# Synthesis of new 3-substituted-2*H*-1,2naphthothiazin-4(3*H*)-one 1,1-dioxides via directed ortho-metalation reaction

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Abstract – The reaction of compounds 6a-i, readily available from  $\alpha$ -amino acids, with an excess of lithium diisopropylamide, leads to new 3-substituted-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1-dioxides 7a-i, with yields ranging between 21 and 70%. The key steps are: the naphthylsulfonyl ortho-deprotonation based on the directed ortho-metalation reaction followed by a regiospecific intramolecular cyclisation reaction. Lithiation–deuteration experiments carried out on the naphthylsulfonamides 8 and 9 using *n*-BuLi and LDA demonstrated the regioselectivity of the deprotonation of the H-3 over the H-1 one of the naphthalene ring. *To cite this article: Y. Kacem et al., C. R. Chimie 5 (2002) 611–621* © 2002 Académie des sciences / Éditions scientifiques et médicales Elsevier SAS

#### regiospecificity / cyclisation / metalation / deprotonation / deuteration

Résumé – Synthèse de nouveaux 1,1-dioxydes de 2*H*-1,2-naphtothiazin-4(3*H*)-one 3-substitués via la réaction de métallation ortho-dirigée. La réaction des composés 6a–i, facilement obtenus à partir acides  $\alpha$ -aminés, en présence d'un excès de diisopropylamide de lithium, conduit à de nouveaux 1,1-dioxydes de 2*H*-1,2-naphtothiazin-4(3*H*)-one 3-substituées 7a–i, avec des rendements entre 21 et 70%. Les étapes clés sont : la déprotonation ortho du naphtylsulfonyle, basée sur la réaction de métallation ortho dirigée, suivie par une réaction de cyclisation intramoléculaire régiospécifique. Des expériences de lithiation– deutération menées sur les naphthylsulfonamides secondaires et tertiaires 8 et 9 en présence de *n*-BuLi et de LDA ont démontré le caractère régiospécifique de la déprotonation du proton H-3 sur le proton H-1 du cycle naphthalène. *Pour citer cet article : Y. Kacem et al., C. R. Chimie 5 (2002) 611–621* © 2002 Académie des sciences / Éditions scientifiques et médicales Elsevier SAS

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### 1. Introduction

Heterocycles involving an arylsulfamido moiety have been reported to possess a variety of interesting biological activities [1–5]. For example, 1,2benzothiazines with basic side chains [6] are claimed to be diuretic agents. Antithrombotic and lipidregulating properties are observed for 1,2benzothiazine-3-carboxamides [7]. Antibacterial activity was found for several penicillin derivatives containing the 1,2-benzothiazinyl fragment [8]. Many reports have referred to the anti-inflammatory analge-

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Fig. 1. Compounds 1-4.

sic and antipyretic activities found for a variety of 1,2-benzothiazines [9]. An initial report by Lombardino et al. [10] indicated that potent anti-inflammatory activity was present in a series of 4-hydroxy-2H-1,2-benzothiazin-3-carboxamide 1,1-dioxides **1** (Fig. 1).

Owing to these important biological properties of the benzo- and the heterocyclic ring-fused 1,2thiazines, many publications have presented interesting methods of their synthesis [11–20]. Meanwhile a few synthetic approaches to 1,2-naphtothiazine derivatives have been developed in the literature. Kaufmann and Zobel [21] first prepared 2,3-dihydro-3-oxonaphtho-[1,8-d,e]-1,2-thiazine 1,1-dioxides **2**. More recently, this compound was prepared by Lombardino according to a short process [22]. Both Trummlitz [23] and Steiner [24] have prepared independently naphtho[2,1e]-1,2-thiazine **3** analogues of **1**. Trummlitz and co-workers [23] also reported the synthesis of *N*-substituted 2H-naphtho[2,1-c]-1,2-thiazin-4(3*H*)-one 1,1-dioxides **4** by ring expansion of 3-bromomethylnaphth-[2,1-d]-isothiazole 1,1-dioxides.

As part of our continued efforts to develop synthetically useful anionic aromatic reactions for the synthesis of biologically active compounds [25], we report a general route based on the well known directed orthometalation reaction [20] in our synthetic approach to 3-substituted-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1dioxides **7a–i** (Fig. 2). These heterocycles are analogues of the 1,2-benzothiazines derivatives that were required for biological activities evaluations and as starting materials to prepare new drugs.



Fig. 2. Synthesis of compounds 5a-i, 6a-i, and 7a-i.

### 2. Results and discussion

The aromatic directed ortho-metalation reaction has been developed into a broadly useful protocol for the regioselective construction of polysubstituted aromatics [26–29] and used in the efficient synthesis of several heterocyclic ring systems and bioactive compounds [30].

The process reported herein (Fig. 2), possibly driven by the Complex Induced Proximity Effect (CIPE) [31], constitutes a mild method for the LDA-mediated regiospecific conversion of *N*-(2-naphthylsulfonyl) amino amides **6a–i** [32–36] readily available from commercial racemic or optically pure (*S*)- $\alpha$ -amino acids into a new 2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1dioxides **7a–i**. According to this process, it was possible to change substitution patterns at the C-3 position of compounds **7a–i**.

In order to select the optimal reaction conditions for the synthesis of compounds **7a-i**, we have carefully examined the ortho-metalation conditions carried out on aminoamides **6a** or **6f** by changing the base (nature and number of equivalents) and the temperature.

The results shown in Table 1 indicate the requirement of an excess of LDA for efficient ortholithiations and to improve the yields (entries 1 to 5). In addition of the possible  $\alpha$ - and  $\alpha$ '-amide carbanion formation, the excess of LDA also serves to displace the equilibrium between the complex formed by the association of the organolithium reagent and the Lewis basic functionality of the substrate and the coordinated ortho-lithiated species in favour of the latter complex [29,31].

The use of other bases, such as LiHMDS, *sec*-BuLi and *n*-BuLi, do not ameliorate the yield of products. With *sec*-BuLi, the cyclisation proceeds very poorly (entry 8), yielding a complex mixture of products. Moreover, either at 0 °C or at -78 °C, the exposure of (*S*)-*N*,*N*-diisopropyl-3-methyl-2-(*N*-2-naphthylsulfonylamino)butanamide **6a** to *n*-BuLi gave the 2-methyl-

Table 1. Yields of products 7a and 7f (optimisition experiments).

Entry	Substrate	BM (equivalent)	Yield (%) <sup>a</sup>
1	6a	LDA (2)	38
2	6a	LDA (3)	47
3	6a	LDA (4)	56
4	6f	LDA (2)	61
5	6f	LDA (3)	70
6	6f	LDA (3) <sup>b</sup>	43
7	6f	LiHMDS (3)	64
8	6f	sec-BuLi (3)	33

<sup>a</sup> Yields correspond to isolated chromatographed materials. <sup>b</sup> Reaction carried out at -78 °C (30 min) then room temperature (2 h).

3-(*N*-2-naphthylsulfonylamino)octan-4-one as a major product along with the starting material **6a**. The ketone results from nucleophilic addition of *n*-BuLi to **6a** before the directed ortho-lithiation reaction. In addition, we have found out that low temperature (-78 °C) decreases significantly the yield of the cyclisation reaction when LDA was used (compare entry 5 with entry 6).

Attempts of these different experimental conditions summarised in Table 1 led to the selection of the following procedure: treatment of compounds **6a–i** with [LDA (four equivalents for N–H derivatives and three equivalents for N–Me derivatives)/THF /–5 to 25 °C over 1 h] gave the corresponding 2*H*-1,2naphthothiazin-4(3*H*)-one 1,1-dioxides **7a–i**. However, the excess of LDA has racemised heterocycles prepared from enantiomerically pure intermediates **6a,c–h**. Yields range between 21 and 70% (Table 2).

Theoretically, ortho-lithiation of *N*-(2-naphthylsulfonyl)amino amides **6a–i** can occur on carbon-1 and carbon-3 of the naphthalene ring. Characterisation of the reaction product proves the regiospecific cyclisation of substrates **5** to provide exclusively 2H-1,2naphthothiazin-4(3*H*)-one 1,1-dioxides **7a-i**. To further prove the selectivity of H-3 deprotonation on expense of H-1 proton, lithiation–deuteration experiments with secondary and tertiary 2-naphthylsulfonamides **8** and **9** were undertaken (Fig. 3).

Thus, treatment of N-methyl-2-naphthylsulfonamide 8 with *n*-BuLi (2.2 equiv), followed by quench with CD<sub>3</sub>OD resulted in selective deuterium incorporation at carbon-3 (RMN <sup>1</sup>H). Likewise, N,N-diethyl-2naphthylsulfonamide 9 when subjected to similar n-BuLi (1.1 equiv) metalation-CD<sub>3</sub>OD quench conditions, afforded only the ortho-deuterated product at carbon-3 (Table 3). As metalation regioselectivity is influenced by variation of metalating agent [37], substrates 8 and 9 were metalated with [LDA (2.2 equiv for 8 and 1.1 equiv for 9)/THF/-5 °C to 25 °C]. Aliquots were quenched at 0 °C and at room temperature during the course of the reaction by addition to CD<sub>3</sub>OD. Deuterium incorporation obtained in these experiments was the same as that obtained when *n*-BuLi was used.

Both results of ortho-lithiation–cyclisation with compounds **5** and ortho-lithiation–deuteration with compounds **8** and **9** demonstrate that the orientation of the sulfonamide group with respect to the proton removed during the ortho-lithiation significantly affects the reaction course [38]. These results lead also to the suggestion of a naphthalenesulfonyl-base complex (Fig. 4), which places the LDA in the proximity of H-3 proton so as to exert a regio-control by a CIPE process.

Entry	Substrate 6	Product 7	Yield (%) <sup>a</sup>
1	$ \begin{array}{c}                                     $		56
2	$Me \underbrace{N(iPr)_2}_{HN_S} \underbrace{N(iPr)_2}_{\mathbf{6b} O_2}$	$\mathbf{\mathbf{B}}_{\mathbf{b}} \mathbf{\mathbf{C}}_{\mathbf{b}} \mathbf{\mathbf{C}}_{\mathbf{b}} \mathbf{\mathbf{C}}_{\mathbf{b}}$	40
3	$MeS \xrightarrow{N(iPr)_2}_{HN} \underbrace{N(iPr)_2}_{6c O_2}$	$\bigcup_{\mathbf{7c}} SMe$	52
4	Ph $(iPr)_2$ $HN_S$ $6d O_2$	$\begin{array}{c} & & \\ & & \\ & & \\ & & \\ & & 7d \\ & & O_2 \end{array}$	48
5	$Me \xrightarrow{HN}_{S} \xrightarrow{N(iPr)_2}$	$7e$ $O_2$ Me Et	50 <sup>b</sup>
6	$\mathbf{MeN}_{S} \mathbf{MeN}_{S}$	$7f O_2$	70
7	$\begin{array}{c} \begin{array}{c} & & \\ $	7g $O$ $Ph7g O_2$	62
8	$ \begin{array}{c}                                     $	$\overbrace{\mathbf{7h}}^{O} \underset{O_2}{\overset{O}{\overset{N}}}$	46
9	$ \begin{array}{c}                                     $	$\vec{i}$	21

Table 2. Synthesis of 3-substituted-2H-1,2-naphthothiazin-4(3H)-one 1,1-dioxides 7a-i.

<sup>a</sup> Yields correspond to isolated chromatographed materials.

<sup>b</sup> Two diastereomers were formed in a 3:1 ratio.

### 3. Conclusion

In summary, a systematic study has provided optimised conditions for the synthesis of new 2H-1,2naphthothiazin-4(3H)-one 1,1-dioxides analogues of the 1,2-benzothiazine class. The synthetic approach applied to elaborate these heterocycles was based on the directed ortho-metalation procedure. This reaction has significant advantages over the more traditional alkoxide-mediated ring expansion of N-acyl-1,2arenethiazole 1,1-dioxides derivatives [13, 24], since it constitutes a general, convenient and regiospecific method for the synthesis of substituted 1,2naphthothiazines. The selectivity of ortho-lithiation



Fig. 3. Lithiation-deuteration experiments with secondary and tertiary 2-naphthylsulfonamides 8 and 9, giving 10 and 11.



Fig. 4. Chemical route from 6 to 7, through a naphthalenesulfonyl-base complex, which places the LDA in the proximity of H-3 proton and so exerts regio-control by a CIPE process.

Table 3. Yields of products 10 and 11 (deuteration experiments).

Product	RLi	Yield (%)
10	<i>n</i> -BuLi	79
10	LDA	38
11	<i>n</i> -BuLi	87
11	LDA	42

Yields correspond to isolated chromatographed deuterated products.

reactions of **6**, **8** and **9** is reasonably attributed to a favourable arrangement during the transfer of the proton to the lithiating reagent within a complex of sulfonamide with the organolithium reagent.

The continuous pharmaceutical interest in the arylsulfonamide compounds [25, 39–40] may justify further exploration of these results in the pharmacological field.

### 4. Experimental section

All reactions were carried out under argon in dried glassware, using syringe-septum cap techniques. Solvents were purified and dried by standard methods. Melting points were determined on a Büchi SMP-20 capillary apparatus and are uncorrected. TLC was carried out on Merck 60F-254 precoated silica gel plates

(0.25 mm) and column chromatography was performed with Merck silica gel (70-230 mesh). NMR spectra were recorded on a Bruker 300 spectrometer (<sup>1</sup>H at 300 MHz and <sup>13</sup>C at 75 MHz) with CDCl<sub>3</sub> as solvent and TMS as internal standard reference. IR-TF spectra were recorded on a BIORAD FTS-6000 spectrometer. Mass spectra (EI) were recorded on a Hewlett Packard 5792 apparatus coupled to a GC HP-5890 chromatograph (column: 5% diphenyl, 95% dimethylpolysiloxane.  $25 \text{ m} \times 0.25 \text{ mm}$ ID. 0.35 µm film). Elemental analyses were carried out by the 'Service de microanalyse' of the University Paris-6. Optical rotations were measured at 21 °C on a Perkin-Elmer 241 polarimeter. n-BuLi and sec-BuLi were purchased from Acros as solutions in hexane stored in a resealable container, and titrated periodically against sec-butanol [41].

## 4.1. *N*-(2-Naphthylsulfonyl)-2-amino acids 5a-e and 5h,i. General procedure

To a solution of an  $\alpha$ -amino acid (20 mmol) in aqueous NaOH (2M, 10 ml, 20 mmol) at 0 °C was added 2-naphthylsulfonyl chloride (21 mmol, 1.05 equiv), followed by EtN(*i*-Pr)<sub>2</sub> (22 mmol, 1.1 equiv) and acetone (10 ml). The mixture was stirred at room temperature for 12 h. The reaction mixture was washed with Et<sub>2</sub>O (2 × 50 ml) and the combined washings extracted with aqueous NaOH (2M, 10 ml). The combined basic aqueous layers were cooled to 0 °C and acidified (pH 1) by the addition of concentrated HCl. The mixture was extracted with EtOAc  $(3 \times 50 \text{ ml})$ , dried over MgSO<sub>4</sub>, filtered and concentrated under reduced pressure to give *N*-(2-naphthylsulfonyl)-2-amino acids, which were purified by crystallisation.

## 4.2. (S)-3-Methyl-2-(N-2-naphthylsulfonylamino)butanoic acid: 5a

Yield 78%. mp 103–105 °C.  $[\alpha]_D = -19.2$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.88$  (d, 3H, J = 6.8 Hz), 0.97 (d, 3H, J = 6.8 Hz), 2.15 (m, 1H), 3.79 (dd, 1H, J = 4.6 Hz and 9.8 Hz), 5.27 (d, 1H, J = 9.8 Hz), 7.54–7.63 (m,2H), 7.77–7.94 (m, 4H), 8.35 (s, 1H), 9.45 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 16.64$ , 19.17, 28.23, 63.59, 123.94, 126.36, 127.32, 129.37, 130.56, 130.29, 130.96, 133.58, 133.80, 137.28, 173.55.

#### 4.3. (±)-2-(N-2-Naphthylsulfonylamino)propanoic acid: 5b

Yield 83%. mp 151–105 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.22$  (d, 3H, J = 7.4 Hz), 3.83 (qd, 1H, J = 7.4 Hz and J = 6.5 Hz), 5.56 (d, 1H, J = 6.5 Hz), 7.60–7.71 (m, 2H), 7.80–7.96 (m, 4H), 8.40 (s, 1H), 9.9 (br,1H). – <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 38.36$ , 57.03, 122.63, 126.59, 128.67, 129.11, 129.87, 131.03, 131.65, 134.78, 135.13, 138.43, 172.98.

### 4.4. (S)-3-Methylsulfanyl-2-(N-2-naphthylsulfonylamino) propanoic acid: 5c

Yield 77%. mp 93–94 °C.  $[\alpha]_{\rm D} = -15.6$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 3.28$  (m, 2H), 3.75 (s, 3H), 4.23 (m, 1H), 6.32 (d, 1H, J = 9.8 Hz), 7.59–7.68 (m, 2H), 7.73–7.90 (m, 4H), 8.38 (s, 1H), 9.60 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 18.76$ , 41.46, 55.79, 124.15, 126.36, 128.65, 129.32, 130.56, 131.08, 131.98, 133.40, 134.74, 137.36, 172.36.

## 4.5. (S)-3-Phenyl-2-(N-2-naphthylsulfonylamino) propanoic acid: 5d

Yield 84%. mp 133–134 °C.  $[\alpha]_D = -43.2$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 3.15$  (m, 2H), 3.98 (m, 1H), 5.59 (d, 1H, J = 10.0 Hz), 7.66–7.74 (m, 7H), 7.86–8.04 (m, 4H), 8.46 (s, 1H), 10.16 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 42.38$ , 57.17, 124.69, 125.61, 126.75, 127.14, 127.61, 128.07, 128.66, 129.14, 129.86, 131.55, 132.18, 135.73, 138.26, 139.10, 175.44.

## 4.6. (25, 35)-3-Methyl-2-(N-2-naphthylsulfonylamino) pentanoic acid: 5e

Yield 70%. mp 111–112 °C.  $[\alpha]_{\rm D} = -8.4$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.92$  (d,

3H, J = 5.4 Hz), 1.18 (t, 3H, J = 3.5 Hz), 1.25–1.39 (m, 2H), 2.48–2.55 (m, 1H), 4.12–4.20 (dd, 1H, J = 3.3 and J = 10.0 Hz), 5.25–5.30 (d, 1H, J = 10.0 Hz), 7.58–7.69 (m, 2H), 7.81–7.96 (m, 4H) 8.28 (s, 1H), 9.6 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 12.03$ , 14.52, 25.71, 34.36, 62.41, 122.69, 125.33, 127.74, 128.72, 129.13, 130.48, 131.11, 133.67, 134.10, 137.15, 173.19.

#### 4.7. (S)-N-(2-Naphthylsulfonyl)pipecolinic acid: 5h

Yield 75%. mp 131–132 °C.  $[a]_D = -35.0$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.9-1.1$  (m, 1H), 1.4–1.6 (m, 4H), 2.5–2.7 (m, 2H), 3.39 (dt, 1H, J = 3.5 Hz and J = 10.8 Hz), 5.34 (t, 1H, J = 3.5 Hz), 7.53–7.64 (m, 2H), 7.79–7.98 (m, 4H), 8.42 (s, 1H), 9.95 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 20.17$ , 24.15, 24.36, 45.59, 61.82, 123.63, 126.28, 128.93, 129.71, 130.79, 131.09, 131.69, 133.73, 134.44, 137.48, 174.42.

#### 4.8. (±)-N-(2-Naphthylsulfonyl)proline: 5i

Yield 74%. mp 128–130 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.76-1.82$  (m, 2H), 2.18–2.32 (m, 2H), 3.34–3.52 (m, 2H), 4.30 (dt, 1H, J = 4.1 Hz and J = 10.7 Hz), 7.61–7.72 (m, 2H), 7.83–8.02 (m, 4H), 8.28 (s, 1H), 9.7 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 24.96$ , 31.18, 47.96, 59.42, 123.80, 126.65, 128.73, 129.51, 130.39, 130.36, 131.31, 133.49, 134.33, 137.47, 172.52.

## 4.9. N-Methyl-N-(2-naphthylsulfonyl)-2-amino acids 5f,g. General procedure

*N*-(2-naphthylsulfonyl)-2-amino acid was suspended in 150 ml of toluene, and paraformaldehyde (2 g) and *p*-toluenesulfonic acid (200 mg) were added. The mixture was refluxed for 30 min with azeotropic water removal. The solution was then cooled, washed with 1 N aqueous NaHCO<sub>3</sub> ( $2 \times 50$  ml) and dried over MgSO<sub>4</sub>. Concentration in vacuo gave the corresponding *N*-(2-naphthylsulfonyl)oxazolidinone, without further purification was dissolved in 20 ml of CHCl<sub>3</sub> and 20 ml of trifluoroacetic acid and triethylsilane (30 mmol) were added. The solution was stirred at room temperature for 24 h; subsequent concentration in vacuo and purification by crystallisation gave pure *N*-methyl-*N*-(2-naphthylsulfonyl)-2-amino acid.

#### 4.10. (S)-3-Methyl-2-(N-methyl-N-2-naphthylsulfonylamino)butanoic acid: 5f

Yield 81%. mp 116–117 °C.  $[\alpha]_D = -25.1$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.89$  (d, 3H, J = 7.2 Hz), 1.13 (d, 3H, J = 7.2 Hz), 2.48 (m, 1H), 2.52 (s, 3H), 4.36 (d, 1H, J = 3.3 Hz), 7.54–7.64

(m,2H), 7.81–7.97 (m, 4H), 8.37 (s, 1H), 10.5 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 17.19, 19.38, 27.86, 38.60, 63.19, 123.78, 126.46, 128.73, 128.98, 130.20, 130.52, 131.25, 133.49, 134.35, 136.88, 172.23.

#### 4.11. (S)-3-Phenyl-2-(N-methyl-N-2-naphthylsulfonylamino)propanoic acid: 5g

Yield 95%. mp 139–141 °C.  $[\alpha]_{\rm D} = -48.5$  (c1, MeOH). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 2.20$  (s, 3H), 3.19 (d, 2H, J = 3.9 Hz), 4.24 (t, 1H, J = 3.9 Hz), 7.37–7.58 (m, 7H), 7.78–7.94 (m, 4H), 8.27 (s, 1H), 9.40 (br, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 37.56$ , 41.61, 56.31, 128.53, 128.94, 129.09, 129.58, 130.01, 130.75, 131.29, 131.96, 133.01, 133.87; 134.25, 134.86, 135.75, 136.93, 173.15.

## 4.12. *N*,*N*-Diisopropyl-2-amino amides 6a–i. General procedure

To a stirred solution of N-(2-naphthylsulfonyl)-2amino acid or N-methyl-N-(2-naphthylsulfonyl)-2amino acid (10 mmol) in dry benzene (70 ml), was added at room temperature SOCl<sub>2</sub> (12 mmol, 1.2 equiv). The mixture was then refluxed for 3 h, cooled and concentrated to oil. The oil was dissolved in CH<sub>2</sub>Cl<sub>2</sub> and reconcentrated two times. The corresponding acid chloride obtained, without further purification, was dissolved in 50 ml of CH2Cl2 and added to a stirred solution of diisopropylamine (11 mmol) and triethylamine (11 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 ml). The mixture was stirred at 0 °C for 2 h and then poured into 100 ml of a diluted HCl solution. The N,Ndiisopropyl-2-amino amide was extracted with CH<sub>2</sub>Cl<sub>2</sub> and the organic phase was washed with saturated NaCl solution, dried over MgSO<sub>4</sub>, filtered and concentrated. The residue was purified by silica gel chromatography [SiO<sub>2</sub>; (EtOAc:hexane)] to afford the N,Ndiisopropyl-2-amino amide.

#### 4.13. (S)-N,N-Diisopropyl-3-methyl-2-(N-2-naphthylsulfonylamino)butanamide: 6a

Yield 84%. mp 118–121 °C.  $[\alpha]_{\rm D} = -31.9$  (c1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.85$  (d, 3H, J = 7.0 Hz), 1.11 (d, 3H, J = 7.0 Hz), 1.14–1.29 (m, 12H), 2.88 (m, 1H), 3.05–3.13 (m, 2H), 4.36 (dd, 1H, J = 4.1 Hz and 10.18 Hz), 5.50 (d, 1H, J = 10.1 Hz), 7.61–7.71 (m, 2H), 7.82–8.9 (m, 4H), 8.44 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.05$ , 12.27, 12.86, 13.51, 16.71, 19.20, 29.02, 40.63, 41.37, 67.84, 123.86, 127.09, 128.73, 129.41, 130.17, 130.52, 131.42, 133.60, 134.19, 137.33, 168.61. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (390.54): calcd C 64.58, H 7.74; found C 64.47, H 7.81.

#### 4.14. (±)-*N*,*N*-Diisopropyl-2-(*N*-2-naphthylsulfonylamino)propanamide: 6b

Yield 78%. mp 113–114 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.0-1.21$  (m, 12H), 1.28 (d, 3H, J = 7.2 Hz), 3.11–3.25 (m, 2H), 3.84 (qd, 1H, J = 7.2 Hz and J = 6.5 Hz), 5.53 (d, 1H, J = 6.5 Hz), 7.73–7.85 (m, 2H), 7.80–7.98 (m, 4H), 8.32 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.13$ , 12.35, 13.06, 13.66, 38.31, 40.49, 41.32, 56.84, 123.62, 126.67, 128.70, 129.43, 130.37, 131.12, 131.53, 134.18, 134.62, 138.19, 167.79. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (362.49): calcd C 62.95, H 7.23; found C 62.89, H 7.26.

#### 4.15. (S)-N,N-Diisopropyl-3-methylsulfanyl-2-(N-2naphthylsulfonylamino)propanamide: 6c

Yield 79%. mp 102–103 °C.  $[\alpha]_{\rm D} = -35$  (c1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.09-1.26$ (m, 12H), 3.02–3.24 (m, 2H), 3.37 (m, 2H), 3.68 (s, 3H), 4.40 (m, 1H), 6.53 (d, 1H, J = 10.5 Hz), 7.59–7.72 (m, 2H), 7.82–8.01 (m, 4H), 8.28 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.10$ , 12.33, 13.04, 13.47, 18.69, 40.58, 41.36, 41.55, 54.68, 123.61, 126.78, 128.73, 129.24, 130.42, 131.50, 132.25, 133.36, 134.25, 137.38, 168.39. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (376.51): calcd C 63.80, H 7.49; found C 63.96, H 7.43.

#### 4.16. (S)-N,N-Diisopropyl-3-phenyl-2-(N-2-naphthylsulfonylamino)propanamide: 6d

Yield 80%. mp 153–155 °C.  $[\alpha]_{\rm D} = -46.3$  (c1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.89-1.02$  (m, 12H), 3.02–3.18 (m, 2H), 3.47 (m, 2H), 4.58 (m, 1H), 6.52 (d, 1H, J = 10.1 Hz), 7.23–7.62 (m, 7H), 7.75–7.91 (m, 4H), 8.25 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.24$ , 12.43, 13.07, 13.60, 40.81, 41.56, 41.69, 53.79, 125.56, 125.78, 126.77, 127.15, 127.61, 128.31, 129.24, 129.69, 130.17, 131.82, 132.54, 135.70, 138.36, 139.58, 168.74. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (438.58): calcd C 68.46, H 6.89; found C 68.38, H 6.94.

## 4.17. (2*S*, 3*S*)-*N*,*N*-Diisopropyl-3-methyl-2-(*N*-2-naphthyl-sulfonylamino)pentanamide: 6e

Yield 78%. mp 106–107 °C.  $[\alpha]_D = -29.7$  (c1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.95-1.29$  (m, 15H), 1.32–1.49 (m, 5H,), 2.72–2.80 (m, 1H), 3.05–3.18 (m, 2H), 4.61–4.73 (dd, 1H, J = 3.2 and J = 8.5 Hz), 5.33–5.46 (d, 1H, J = 10.1 Hz), 7.60–7.72 (m, 2H), 7.84–8.02 (m, 4H) 8.39 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.16$ , 11.49, 12.32, 12.91, 13.62, 15.28, 25.83, 34.66, 40.82, 42.05, 67.68, 123.16, 125.59, 127.87, 128.59, 129.69, 130.99, 131.63, 133.43, 134.46, 137.21, 168.32. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (404.57): calcd C 65.31, H 7.97; found C 65.42, H 7.91.

#### 4.18. (S)-N,N-Diisopropyl-3-methyl-2-(N-methyl-N-2naphthylsulfonylamino)butanamide: 6f

Yield 85%. mp 121–123 °C.  $[\alpha]_{\rm D} = -27.3$  (c 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.93$  (d, 3H, J = 7.2 Hz), 1.10 (d, 3H, J = 7.2 Hz), 1.13–1.29 (m, 12H), 2.57 (m, 1H), 3.01 (s, 3H), 3.14–3.28 (m, 2H), 4.49 (d, 1H, J = 3.5 Hz), 7.66–7.71 (m, 2H), 7.72–7.89 (m, 4H), 8.37 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.32$ , 12.18, 12.88, 13.57, 17.19, 19.24, 27.86, 38.76, 41.09, 42.11, 68.01, 123.82, 126.58, 128.25, 129.39, 130.52, 131.28, 131.85, 133.48, 134.68, 137.72, 169.14. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (404.57): calcd C 65.31, H 7.97; found C 65.40, H 7.94.

#### 4.19. (S)-N,N-Diisopropyl-3-phenyl-2-(N-methyl-N-2naphthylsulfonylamino)propanamide: 6g

Yield 77%. mp 129–130 °C.  $[\alpha]_D = -31.5$  (c 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.92-1.11$  (m, 12H), 2.34 (s, 3H), 3.12–3.22 (m, 2H), 3.41 (d, 2H, J = 3.7 Hz), 4.37 (t, 1H, J = 3.7 Hz), 7.47–7.62 (m, 7H), 7.86–7.90 (m, 4H), 8.29 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.29$ , 12.37, 12.74, 13.71, 37.15, 40.93, 41.73, 42.57, 54.39, 128.17, 128.24, 128.69, 129.10, 129.49, 130.08, 130.96, 131.87, 132.11, 133.27; 133.65, 134.54, 135.69, 136.81, 167.58. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (452.61): calcd C 68.99, H 7.12; found C 68.90, H 7.21.

#### 4.20. (S)-N,N-Diisopropyl-2-(N-2naphthylsulfonyl)pipecolinamide: 6h

Yield 82%. mp 138–140 °C.  $[\alpha]_D = -16.4$  (c 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.03-1.28$  (m, 12H), 1.32–1.37 (m,1H), 1.57–1.79 (m, 4H), 2.37–2.65 (m, 2H), 3.10–3.19 (m, 2H), 3.64 (dt, 1H, J = 4.2 Hz and J = 11.9 Hz), 5.46 (t, 1H, J = 4.2 Hz), 7.55–7.66 (m, 2H), 7.91–8.10 (m, 4H), 8.47 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.23$ , 12.29, 12.82, 13.67, 21.10, 24.37, 25.21, 40.80, 41.93, 46.31, 62.72, 123.69, 127.41, 128.74, 129.65, 130.46, 130.59, 131.37, 133.76, 134.41, 137.49, 169.04. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (402.55): calcd C 65.64, H 7.51; found C 65.68, H 7.48.

#### 4.21. (±)-*N*,*N*-diisopropyl-2-(*N*-2naphthylsulfonyl)prolinamide: 6i

Yield 78%. mp 134–135 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.98$ –1.21 (m, 12H), 1.53–1.67 (m, 2H), 2.16–2.35 (m, 2H), 3.02–3.18 (m, 2H), 3.56–3.78 (m, 2H), 4.87 (dt, 1H, J = 3.7 Hz and J = 11.0 Hz), 7.64–7.70 (m, 2H), 7.84–7.99 (m, 4H), 8.39 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.09$ , 12.37, 13.05, 13.74, 23.66, 32.17, 48.32, 56.41, 123.98, 126.53, 128.87, 130.28, 130.86, 131.11, 131.73, 133.68,

134.69, 137.79, 168.31. C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (388.52): calcd C 64.92, H 7.26; found C 65.09, H 7.30.

## 4.22. 2H-1,2-Naphthothiazin-4(3H)-one 1,1-dioxides 7a-i. General procedure

To a solution of LDA (4 equiv for N–H derivatives and 3 equiv for N–Me derivatives of **6**) freshly prepared in THF at –5 °C was added dropwise a solution of *N*-(2-naphthylsulfonyl)amino amide (5 mmol). The reaction mixture was allowed to warm to room temperature over 1 h, and then quenched with a hydrochloric acid solution (1 M, 25 ml). The organic layer was separated and the aqueous layer extracted with methylene chloride ( $3 \times 20$  ml). The organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), and evaporated. The residue was purified by silica gel chromatography [SiO<sub>2</sub>; (EtOAc:hexane)] to afford the 2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1-dioxide.

#### 4.23. 3-Isopropyl-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1dioxide: 7a

Yield 56%. mp 145–147 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.98$  (d, 3H, J = 6.9 Hz), 1.17 (d, 3H, J = 6.9 Hz), 2.85 (m, 1H), 4.48 (dd, 1H, J = 3.6 Hz and 10.2 Hz), 5.47 (d, 1H, J = 10.2 Hz), 7.65 (m,2H), 7.85 (m, 2H), 8.15 (s, 1H), 8.45 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 16.66$ , 19.19, 28.26, 68.45, 123.90, 126.56, 128.79, 129.27, 130.18, 130.36, 131.11, 133.63, 134.10, 137.21, 191.91. IR (KBr):  $\nu = 1679$  (C=O), 1128, 1323 (SO<sub>2</sub>N), 1552 (NH), 1044 (SO<sub>2</sub>). C<sub>15</sub>H<sub>15</sub>NO<sub>3</sub>S (289.33): calcd C 62.26, H 5.22; found C 62.34, H 5.30.

#### 4.24. 3-Methyl-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1dioxide: 7b

Yield 40%. mp 126–128 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.33$  (d, 3H, J = 7.2 Hz), 3.92(qd, 1H, J = 7.2 Hz and J = 6.6 Hz), 5.63 (d, 1H, J = 6.6 Hz), 7.70 (m, 2H), 7.82 (m, 2H), 8.12 (s, 1H), 8.51 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 38.49$ , 56.94, 123.91, 126.51, 128.72, 129.32, 130.29, 130.33, 131.22, 134.01, 134.72, 138.13, 192.90. IR (KBr):  $\nu = 1669$  (C=O), 1126, 1339 (SO<sub>2</sub>N), 1569 (NH), 1048 (SO<sub>2</sub>). C<sub>13</sub>H<sub>11</sub>NO<sub>3</sub>S (261.28): calcd C 59.75, H 4.23; found C 59.62, H 4.31.

#### 4.25. 3-Methylsulfanylmethyl-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1-dioxide: 7c

Yield 52%. mp 175–177 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 3.32$  (m, 2H), 3.91 (s, 3H), 4.45 (m, 1H), 6.60 (d, 1H, J = 10.6 Hz), 7.62 (m, 2H), 7.83 (m, 2H), 8.21 (s, 1H), 8.50 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 18.78$ , 41.57, 53.98, 123.89, 126.46,

128.61, 129.34, 130.28, 130.38, 131.25, 133.56, 134.18, 137.30, 198.18. IR (KBr):  $\nu = 1695$  (C=O), 1125, 1323 (SO<sub>2</sub>N), 1570 (NH), 1032 (SO<sub>2</sub>). C<sub>14</sub>H<sub>13</sub>NO<sub>3</sub>S<sub>2</sub> (307.38): calcd C 54.70, H 4.25; found C 54.81, H 4.29.

#### 4.26. 3-Benzyl-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1dioxide: 7d

Yield 48%. mp 132–134 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 3.41 (m, 2H), 4.46 (m, 1H), 6.66 (d, 1H, J = 10.2 Hz), 7.26–7.61 (m, 7H), 7.87–7.94 (m, 2H), 8.06 (s, 1H), 8.10 (s, 1H). – <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 41.56, 54.08, 125.62, 125.71, 126.81, 127.12, 127.71, 128.16, 128.59, 128.74, 129.00, 131.62, 132.04, 135.90, 138.18, 139.03, 198.08. IR (KBr):  $\nu$  = 1671 (C=O), 113, 1326 (SO<sub>2</sub>N), 1549 (NH), 1050 (SO<sub>2</sub>). C<sub>19</sub>H<sub>15</sub>NO<sub>3</sub>S (337.38): calcd C 67.64, H 4.47; found C 67.48, H 4.34.

#### **4.27. 3**-(1-(*S*)-Methylpropyl)-2*H*-1,2-naphthothiazin-4(3*H*)-one 1,1-dioxide: 7e

Yield 50%. mp 112–114 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.95$  (d, 3H, J = 5.2 Hz), 1.01 (t, 3H, J = 3.5 Hz), 1.09–1.31 (m, 2H), 2.58–2.71 (m, 1H), 4.54–4.72 (dd, 1H, J = 3.2 and J = 10.1 Hz), 5.21–5.33 (d, 1H, J = 10.1 Hz), 7.64–7.76 (m, 2H), 7.95–8.01 (m, 2H) 8.25 (s, 1H), 8.56 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 11.5$ , 14.62, 25.92, 34.72, 67.45, 122.90, 125.55, 127.76, 128.69, 129.24, 130.35, 131.14, 133.53, 134.18, 137.01, 194.85. IR (KBr):  $\nu = 1687$  (C=O), 1139, 1313 (SO<sub>2</sub>N), 1558 (NH), 1034 (SO<sub>2</sub>).

## 4.28. 3-Isopropyl-*N*-methyl-1,2-naphthothiazin-4(3*H*)-one 1,1-dioxide: 7f

Yield 70%. mp 152–154 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.96$  (d, 3H, J = 7.0 Hz), 1.18 (d, 3H, J = 7.0 Hz), 2.40 (m, 1H), 2.82 (s, 3H), 4.47 (d, 1H, J = 3.5 Hz), 7.67 (m,2H), 7.86 (m, 2H), 8.17 (s, 1H), 8.46 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 16.49$ , 19.03, 27.96, 38.65, 68.07, 123.88, 126.49, 128.66, 129.41, 130.22, 130.40, 131.09, 133.58, 134.18, 137.11, 191.72. IR (KBr):  $\nu = 1672$  (C=O), 1138, 1342 (SO<sub>2</sub>N), 2810 (NCH<sub>3</sub>), 1040 (SO<sub>2</sub>). C<sub>16</sub>H<sub>17</sub>NO<sub>3</sub>S (303.36): calcd. C 63.34, H 5.64; found C 63.22, H 5.61.

## 4.29. 3-Benzyl-*N*-methyl-1,2-naphthothiazin-4(3*H*)-one 1,1-dioxide: 7g

Yield 62%. mp 138–140 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 2.29$  (s, 3H), 3.38 (d, 2H, J = 3.8 Hz), 4.33 (t, 1H, J = 3.8 Hz), 7.25–7.77 (m, 7H), 7.87–7.93 (m, 2H), 8.08 (s, 1H), 8.10 (s, 1H). <sup>13</sup>C NMR

(75 MHz, CDCl<sub>3</sub>):  $\delta$  = 37.11, 42.06, 54.21, 128.03, 128.34, 128.49, 129.12, 129.21, 129.65, 130.86, 131.76, 132.21, 133.03; 133.75, 134.00, 135.35, 136.51, 197.71. IR (KBr):  $\nu$  = 1675 (C=O), 1132, 1340 (SO<sub>2</sub>N), 2817 (NCH<sub>3</sub>), 1047 (SO<sub>2</sub>). C<sub>20</sub>H<sub>17</sub>NO<sub>3</sub>S (351.40): calcd C 68.35, H 4.87; found C 68.48, H 5.01.

#### 4.30. 2,3,4,5-tetrahydropyrido[1,2-b] [1,2]naphthothiazin-7-one 12,12-dioxide: 7h

Yield 46%. mp 98–101 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.1-1.4$  (m, 1H), 1.5–1.8 (m, 4H), 2.4–2.7 (m, 2H), 3.66 (dt, 1H, J = 3.6 Hz and J = 11.4 Hz), 5.23 (t, 1H, J = 3.6 Hz), 7.61–7.66 (m, 2H), 7.82–7.86 (m, 2H), 8.12 (s, 1H), 8.47 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 20.10, 24.22, 24.31, 46.16, 63.92, 123.81, 127.01, 128.82, 129.55, 130.29, 130.46, 131.19, 133.78, 134.30, 137.42, 192.94. IR (KBr): <math>\nu = 1692$  (C=O), 1127, 1343 (SO<sub>2</sub>N), 1042 (SO<sub>2</sub>). C<sub>16</sub>H<sub>15</sub>NO<sub>3</sub>S (301.35): calcd C 63.77, H 5.01; found C 63.54, H 5.11.

#### 4.31. 2,3,4- trihydropyrro[1,2-b] [1,2]naphthothiazin-6one 11,11-dioxide: 7i

Yield 21%. mp 86–88 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.80-1.92$  (m, 2H), 2.05–2.22 (m, 2H), 3.45–3.62 (m, 2H), 4.70 (dt, 1H, J = 3.9 Hz and J = 10.9 Hz), 7.63–7.68 (m, 2H), 7.83–7.87 (m, 2H), 8.16 (s, 1H), 8.49 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 24.76$ , 31.14, 48.17, 57.43, 123.90, 126.93, 128.75, 129.48, 130.16, 130.37, 131.15, 133.61, 134.23, 137.51, 193.16. IR (KBr):  $\nu = 1689$  (C=O), 1130, 1335 (SO<sub>2</sub>N), 1040 (SO<sub>2</sub>). C<sub>15</sub>H<sub>13</sub>NO<sub>3</sub>S (287.32): calcd C 62.70, H 4.55; found C 62.75, H 4.42.

#### 4.32. N-methyl-2-naphthylsulfonamide: 8

To a solution of 2-naphthylsulfonyl chloride (10 mmol) in  $CH_2Cl_2$  (50 ml) stirred at 0 °C was added dropwise a solution of methylamine 30% in water (22 mmol). The reaction mixture was allowed to warm to room temperature over 1 h. Water (30 ml) was then added, and the aqueous phase was extracted with  $CH_2Cl_2$  (3 × 30 ml). The extract was dried (Na<sub>2</sub>SO<sub>4</sub>), subjected to filtration and concentrated. The residue was purified by flash column chromatography [SiO<sub>2</sub>; (EtOAc:cyclohexane)].

Yield 88%. mp 101–102 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 2.76$  (d, 3H, J = 5.3 Hz), 5.45 (q, 1H, J = 5.3 Hz), 7.56–7.67 (m, 2H), 7.78–7.95 (m, 4H), 8.32 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta = 29.91$ , 122.69, 127.33, 127.73, 128.25, 128.60, 128.88, 129.12, 131.86, 134.23, 136.61.

#### 4.33. N,N-Diethyl-2-naphthylsulfonamide: 9

To a solution of diethylamine (12 mmol) and triethylamine (12 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (100 ml) stirred at 0 °C was added dropwise solution а of 2-naphthylsulfonyl chloride (10 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 ml). The reaction mixture was allowed to warm to room temperature over 1 h. Water (30 ml) was then added, and the aqueous phase was extracted with  $CH_2Cl_2$  (3 × 30 ml). The extract was dried (Na<sub>2</sub>SO<sub>4</sub>), subjected to filtration and concentrated. The residue obtained was purified by flash column chromatography [SiO<sub>2</sub>; (EtOAc:cyclohexane)].

Yield 92%. mp 89–91 °C. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.15 (t, 6H, *J* = 6.9 Hz), 3.24 (q, 4H, *J* = 6.9 Hz), 7.65–7.78 (m, 2H), 7.84–7.98 (m, 4H), 8.25 (s, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 16.38, 37.42, 122.58, 127.41, 127.86, 128.11, 128.74, 129.05, 129.32, 131.56, 134.85, 135.96.

#### 4.34. Deuteration experiments

#### 4.34.1. Using n-BuLi. General procedure

A solution of **8** or **9** in THF (0.5 M) was cooled to  $-5 \,^{\circ}$ C under argon atmosphere and treated with *n*-BuLi in hexane (2.2 and 1.1 equiv, respectively). After stirring at  $-5 \,^{\circ}$ C for 20 min, the reaction mixture was warmed quickly to room temperature and stirred for an additional 1 h before quenching with CD<sub>3</sub>OD (10 equiv). The resulting solution was stirred for 1 h and quenched with saturated NH<sub>4</sub>Cl solution. The aqueous portion was extracted with CH<sub>2</sub>Cl<sub>2</sub>, and the extract was dried (Na<sub>2</sub>SO<sub>4</sub>), subjected to filtration, and concentrated. The residue was purified by flash column chromatography [SiO<sub>2</sub>; (EtOAc:cyclohexane)]. **10.** Use of the general procedure with the following materials [**8** (0.221 g, 1 mmol), *n*-BuLi (0.88 ml, 2.5 M, 2.2 mmol), CD<sub>3</sub>OD (0.40 ml, 10 mmol) in THF (10 ml)], followed by flash column chromatography (EtOAc/cyclohexane 2:8) afforded a light yellow solid. Yield 79%. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 2.78$  (d, 3H, J = 5.3 Hz), 5.21 (bs, 1H), 7.54–7.61 (m, 2H), 7.87–7.94 (m, 2H), 8.06 (s, 1H), 8.15 (s, 1H).

**11.** Use of the general procedure with the following materials [9 (0.263 g, 1 mmol), *n*-BuLi (0.44 ml, 2.5 M, 1.1 mmol), CD<sub>3</sub>OD (0.40 ml, 10 mmol) in THF (10 ml)], followed by flash column chromatography (EtOAc/cyclohexane 2:8) afforded a light yellow solid. Yield 87%. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.14$  (t, 6H, J = 6.9 Hz), 3.22 (q, 4H, J = 6.94 Hz), 7.62–7.77 (m, 2H), 7.80–7.96 (m, 2H), 8.10 (s, 1H), 8.18 (s, 1H).

#### 4.34.2. Using LDA. General procedure

A solution of **8** or **9** in THF (0.2 M) was cooled to -5 °C under argon atmosphere and treated with LDA (2.2 and 1.1 equiv, respectively, prepared by dropwise addition of *n*-BuLi to a solution of HN(*i*-Pr)<sub>2</sub> in THF (1 M) precooled to 0 °C). After stirring for 20 min, a 10 ml aliquot was quenched by addition to CD<sub>3</sub>OD precooled to -5 °C. The reaction mixture was warmed to room temperature and stirred for an additional 1 h before quenching with CD<sub>3</sub>OD. Each aliquot was extracted with CH<sub>2</sub>Cl<sub>2</sub>, and the extract was dried (Na<sub>2</sub>SO<sub>4</sub>), subjected to filtration, and concentrated. The residue was purified by flash column chromatography [SiO<sub>2</sub>; (EtOAc:cyclohexane)].

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### References

- V.J. Aran, P. Goya, C. Ochoa, Adv. Heterocycl. Chem. 44 (1988) 81.
- [2] J.G. Lombardino, D.E. Kuhla, Adv. Heterocycl. Chem. 28 (1981) 73.
- [3] M.S. Chamber, S.C. Hobbs, M.I. Graham, A.P. Watt, S.R. Fletcher, R. Baker, S.B. Freedman, S. Patel, A.J. Smith, V.G. Matassa, Bioorg. Med. Chem. Lett. 5 (1995) 2303.
- [4] W.G. Rice, J.G. Supko, L. Malspeis, R.W. Buckheit, D. Clanton, M. Bu, L. Graham, C.A. Schaeffer, J.A. Turpin, J. Domagala, R. Gogliotti, J.P. Bader, S.M. Halliday, L. Coren, R.C. Sowder, L.O. Arthur, L.E. Henderson, Science 270 (1995) 1194.
- [5] E.S. Lazer, C.K. Miao, C.L. Cywin, R. Sorcek, H.C. Wong, Z. Meng, I. Potocki, M. Hoermann, R.J. Snow, M.A. Tschantz, T.A. Kelly, D.W. McNeil, S.J. Coutts, L. Churchill, A.G. Graham, E. David, P.M. Grob, W. Engel, H. Meier, G. Trummlitz, J. Med. Chem. 40 (1997) 980.
- [6] T.M. Hasegawa, T. Furuta, T. Tsuda, Japanese Patent 71/22,027, 1971.

- [7] J.G. Lombardino, E.H. Wiseman, US Patent 3,862,319, 1975.
- [8] J.C. Sircar, H. Zinnes, J. Shavel, US Patent 3,878,198, 1975.
- [9] J.G. Lombardino, Antiinflammatory Agents: Chemistry and Pharmacology, Academic Press, New York, 1974 p. 129.
- [10] J.G. Lombardino, E.H. Wiseman, W. McLamore, J. Med. Chem. 14 (1971) 1171.
- [11] J.G. Lombardino, E.H. Wiseman, J. Chiaini, J. Med. Chem. 16 (1973) 493.
- [12] P. Catsoulacos, C. Camoutsis, J. Heterocycl. Chem. 16 (1979) 1503.
- [13] R.A. Abramovitch, K.M. More, I. Shinkai, P.C. Srinivasan, J. Chem. Soc. Chem. Commun. (1976) 771.
- [14] M.R. Panara, G. Renda, M.G. Sciulli, J. Pharmacol. Exp. Ther 290 (1999) 276.
- [15] C.B. Schapira, I.A. Perillo, S. Lamdan, J. Heterocycl. Chem. 17 (1980) 1281.
- [16] I.A. Perillo, C.B. Schapira, S. Lamdan, J. Heterocycl. Chem. 20 (1983) 155.

- [17] R. Pfister, P. Zeller, B. Binder, O. Hromatka, Britsh Patent 2,003,877, 1979.
- [18] G. Trummlitz, W. Engel, E. Seeger, W. Haarmann, G. Engelhardt, US Patent 4,137,313, 1979.
- [19] D. Binder, O. Hromatka, F. Geissler, K. Schmied, C.R. Noe, K. Burri, R. Pfister, K. Strub, P. Zeller, J. Med. Chem. 30 (1987) 678.
- [20] W.I.I. Bakker, O.B. Familoni, J. Padfield, V. Snieckus, Synlett (1997) 1079.
- [21] H. Kaufmann, H. Zobel, Chem Ber. B 55 (1922) 1499.
- [22] J.G. Lombardino, J. Org. Chem. 36 (1971) 1843.
- [23] G. Trummlitz, E. Seeger, W. Engel, H. Teufel, G. Englehardt, W. Haarmann, US Patent 3,992,535, 1976.
- [24] G. Steiner, Justus Liebigs Ann. Chem. (1978) 635.
- [25] Y. Kacem, J. Kraiem, E. KerKeni, A. Bouraoui, B. Ben Hassine, Eur. J. Pharm. Sci. 16 (2002) 221.
- [26] H.W. Gschwend, H.R. Rodriguez, Organic Reactions, vol. 1, Am. Chem. Soc., New York, 1979, p. 26.
- [27] I. Omae, Chem. Rev. 79 (1979) 287.

- [28] V. Snieckus, Bull. Soc. Chim. Fr. (II) (1988) 67.
- [29] V. Snieckus, Chem. Rev. 90 (1990) 879.
- [30] N.S. Narasimhan, R.S. Mali, Synthesis (1983) 957.
- [31] P. Beak, A.I. Meyers, Acc. Chem. Res. 19 (1986) 356.
- [32] R.M. Freidinger, J.S. Hinkle, D.S. Perlow, B.H. Arison, J. Org. Chem. 48 (1983) 77.
- [33] M.B. Berry, D. Craig, Synlett (1992) 41.
- [34] P.J. Maurer, H. Takahata, H. Rapoport, J. Am. Chem. Soc. 106 (1984) 1095.
- [35] F.B. Thomas, H. Rapoport, J. Am. Chem. Soc. 103 (1981) 6157.
- [36] W.O. Moss, E. Wakefield, M.F. Mahon, K.C. Molloy, R.H. Bradbury, N.J. Hales, T. Gallagher, Tetrahedron 48 (1992) 7551.
- [37] P. Beak, R.A. Brown, J. Org. Chem. 47 (1982) 34.
- [38] P. Beak, S.T. Kerrick, D.J. Gallagher, J. Am. Chem. Soc. 115 (1993) 10628.
- [39] B.L. Eun, K.K. Soon, G.K. Sang, Arch. Pharm. Res. 22 (1999) 44.
- [40] G. Dannhardt, W. Kiefer, Eur. J. Med. Chem. 36 (2001) 109.
- [41] S.C. Watson, J.F. Eastham, J. Organomet. Chem. 9 (1967) 165.