponding authors.

2.1. 2-acetyl-2-allylcyclohexanone¹ (**3b**)

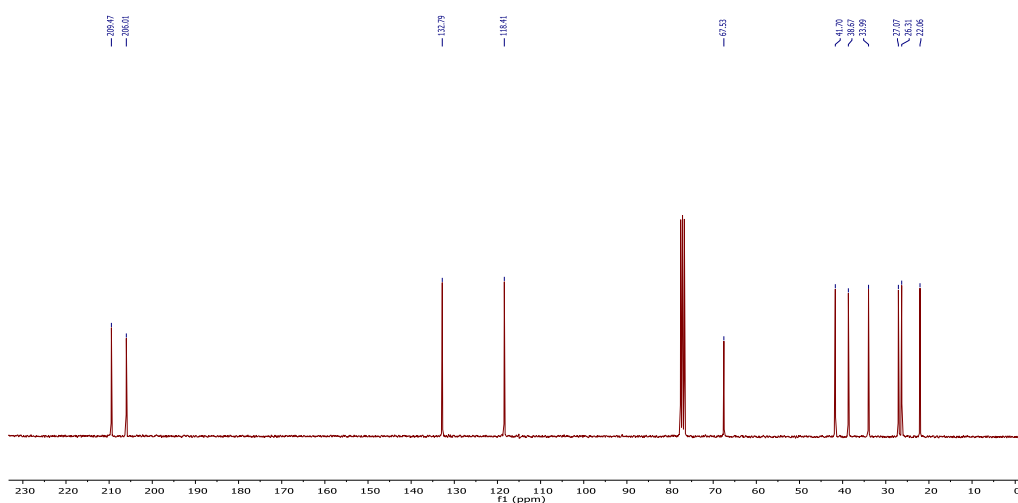
Yield: 72%



¹H NMR (300 MHz, CDCl₃): δ 5.68–5.57 (m, 1H, **-CH=**), 5.28–4.99 (m, 2H, **CH₂=CH**), 2.57–2.48 (m, $J = 5.4$ Hz, 1H, **-CH₂-CO**), 2.45–2.36 (m, 2H, **-CH₂CH=**), 2.35–2.28 (m, 1H, **CH₂-CO**), 2.10 (s, 3H, **CH₃**), 1.76–1.33 (m, 6H, **CCH₂CH₂CH₂CH₂CO**).

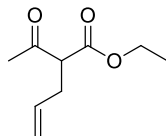
Supplementary Figure S1. ¹H NMR spectrum of **3b**.

¹³C NMR (75 MHz, CDCl₃) δ 209.47 (**C=O**), 206.01 (**C=O**), 132.79 (**CH=CH₂**), 118.41 (**CH₂=CHCH₂**), 67.53 (**Cq-CO**), 41.70 (**CH₂-CH₂CO**), 38.67 (**CH₂=CHCH₂**), 33.99 (**CH₂-CH₂CO**), 27.07 (**CH₃-CO**), 26.31 (**CH₂-CqCO**), 22.06 (**CH₂-CH₂Cq**).

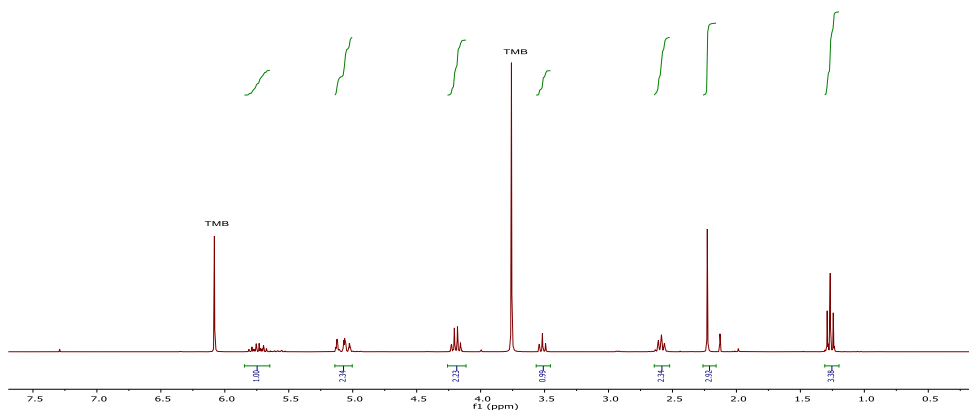
Supplementary Figure S2. ¹³C NMR spectrum of **3b**.

2.2. Ethyl 2-acetylpent-4-enoate¹ (**3c**)

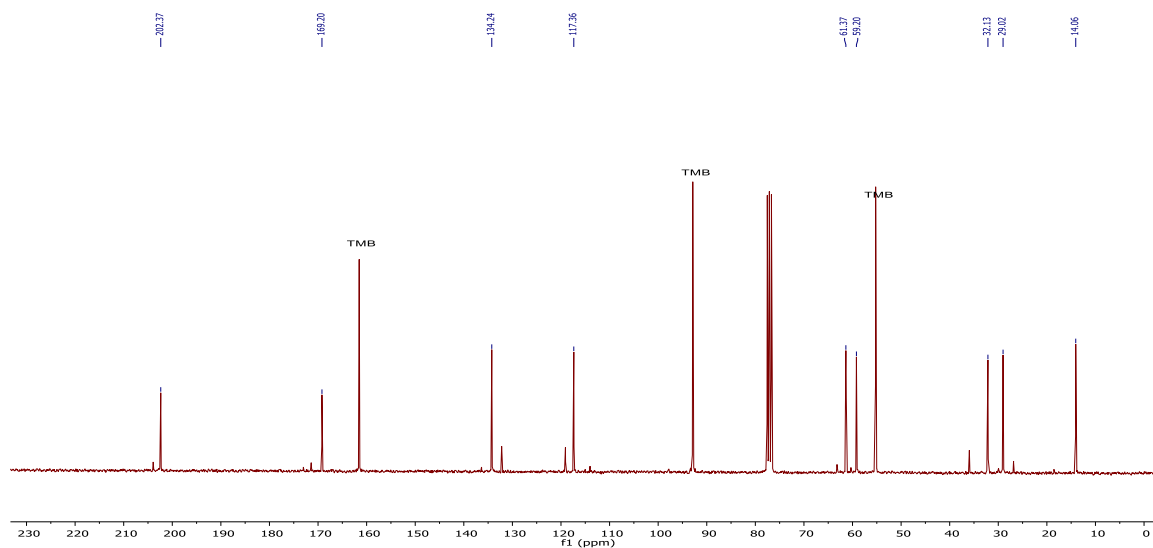
Yield: 42%



¹H NMR (300 MHz, CDCl₃) δ 5.74 (m, 1H, -CH=), 5.17–4.98 (m, 2H, CH₂=CH), 4.19 (q, *J* = 7.1 Hz, 2H, CH₃CH₂O), 3.52 (t, *J* = 7.4 Hz, 1H, CH-CO), 2.59 (m, *J* = 7.7, 6.7, 1.6, 0.8, 2H, -CH₂CH=), 2.23 (s, 3H, CH₃CO), 1.32–1.20 (t, 3H, CH₃CH₂O).

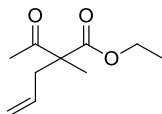
Supplementary Figure S3. ¹H NMR spectrum of **3c**.

¹³C NMR (75 MHz, CDCl₃) δ 202.37 (C=O), 169.20 (C=O), 134.24 (CH₂=CH-CH₂), 117.36 (CH₂=CH-CH₂), 61.37 (CH₃-CH₂O), 59.20 (CH-COOCH₂CH₃), 32.13 (CH₂=CH-CH₂-CH), 29.02 (CH₃-CO), 14.06 (CH₃-CH₂O).

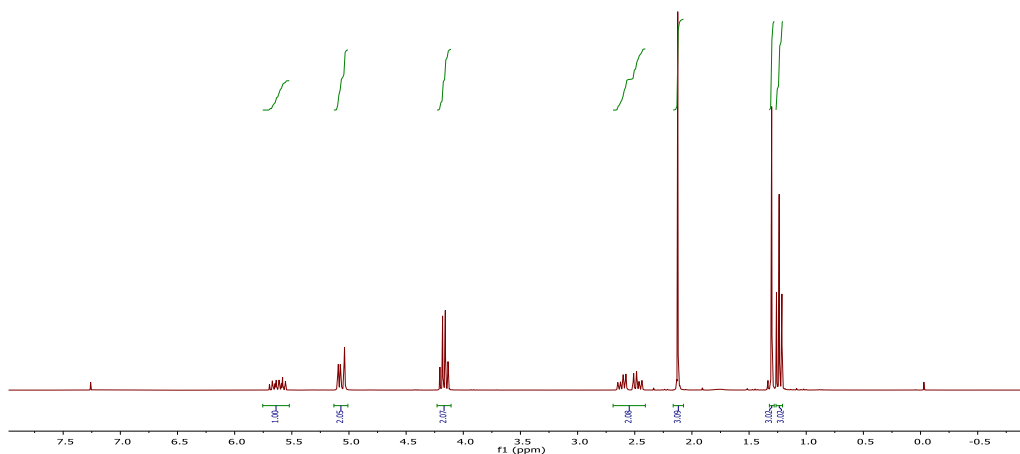
Supplementary Figure S4. ¹³C NMR spectrum of **3c**.

2.3. Ethyl 2-acetyl-2-methylpent-4-enoate¹ (**3d**)

Yield: 61%

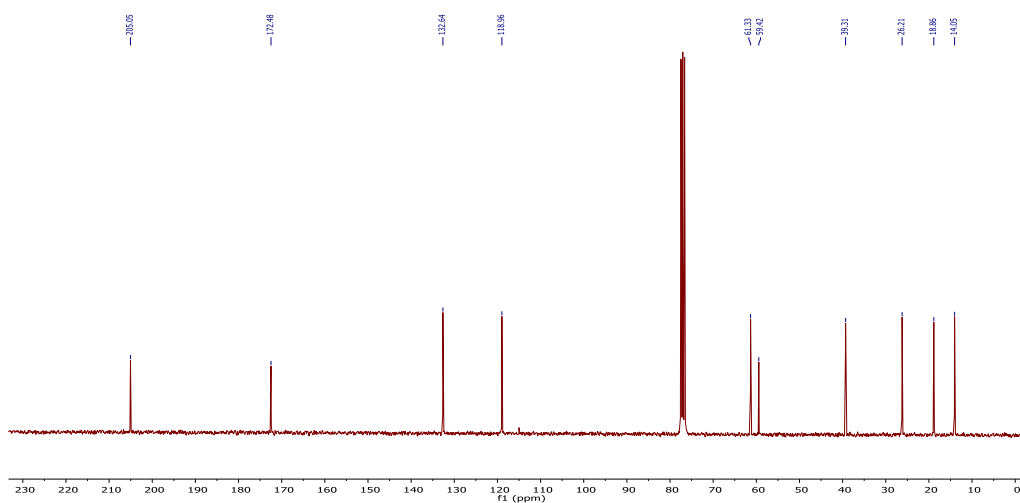


¹H NMR (300 MHz, CDCl₃) δ 5.72–5.52 (m, 1H, **-CH=**), 5.14–4.99 (m, 2H, **CH₂=CH**), 4.17 (q, *J* = 7.1 Hz, 2H, **CH₃CH₂O**), 2.69–2.40 (m, 2H, **-CH₂CH=**), 2.12 (s, 3H, **CH₃CO**), 1.30 (s, 3H, **CH₃-CCO**), 1.24 (t, *J* = 7.1 Hz, 3H, **CH₃CH₂O**).



Supplementary Figure S5. ¹H NMR spectrum of **3d**.

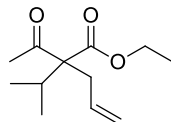
¹³C NMR (75 MHz, CDCl₃) δ 205.05 (**C=O**), 172.48 (**C=O**), 132.64 (**CH₂-CH=CH₂**), 118.96 (**CH₂=CH**), 61.33 (**CH₃-CH₂O**), 59.42 (**Cq-CO**), 39.31 (**=CH-CH₂Cq**), 26.21 (**CH₃-CO**), 18.86 (**CH₃-Cq**), 14.05 (**CH₃-CH₂O**).



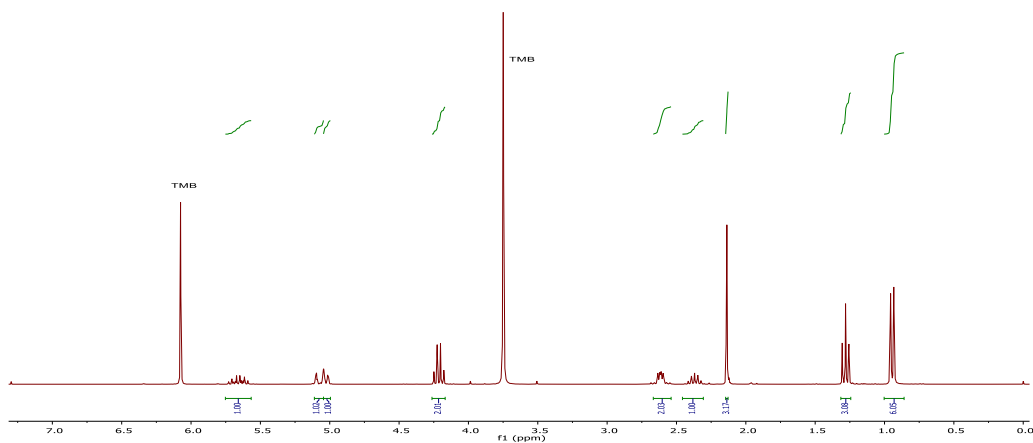
Supplementary Figure S6. ¹³C NMR spectrum of **3d**.

2.4. Ethyl 2-acetyl-3-isopropylpent-4-enoate (**3e**)

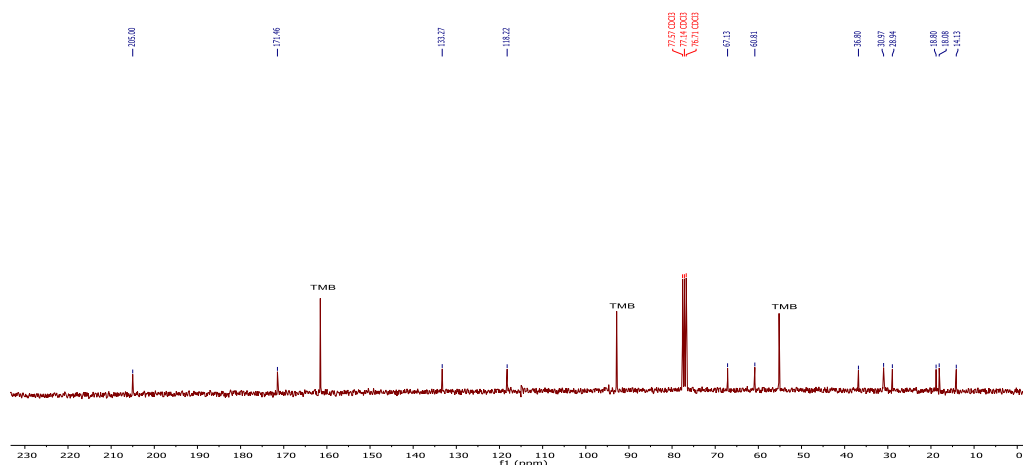
Yield: 24%



^1H NMR (300 MHz, CDCl_3) δ 5.66 (m, 2H, $J = 17.3, 10.1, 7.3$ Hz, $-\text{CH}=\text{}$), 5.04–4.99 (m, 2H, $\text{CH}_2=\text{}$), 4.24 (q, $J = 7.1$ Hz, 2H, $\text{CH}_3\text{CH}_2\text{O}$), 2.69–2.55 (m, 2H, $-\text{CH}_2\text{CH}=\text{}$), 2.37 (m, 1H, $\text{CH}^{\text{isopro}}$), 2.13 (s, 3H, CH_3-CO), 1.28 (t, $J = 7.1$ Hz, 3H, $\text{CH}_3\text{CH}_2\text{O}$), 0.94 (d, $J = 6.8$ Hz, 6H, $\text{CH}_3^{\text{isopro}}$).

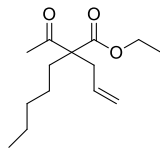
Supplementary Figure S7. ^1H NMR spectrum of **3e**.

^{13}C NMR (75 MHz, CDCl_3) δ 205.00 (C=O), 171.46 (C=O), 133.27 ($\text{CH}_2-\text{CH}=\text{CH}_2$), 118.22 ($\text{CH}_2=\text{CH}$), 67.13 ($\text{Cq}-\text{CH}-(\text{CH}_3)_2$), 60.81 ($\text{CH}_3-\text{CH}_2\text{O}$), 36.80 ($\text{CH}_2=\text{CH}-\text{CH}_2\text{Cq}$), 30.97 (CH_3-CO), 28.94 ($\text{CH}^{\text{isopro}}$), 18.80 (CH_3-CHC), 18.08 (CH_3-CHC), 14.13 ($\text{CH}_3-\text{CH}_2\text{O}$).

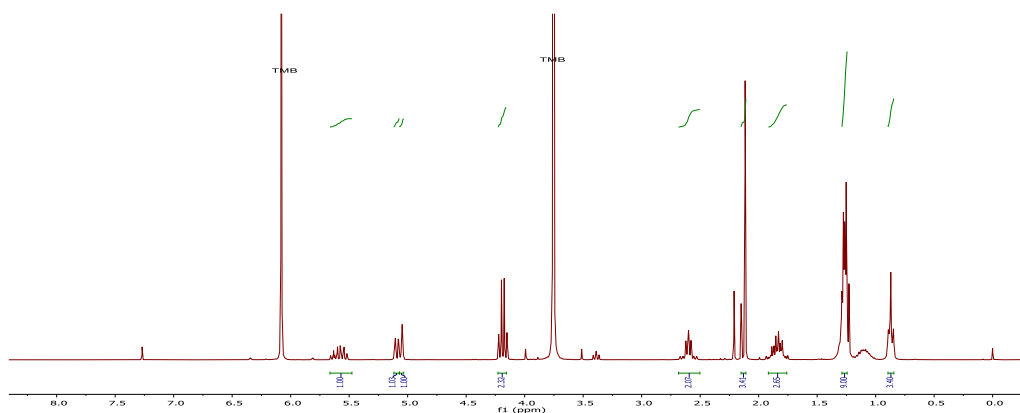
Supplementary Figure S8. ^{13}C NMR spectrum of **3e**.

2.5. Ethyl 2-acetyl-2-allylheptanoate¹ (**3f**)

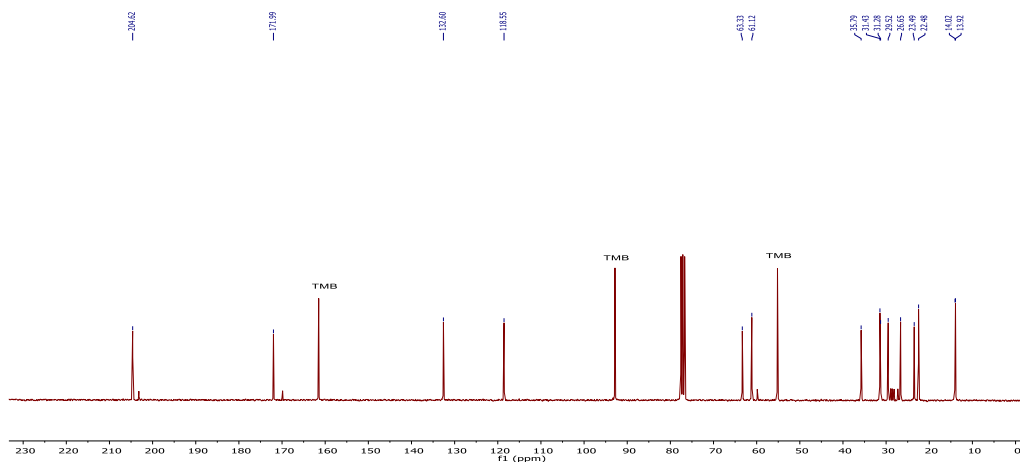
Yield: 20%



¹H NMR (300 MHz, CDCl₃) δ 5.66–5.48 (m, 1H, –CH=), 5.12–5.02 (m, 2H, CH₂=), 4.23–4.16 (q, *J* = 7.1 Hz 2H, CH₃CH₂O), 2.60 (m, *J* = 7.3, 6.0 Hz, 2H, –CH₂CH=), 2.15 (s, 3H, CH₃–CO), 1.92–1.76 (m, 2H, (CH₂)^{hept}CCO), 1.29–1.24 (m, 9H, –(CH₂)₃–CH₃ and CH₃CH₂O), 0.90–0.82 (m, 3H, CH₃^{hept}).

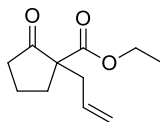
Supplementary Figure S9. ¹H NMR spectrum of **3f**.

¹³C NMR (75 MHz, CDCl₃) δ 204.62 (C=O), 171.99 (C=O), 132.60 (CH=CH₂), 118.55 (CH=CH₂), 63.33 (Cq–CO), 61.12 (CH₃–CH₂O), 35.79 (CH₂CH=CH₂), 31.36 (CH₃–CO), 29.52 (CqCH₂CH₂CH₂CH₂CH₂CH₃), 26.65 (CqCH₂CH₂–CH₂–CH₂CH₃), 23.49 (CqCH₂–CH₂–CH₂CH₂CH₃), 22.48 (CqCH₂CH₂–CH₂–CH₃), 14.02 (CH₃–CH₂O), 13.92 (CH₃–CH₂CH₂CH₂CH₂Cq).

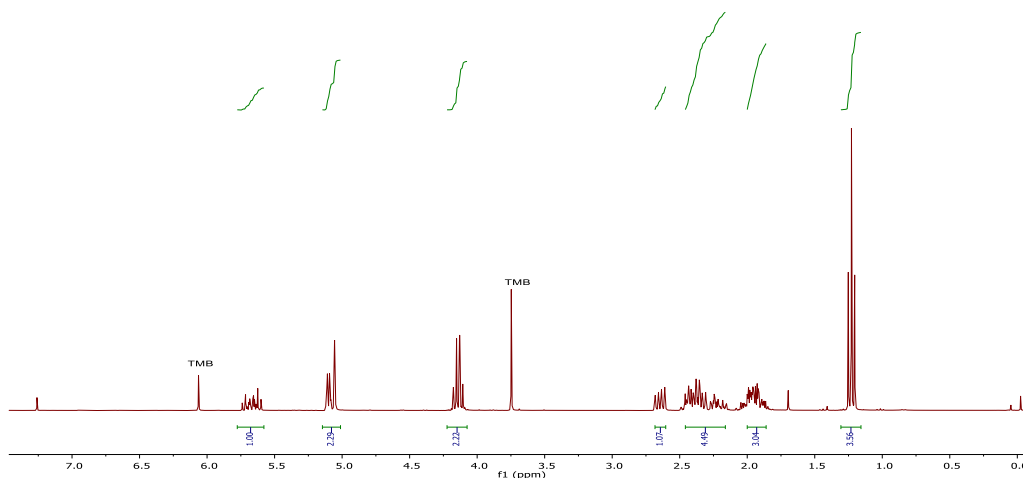
Supplementary Figure S10. ¹³C NMR spectrum of **3f**.

2.6. Ethyl 1-allyl-2-oxocyclopentanecarboxylate¹ (**3g**)

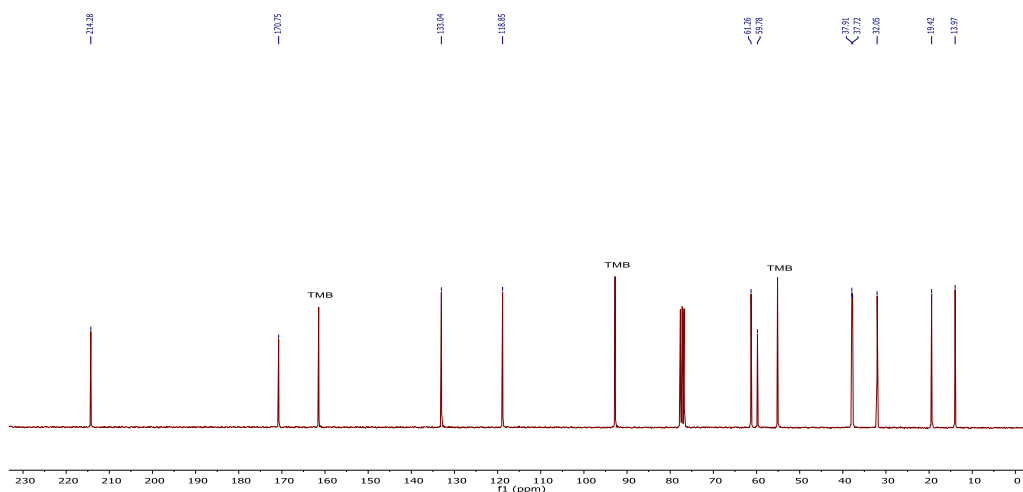
Yield: 64%



¹H NMR (300 MHz, CDCl₃): δ 5.75–5.64 (m, 1H, –CH=), 5.28–5.07 (m, 2H, CH₂=CH), 4.15 (q, *J* = 7.1 Hz, 2H, OCH₂), 2.73–1.80 (m, 8H, CH₂CH₂CH₂ and –CH₂CH=), 1.24 (t, *J* = 7.1 Hz, CH₃CH₂O).

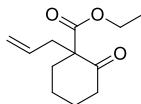
Supplementary Figure S11. ¹H NMR spectrum of **3g**.

¹³C NMR (75 MHz, CDCl₃) δ 214.28 (C=O), 170.75 (C=O), 133.04 (CH₂=CH–CH₂), 118.85 (CH₂=CH–CH₂), 61.26 (CH₃–CH₂O), 59.78 (Cq–CO), 37.91 (=CH–CH₂), 37.72 (CH₂–CO), 32.05 (CH₂–CqCO), 19.42 (CH₂–CH₂CO), 13.97 (CH₃–CH₂O).

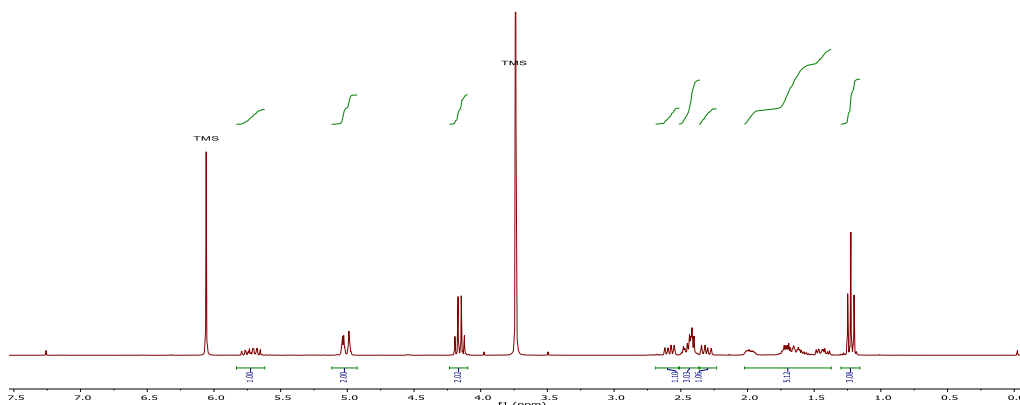
Supplementary Figure S12. ¹³C NMR spectrum of **3g**.

2.7. Ethyl 1-allyl-2-oxocyclohexanecarboxylate¹ (**3h**)

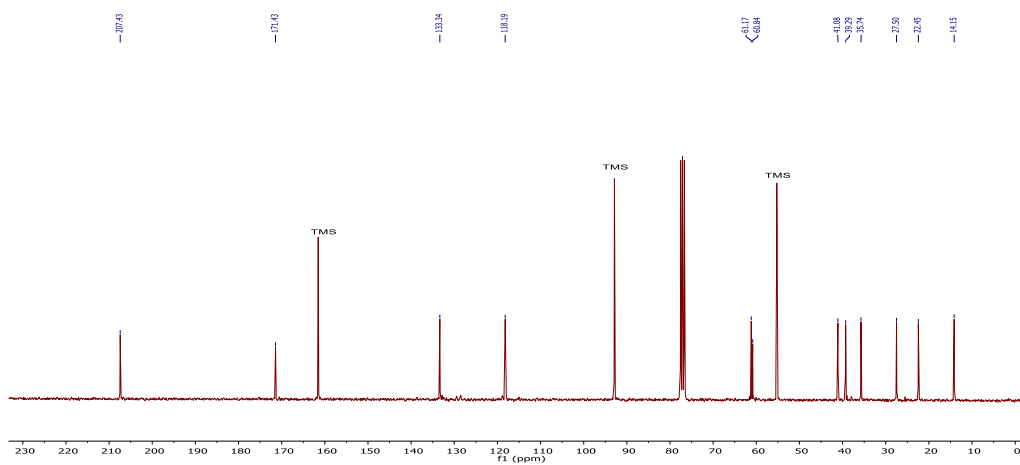
Yield: 75%



¹H NMR (300 MHz, CDCl₃) δ 5.82–5.62 (m, 1H, $-\text{CH}=\text{}$), 5.08–4.94 (m, 2H, $\text{CH}_2=\text{CH}$), 4.16 (q, $J = 7.1$ Hz, 2H, OCH_2), 2.58 (m, $J = 13.9, 7.0, 1.3$ Hz, 1H, $-\text{CH}_2\text{CH}=\text{}$), 2.31 (m, $J = 13.9, 7.8, 1.1$ Hz, 1H, $-\text{CH}_2\text{CH}=\text{}$), 2.49–1.35 (m, 8H, $\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{CO}$), 1.22 (t, $J = 7.1$ Hz, 3H, CH_3).

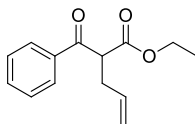
Supplementary Figure S13. ¹H NMR spectrum of **3h**.

¹³C NMR (75 MHz, CDCl₃) δ 207.43 (C=O), 171.43 (C=O), 133.34 ($\text{CH}_2=\text{CH}$), 118.19 ($\text{CH}_2=\text{CH}$), 61.01 ($\text{CH}_3-\text{CH}_2\text{O}$), 60.84 (Cq), 41.08 (CH_2-COCq), 39.29 (CH_2-CqCO), 35.74 ($\text{CH}_2-\text{CH}=\text{}$), 27.50 ($\text{CH}_2-\text{CH}_2\text{CO}$), 22.45 ($\text{CqCH}_2-\text{CH}_2$), 14.15 ($\text{CH}_3-\text{CH}_2\text{O}$).

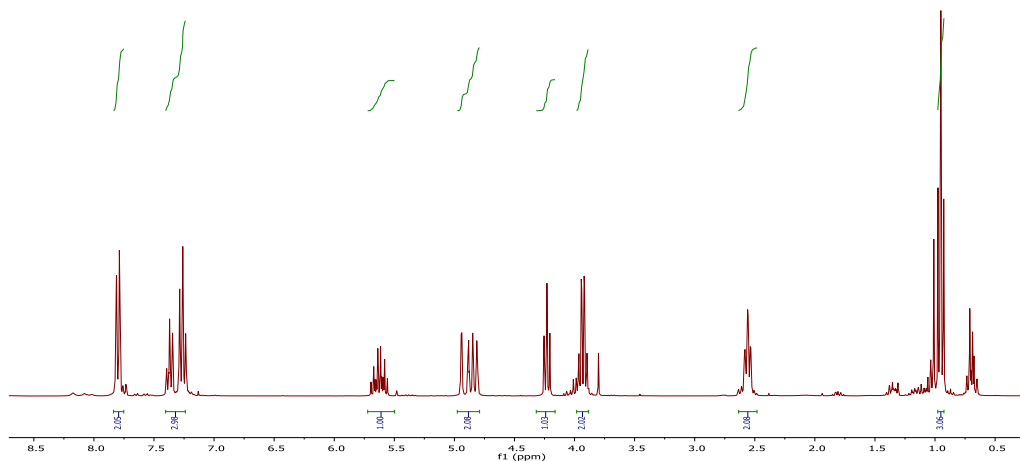
Supplementary Figure S14. ¹³C NMR spectrum of **3h**.

2.9. Ethyl 2-benzoylpent-4-enoate¹ (**3j**)

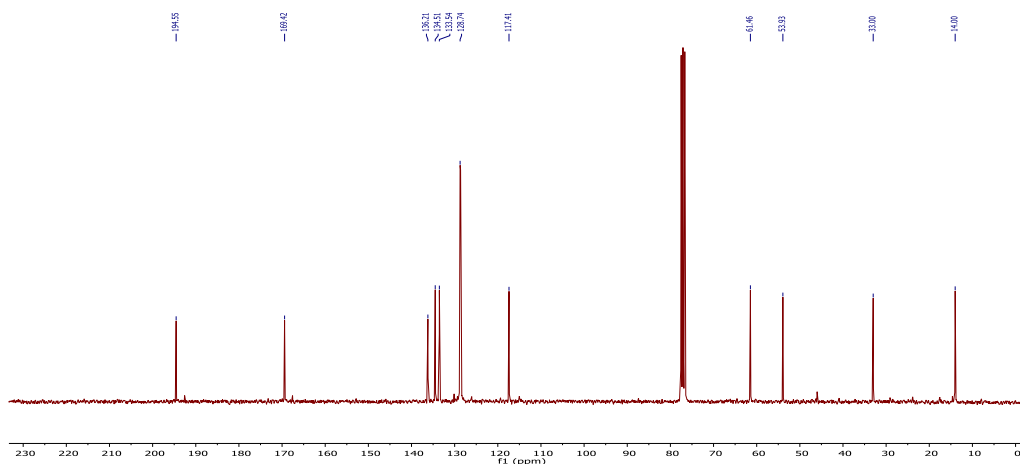
Yield: 31%



¹H NMR (300 MHz, CDCl₃) δ 7.88–7.20 (m, 5H, C₆H₅), 5.63 (m, *J* = 17.0, 10.1, 6.8 Hz, 1H, m, –CH=), 5.02–4.75 (m, 2H, CH₂=), 4.23 (t, *J* = 7.2 Hz, 1H, –CHCO), 3.93 (q, *J* = 7.1, 2H, CH₃CH₂O), 2.56 (m, *J* = 9.5, 6.9, 1.4 Hz, 2H, –CH₂–CH=CH), 1.07–0.90 (t, *J* = 7.1, 3H, CH₃CH₂O).

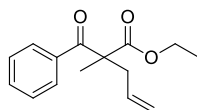
Supplementary Figure S17. ¹H NMR spectrum of **3j**.

¹³C NMR (75 MHz, CDCl₃) δ 194.55 (C=O), 169.42 (C=O), 136.21 (C_q-CO), 134.51 (CH₂=CHCH₂), 133.54 (CH^{para}), 128.74 (CH^{aromatic}), 117.41 (CH₂=CH), 61.46 (CH₃-CH₂O), 53.93 (CH-CO), 33.00 (CH₂-CH=), 14.00 (CH₃-CH₂O).

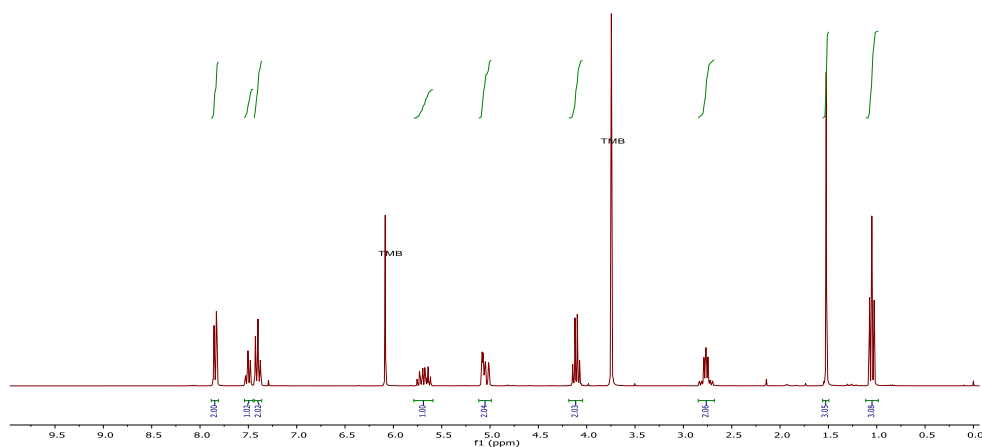
Supplementary Figure S18. ¹³C NMR spectrum of **3j**.

2.10. Ethyl 2-benzoyl-2-methylpent-4-enoate (**3k**)

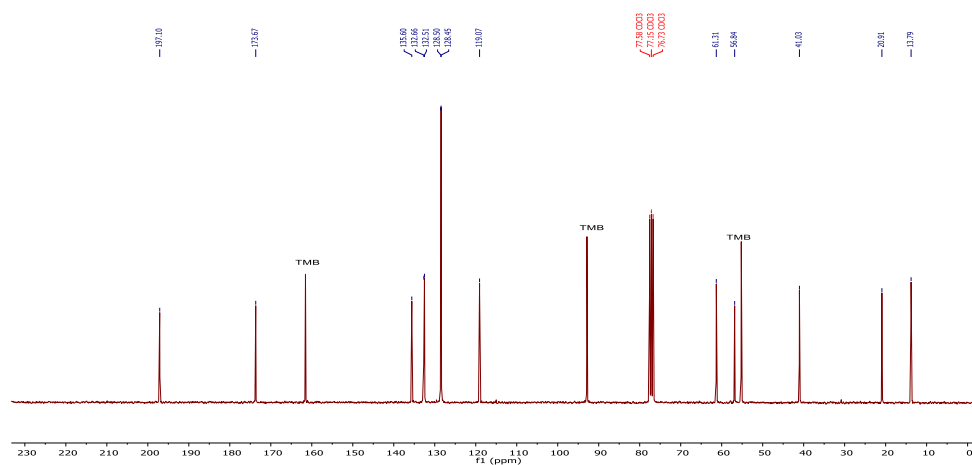
Yield: 62%



^1H NMR (300 MHz, CDCl_3) δ 7.90–7.38 (m, 5H, C_6H_5), 5.83–5.59 (m, 1H, $-\text{CH}=\text{}$), 5.13–4.98 (m, 2H, $\text{CH}_2=\text{}$), 4.11 (q, $J = 7.1$ Hz, 2H, $\text{CH}_3\text{CH}_2\text{O}$), 2.76 (m, $J = 7.1$ Hz, 2H, $\text{CqCH}_2\text{CH}=\text{}$), 1.52 (s, 3H, CH_3CCO), 1.05 (t, $J = 7.1$ Hz, 3H, $\text{CH}_3\text{CH}_2\text{O}$).

Supplementary Figure S19. ^1H NMR spectrum of **3k**.

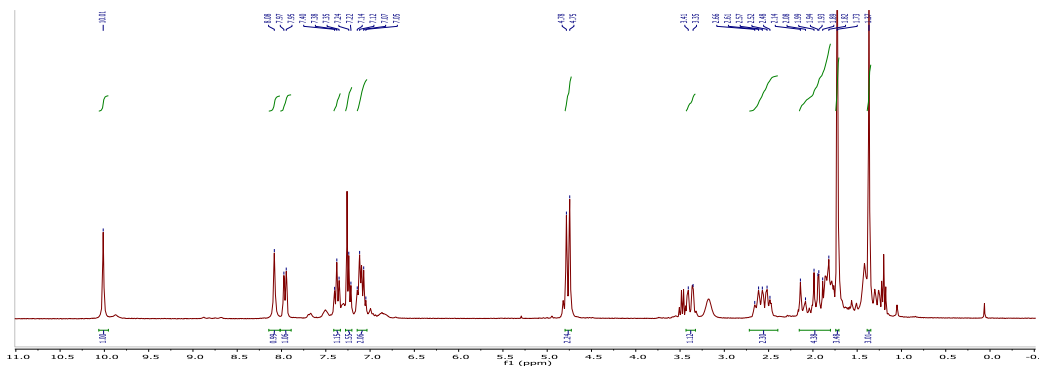
^{13}C NMR (75 MHz, CDCl_3) δ 197.10 ($\text{C}=\text{O}$), 173.67 ($\text{C}=\text{O}$), 135.60 ($\text{Cq}^{\text{aromatic}}$), 132.66 ($\text{CH}^{\text{aromatic}}$), 132.51 ($\text{CH}^{\text{aromatic}}$), 128.50 ($\text{CH}^{\text{aromatic}}$), 128.45 ($\text{CH}^{\text{aromatic}}$), 119.07 ($\text{CH}_2=\text{}$), 61.31 ($\text{CH}_3-\text{CH}_2\text{O}$), 56.84 ($\text{Cq}-\text{COOEt}$), 41.03 ($\text{CH}_2-\text{CH}=\text{CH}_2$), 20.91 (CH_3-CCO), 13.79 ($\text{CH}_3-\text{CH}_2\text{O}$).

Supplementary Figure S20. ^{13}C NMR spectrum of (**3k**).

3. NMR spectra of PdL_nCl₂ complexes

3.1. ¹H NMR of Pd(L1)Cl₂ (C1)

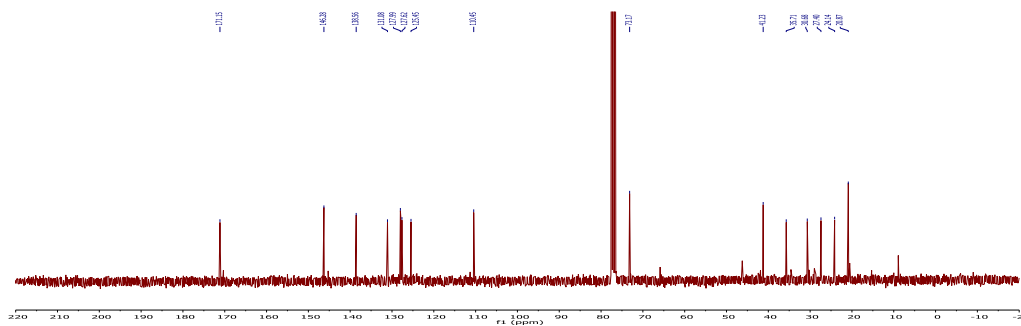
¹H NMR (300 MHz, CDCl₃) δ 10.01 (s, 1H), 8.08 (s, 1H), 7.96 (d, *J* = 8.0 Hz, 1H), 7.38 (t, *J* = 7.6 Hz, 2H), 7.23 (d, *J* = 7.2 Hz, 1H), 7.17–7.02 (m, 3H), 4.76 (d, *J* = 11.2 Hz, 3H), 3.38 (d, *J* = 16.2 Hz, 1H), 2.57 (dt, *J* = 25.8, 11.4 Hz, 3H), 2.18–1.80 (m, 5H), 1.73 (s, 4H), 1.37 (s, 3H).



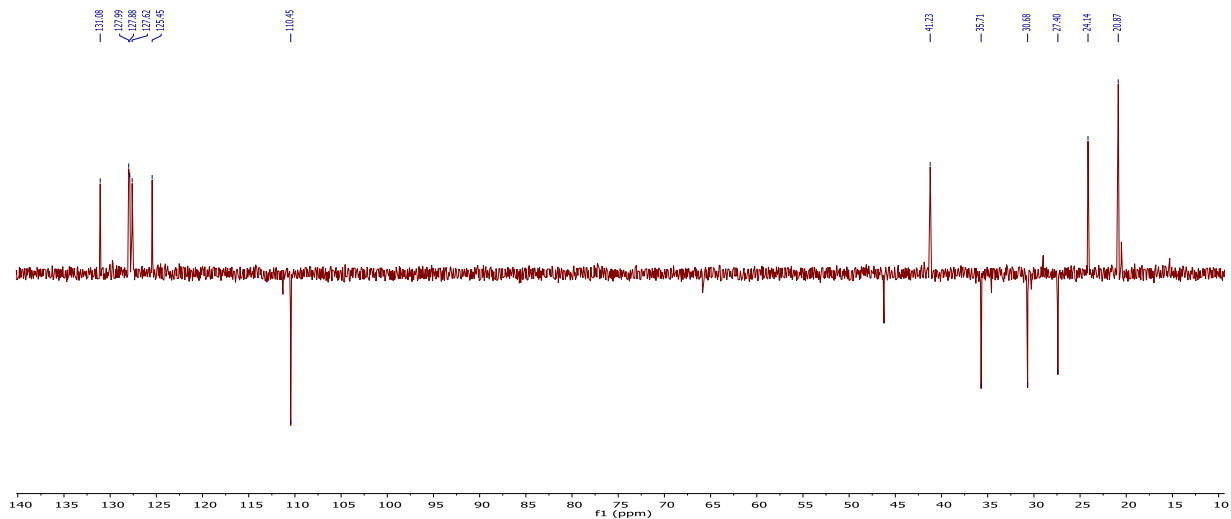
Supplementary Figure S21. ¹H NMR of Pd(L1)Cl₂ in CDCl₃.

3.2. ¹³C NMR of Pd(L1)Cl₂ (C1)

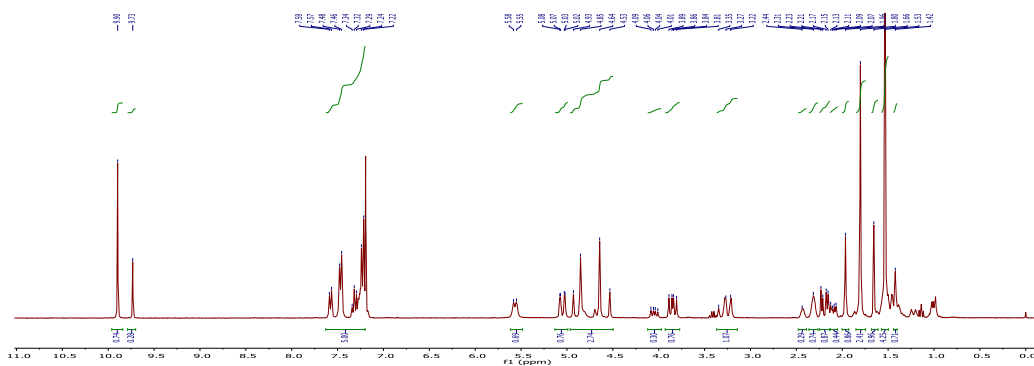
¹³C NMR (75 MHz, CDCl₃) δ 171.15 (C_q=NOH), 146.28 (C_q=CH₂), 138.56 (C_q-C₆H₅), 131.08, 127.99, 127.62, 125.45 (C_{aromatic}), 110.45 (CH₂=), 73.17 (C_q-NH), 41.23 (CH₂-C*-NH), 35.71 (CH₂-CH-C_q), 30.68 (CH₂-C=NOH), 27.40 (CH₂-CH-C_q), 24.14 (CH₃-(C=CH₂)), 20.87 (CH₃-(C-NH)).



Supplementary Figure S22. ¹³C NMR of Pd(L1)Cl₂ in CDCl₃.

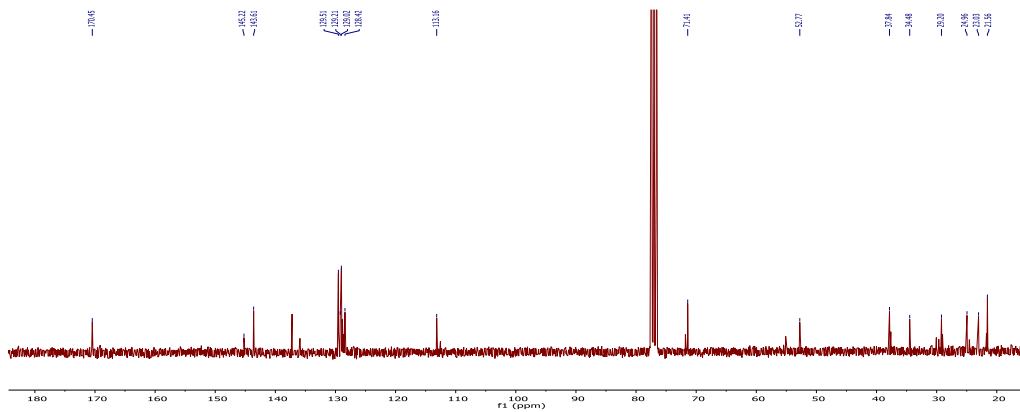
3.3. DEPT 135 NMR of Pd(L1)Cl₂ (C1)Supplementary Figure S23. DEPT 135 NMR of Pd(L1)Cl₂ in CDCl₃.3.4. ¹H NMR of Pd(L2)Cl₂ (C2 and C'2)

¹H NMR (300 MHz, CDCl₃) δ 9.90 (s, 1H), 9.73 (s, 1H), 7.66–7.31 (m, 10H), 5.57 (d, *J* = 9.7 Hz, 1H), 5.04 (dd, *J* = 14.9, 2.4 Hz, 2H), 4.93–4.52 (m, 4H), 4.05 (dd, *J* = 14.4, 8.3 Hz, 1H), 3.85 (dd, *J* = 15.0, 10.0 Hz, 1H), 3.24 (d, *J* = 15.5 Hz, 2H), 2.31 (s, 1H), 2.28–2.13 (m, 1H), 1.96 (s, 1H), 1.80 (s, 6H), 1.65 (s, 1H), 1.44 (s, 1H), 1.53 (s, 3H), 1.42 (m, 1H).

Supplementary Figure S24. ¹H NMR of Pd(L2)Cl₂ in CDCl₃.

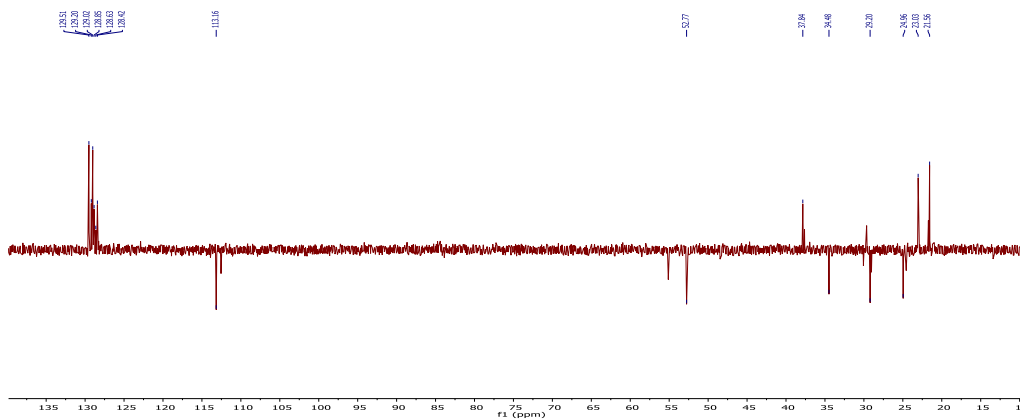
3.5. ^{13}C NMR of $\text{Pd}(\text{L}2)\text{Cl}_2$ (**C2**)

^{13}C NMR (75 MHz, CDCl_3) δ 170.45 (**Cq=N-OH**), 143.61 (**Cq=CH₂**), 137.22 (**Cq-C₆H₅**), 129.51 (**C_{aromatic}**), 129.20, 129.02, 128.85, 113.16 (**CH₂=**), 71.41 (**Cq-NH**), 52.68 (**CH₂-(C₆H₅)**), 37.84 (**CH***), 34.48 (**CH₂-(C*-NH)**), 29.20 (**CH₂-C-NOH**), 24.96 (**CH₂-CH-Cq**), 23.03 (**CH₃-(C=CH₂)**), 21.56 (**CH₃-(C-NH)**).

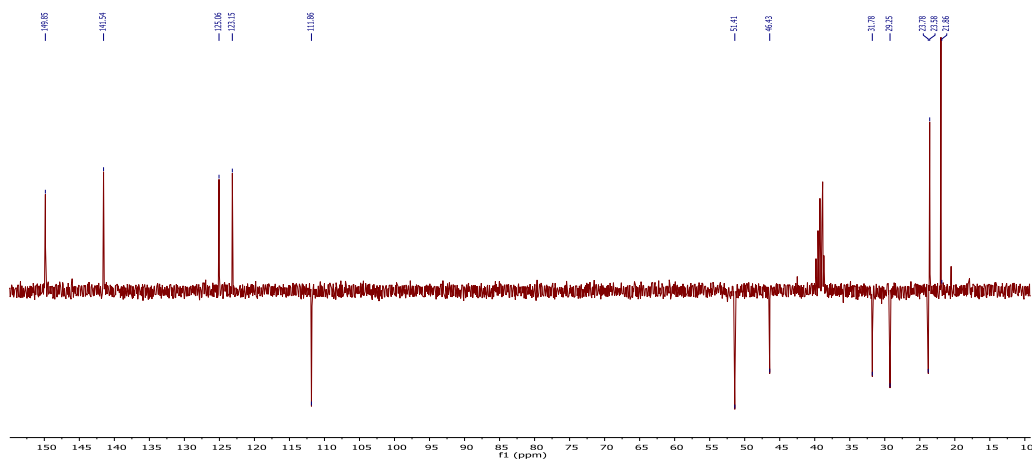


Supplementary Figure S25. ^{13}C NMR of $\text{Pd}(\text{L}2)\text{Cl}_2$ in CDCl_3 .

3.6. DEPT 135 NMR of $\text{Pd}(\text{L}2)\text{Cl}_2$ (**C2**)



Supplementary Figure S26. DEPT 135 NMR of $\text{Pd}(\text{L}2)\text{Cl}_2$ in CDCl_3 .

3.9. DEPT 135 of Pd(L3)Cl₂Supplementary Figure S29. DEPT 135 NMR of PdL₃ in DMSO-*d*₆.

4. Diffraction studies

4.1. Crystal data and structure refinement for complexes C1 and C2

Parameter	C1	C2
<i>Crystal data</i>		
Formula	4(C ₁₆ H ₂₂ Cl ₂ N ₂ OPd)·C ₄ H ₁₀ O	PdCl ₂ C ₁₇ H ₂₄ N ₂ O
Molecular weight	1816.74	449.68
Crystal colour	Orange	Orange
Crystal size (mm)	0.44 × 0.39 × 0.26	0.23 × 0.19 × 0.11
Temperature (K)	100	100
Crystal system	Trigonal	Orthorhombic
Space group	P 3 ₁	P 2 ₁ 2 ₁ 2 ₁
a (Å)	13.1530(4)	8.6502 (3)
b (Å)	13.1530(4)	11.3195 (4)
c (Å)	38.2924(12)	18.1538 (6)
α (°)	90	90
β (°)	90	90
γ (°)	120	90
V (Å ³)	5737.1(4)	1777.55(11)
D _{calc} (mg/m ³); Z	1.578; 3	1.680; 4
F(0 0 0)	2766	912
μ (mm ⁻¹)	1.26	1.35

(continued on next page)

Parameter	C1	C2
<i>Data collection</i>		
Instrument	Bruker APEX-II CCD	Bruker APEX-II CCD
Θ range for data collection (deg)	1.86 to 36.29	2.12 to 33.12
Index ranges	$-21 \leq h \leq 21$	$-13 \leq h \leq 11$
	$-21 \leq k \leq 21$	$-17 \leq k \leq 17$
	$-63 \leq l \leq 63$	$-22 \leq l \leq 27$
Number of measured reflections	198742	45344
Number of independent reflections	36836	6736
Number of reflections with $I > 2\sigma(I)$	35926	6627
R_{int}	0.043	0.038
<i>Refinement</i>		
Refinement method	Full matrix least square on F^2	Full matrix least square on F^2
$R[F^2 > 2\sigma(F^2)]$	0.030	0.018
$wR(F^2)$	0.065	0.043
S	1.09	1.04
Data/restraints/parameters	36836/7/861	6736/0/218
$\Delta\rho_{\text{max}}/\Delta\rho_{\text{min}} [e \text{ \AA}^{-3}]$	1.25/−0.60	0.79/−0.60
Absolute structure Flack parameter	−0.007(4)	−0.011(8)

4.2. Selected bond lengths d and bond angles ω in complex **C1**

Bond	d (Å)
Pd(1)–Cl(1)	2.288(1)
Pd(1)–Cl(2)	2.307(8)
Pd(1)–N(1)	2.071(3)
Pd(1)–N(2)	1.981(4)
N(1)–C(1)	1.458(5)
N(1)–C(7)	1.543(6)
N(2)–O(1)	1.382(6)
N(2)–C(12)	1.289(4)

Angle	ω (deg)
Cl(1)–Pd(1)–Cl(2)	91.80(4)
N(2)–Pd(1)–Cl(2)	91.43(1)
N(1)–Pd(1)–Cl(1)	96.50(8)
N(1)–Pd(1)–N(2)	80.1(1)
O(1)–N(2)–Pd(1)	123.4(3)
O(1)–N(2)–C(12)	116.2(3)
Pd(1)–N(1)–C(7)	109.3(2)
Pd(1)–N(1)–C(1)	121.4(2)
N(2)–C(12)–C(7)	115.5(3)
N(2)–C(12)–C(11)	123.7(3)
C(1)–N(1)–C(7)	112.2(3)
N(1)–C(1)–C(6)	120.1(3)

4.3. Selected bond lengths d and angles ω in Complex **C2**

Angle	ω (deg)
Cl(2)–Pd(1)–Cl(1)	94.56(2)
N(1)–Pd(1)–Cl(1)	92.20(4)
N(1)–Pd(1)–Cl(2)	172.79(4)
N(1)–Pd(1)–N(2)	80.60(6)
N(2)–Pd(1)–Cl(1)	172.79(4)
N(2)–Pd(1)–Cl(2)	92.64(4)
O(1)–N(1)–Pd(1)	124.1(1)
C(1)–N(1)–Pd(1)	119.3(1)
C(1)–N(1)–O(1)	116.6(1)

Bond	d (Å)
Pd(1)–Cl(1)	2.3187(5)
Pd(1)–Cl(2)	2.2886(5)
Pd(1)–N(1)	1.990(1)
Pd(1)–N(2)	2.046(1)
N(1)–O(1)	1.383(2)
N(1)–C(1)	1.285(2)
N(2)–C(6)	1.530(2)
N(2)–C(11)	1.512(2)