



Full paper / Mémoire

The “Ouzo effect”: Following the spontaneous emulsification of *trans*-anethole in water by NMR

David Carteau, Dario Bassani*, Isabelle Pianet*

ISM – UMR 5255, Université Bordeaux-1, 351, cours de la Libération, 33405 Talence cedex, France

Received 1 May 2007; accepted after revision 6 November 2007

Available online 4 March 2008

Abstract

Some aperitifs, like Pastis or Ouzo, are ethanol extracts of anis seeds. When diluted with water, these aperitifs, which are principally composed of water, ethanol and *trans*-anethole, become cloudy instantaneously. This phenomenon is the result of a spontaneous emulsification sometimes termed “Ouzo effect”. By employing deuterated ethanol and water, it is possible to follow the aggregation process by NMR, using DOSY and TOCSY experiments. Analysis of these results permits us to propose an aggregation process mechanism by which *trans*-anethole initially forms small aggregates that are visible by NMR, which then coalesce to form small droplets of micron-size that are “NMR invisible”. **To cite this article:** D. Carteau et al., *C. R. Chimie* 11 (2008).

© 2007 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

Résumé

Certains apéritifs, comme le Pastis ou l’Ouzo, sont des extraits de l’anis étoilé. Lorsqu’on dissout ces apéritifs dans l’eau, majoritairement composés d’eau, d’éthanol et de *trans*-anéthole, le mélange prend immédiatement une consistance laiteuse. Ce phénomène est le résultat d’une émulsion spontanée qui s’opère entre une petite quantité d’eau et une solution riche en huile : « l’effet Ouzo ». En utilisant de l’éthanol D6 et D₂O, il est possible de suivre le mécanisme d’agrégation par RMN, en utilisant des expériences de type DOSY et EXSY. L’analyse des expériences permet de proposer un mécanisme d’agrégation par lequel le *trans*-anéthol s’auto-associerait pour former des petits agrégats visible en RMN, avant de coalescer pour former des microgouttelettes de plus grandes tailles (de l’ordre du micromètre), invisible en RMN. **Pour citer cet article :** D. Carteau et al., *C. R. Chimie* 11 (2008).

© 2007 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

Keywords: *Trans*-anethole; “Ouzo effect”; Aggregation; DOSY; EXSY

Mots-clés : *trans*-anéthole ; « effet Ouzo » ; agrégation ; DOSY ; EXSY

1. Introduction

The “Ouzo effect” was coined by Vitale and Katz [1] to describe the spontaneous emulsification accompanying the preparation of the Greek beverage Ouzo, because

* Corresponding authors.

E-mail addresses: d.bassani@ism.u-bordeaux1.fr (D. Bassani), i.pianet@ism.u-bordeaux1.fr (I. Pianet).

Ouzo is more drunk than Pastis in the United States due to the presence of an important Greek community. Nevertheless, French Pastis lovers are familiar with this process, and revel in it when they prepare their favourite aperitif. This “Ouzo effect” is the result of a spontaneous emulsification between two immiscible components, thanks to a third component in which both components are soluble. This phenomenon arouses intermittent interest in the scientific community, principally due to a number of potential applications [2]. Thus, mixing an ethanolic solution of the essential oil *trans*-anethole (*t*-A) in water instantaneously gives a metastable solution of cloudy aspect. Despite its noticeable visual stability over several few hours, continuous molecular reorganizations occur than can be probed by using NMR.

In a previous paper, we reported the mechanism of the aggregation process that occurs at the early beginning of *t*-A emulsification [3] and the effect of aggregation processes on its photochemical behaviour [4]. Herein, a more detailed analysis of each state of *t*-A emulsification is described, showing that before reaching a molecular metastable state, *t*-A molecules initially reorganize into small aggregates of angstrom-size, thus visible by using standard liquid NMR experiments. Simultaneously, these small aggregates coalesce inbetween them to form small micrometer-size droplets.

2. Materials and methods

2.1. Sample preparation

Microemulsions were freshly prepared using a set procedure to ensure reproducibility: an amount of *trans*-Anethole (*t*-A, Aldrich, >99% pure) was dissolved in ethanol D6 (Euriso-Top) in order to obtain a homogeneous single-phase solution. Subsequently, deuterium oxide (Euriso-Top) was added to the alcoholic solution to reach a concentration of 5 mM in *t*-A and a solvent composition of 5% C₂D₅OD/D₂O. A 0.5-mL aliquot was used for the NMR experiments.

2.2. NMR experiments

NMR experiments were performed at 298 K on a Bruker DPX 400 equipped with a 5-mm gradient inverse broad band probe. All proton chemical shifts are given with respect to TMS as an external reference. Three types of experiments were recorded.

2.2.1. Kinetics

A sample containing 5 mM of *trans*-anethole in a solvent mixture containing 5% C₂D₅OD/D₂O was

prepared as previously described. An external capillary containing 5 mM of methanol was added in the NMR tube and was used as an external quantitative. Then, proton spectra were recorded every 10 min immediately after the sample preparation, and the ratio of aggregated form/free form was estimated with respect to the total amount of *trans*-anethole present in the sample.

2.2.2. Exchange rate measurements

The 2D ¹H EXSY NMR spectra were recorded using the pulse sequence of Jeener et al. [5] in the phase-sensitive mode with time-proportional phase incrementation [6]. Twenty-four experiments were recorded at two successive mixing times (60 and 0 ms) every 15 min during the first hour and then every 30 min during the 10 following hours. The auto and cross-peak volumes were determined after phase and baseline correction of the two dimensions using the Bruker UXNMR software. Rate exchanges were obtained from these experiments by using the method of Perrin, Gipe and Dwyer [7,8] that allows extraction of rates from a single experiment recorded at one mixing time by resolving the kinetic matrix *L*:

$$L = (1/tm)\ln(A) = (1/tm)U\ln(\lambda)U^{-1}$$

where the elements of the matrix *A* are composed of the intensity of the peaks obtained from the EXSY spectra, as previously described [9]. In the case of an exchange between two species (*t*-A_{aggregate} and *t*-A_{free}), *L* is a 4 × 4 matrix:

$$L = - \begin{bmatrix} R_1t - A_{\text{aggregate}} + k_{-a} & -k_a \\ -k_{-a} & R_1t - A_{\text{free}} + k_a \end{bmatrix}$$

where *R*₁ is the relaxation rate for each species and *k*_a, *k*_{-a}, the rate constants for the exchange.

2.2.3. Diffusion measurements

Diffusion measurements were performed using a 2D ¹H NMR pulsed-gradient experiment: the stimulated spin-echo sequence [10] which leads to a 2D spectrum in which the second dimension is obtained after a Fourier Transformation and gives the ¹H NMR spectrum, and the first dimension is obtained after a Laplace Transformation using the Bruker software and gives the translational Diffusion Coefficients' dimension. The 2D DOSY spectrum was recorded using the following parameter: intergradient delay Δ, 150 ms; gradient pulse duration δ, 1,3 ms and 16 different gradient values varying linearly from 0.01 to 0.4 G/m).

3. Results

3.1. Behaviour of *trans*-anethole in water

Fig. 1 shows a proton spectrum of *t*-A obtained approximately 10 min after emulsification. According to the expected pattern for the proton spectrum, two distinct sets of readily assigned resonances can be distinguished, suggesting that *t*-A can exist in at least two forms. Three types of NMR experiments confirm that these two forms correspond to free and aggregated *t*-A. First, EXSY experiments acquired on the mixture clearly demonstrate a slow exchange between the two species (Fig. 2), confirming that the same compound is undergoing exchange between two states. Second, DOSY experiments performed on the mixture shows that the two species have very different diffusion coefficient values: about 6×10^{-10} and 1×10^{-10} $\text{m}^2 \text{s}^{-1}$, allowing the unambiguous assignment of the free and the aggregated forms, respectively, as shown in the DOSY spectrum displayed in Fig. 3. Since we are in the slow exchange limit (Δ/τ between 0.075 and 0.18 for the aggregated specie, and close to 0.11 for the free form) [11], we can consider that the two diffusion coefficients measured are representatives of those belonging to the individual species. Third, proton spectra recorded at different ethanol concentrations (up to 60% ethanol) show that only the free form of *t*-A is present above 40% of ethanol [12]. However, the most interesting feature of these experiments is the systematic deshielding effect of ca. 0.6 ppm for the overall ^1H resonances of the free form as compared to the aggregated form. This phenomenon can be easily explained by the hydrophilic/hydrophobic environment felt by *t*-A in the free and aggregated forms.

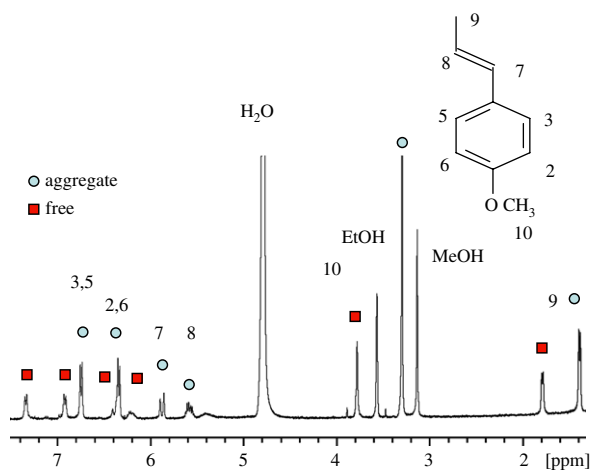


Fig. 1. Proton spectrum of *t*-A (400.13 MHz), 10 min after the mixture preparation.

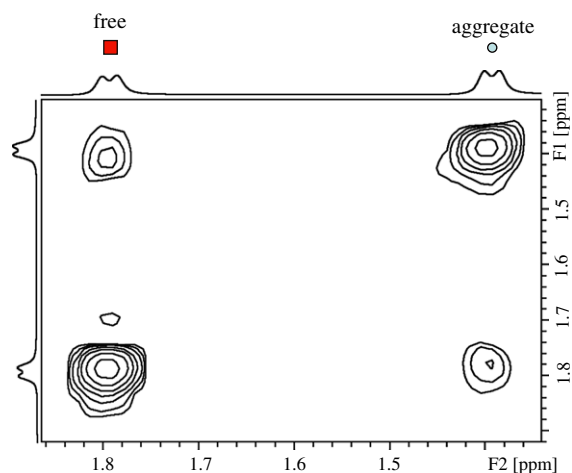


Fig. 2. EXSY spectrum recorded with a 60 ms mixing time showing the interconversion between the two forms of *t*-A.

3.2. Aggregation process

When water is added to an alcoholic solution of *t*-A, the mixture instantaneously takes a cloudy aspect that appears visually stable for at least 6–8 h, after which a two-phase solution can be observed [13]. But this stable appearance is only visual: as an example, Fig. 4A displays the methyl 9 part of the proton spectra acquired in the course of time. It shows that if the intensity of the free form (1.78 ppm) remains constant along at least 70 min, a decrease of the signal assigned to the aggregated form (1.39 ppm) is observed over the same period. This decrease has been followed and quantified with respect to the intensity of the methyl signal of methanol added to the mixture as an internal standard (5 mM, final concentration). From these results, it is possible to understand that anethole exists in three distinct forms, of which only two are observable in the NMR ^1H spectra. It is thus possible to follow, all along the duration of the experiment, the proportion of *t*-A observed in two forms (small aggregates or free) and to deduct the proportion of *t*-A that is invisible by NMR (Fig. 4B). After 40 min, the latter reaches a plateau value close to 3.5 mM, which represents up to 70% of the total *t*-A; at the same time, the proportion of small aggregates decreases to a concentration close to 0.7 mM, whereas the proportion of free *t*-A remains constant (0.8 mM). It is only after ca 40 min that a metastable state is reached, which remains stable for several hours, as shown by liquid NMR observations.

The fraction of *t*-A not visible in the NMR proton spectrum recorded using classical liquid techniques is assigned to *t*-A present as small droplets that are several micrometers in size and that are responsible for the

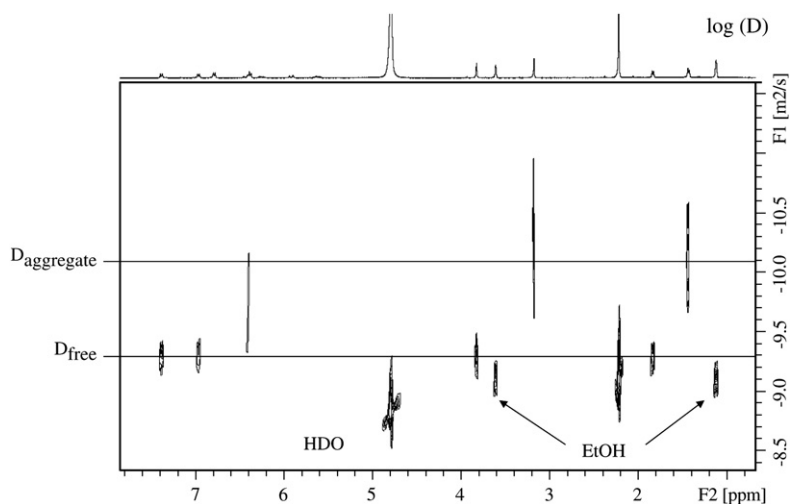


Fig. 3. DOSY 2D spectrum of a hydroalcoholic solution of *t*-A 5 mM, showing two distinct diffusion coefficient values allowing the assignment of the free and aggregated forms.

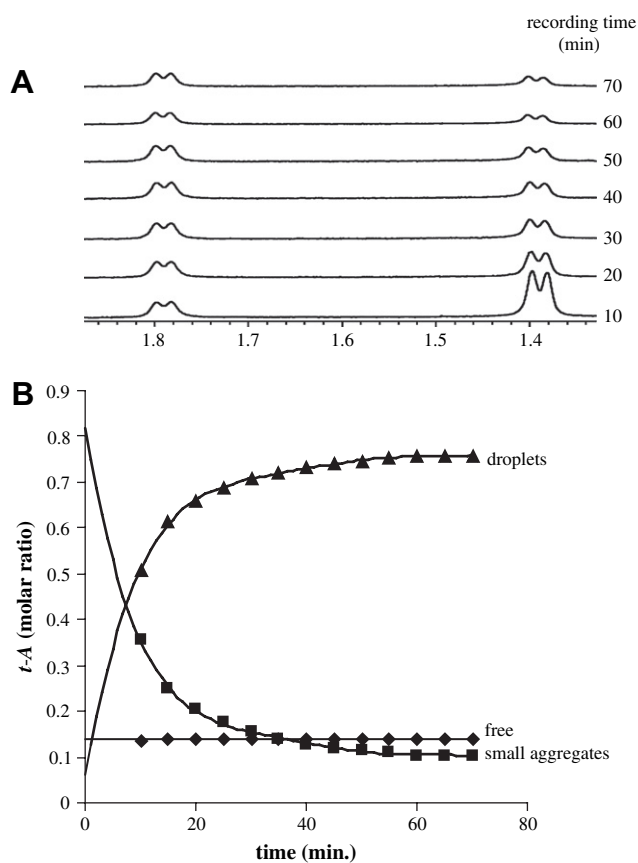
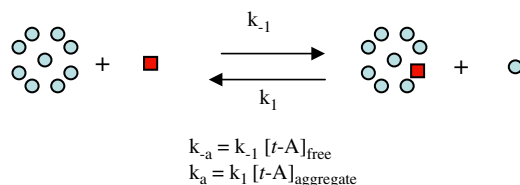


Fig. 4. Evolution of the different forms of *t*-A along the time. (A) Part of proton spectra showing the doublet representative of each form of methyl 9. (B) Evolution of *trans*-anethole molar fraction against time under its free (diamond), aggregated (square) and NMR invisible (triangle) forms. The solid lines are the best-fit using a monoexponential function for the aggregates ($y = Ae^{-kt}$, $k = 11.5 \text{ min}^{-1}$).

cloudy aspect of the solution. Two observations support this hypothesis: first, the proportion of *t*-A, which is being indirectly followed through the disappearance of *t*-A in the NMR spectra (Fig. 4B, droplets) can be fitted to a monoexponential function consistent with an Ostwald ripening process. This behaviour is in agreement with previous investigations of *t*-A droplet growth by Grillo et al. using Small-Angle Neutron Scattering [13]. Second, the part of *t*-A that escapes detection does so either due to differences in magnetic susceptibility of anethole in different environments or due to the predominance of interactions specific to emulsions such as a decrease in T_2 , chemical shift anisotropy or/and dipolar coupling. Concerning this last point, it is noteworthy that HR-MAS spectra recorded at different rotation speeds up to 10 kHz (Fig. 5) show that only a very small part of the signal is recovered, suggesting that the disappearance of a large part of the anethole is due to residual dipolar couplings/anisotropy, since the effect of magnetic susceptibility, which does not exceed 10–100 Hz in a solution, should be averaged at a speed of 10 kHz [14].

3.3. Kinetics

EXSY experiments were recorded over the course of time using a constant mixing time value of 60 ms. EXSY experiments with a zero mixing time were intercalated inbetween each experiment in order to have a reference area matrix for each measurement. The system was considered to undergo only small variations during the time required to acquire the NMR data. Therefore, EXSY NMR experiments allow one to observe the exchange of magnetization between the free



Scheme 1. Study of exchange of magnetization in a bimolecular process by EXSY NMR experiment.

and the aggregated forms: such an exchange is adequately described by the bimolecular process illustrated in Scheme 1. Thus, the rate constant values k_a and k_{-a} , extracted from the area matrices as previously described [9], are, in fact pseudo-order-one rate constants intrinsically expressed in Hz or s^{-1} , and are concentration-dependent (k_a depends on $[t-A]_{\text{aggregate}}$ and k_{-a} on $[t-A]_{\text{free}}$ as explained in Scheme 1).

Pseudo-order-one rate constant values k_a and k_{-a} evolved till 4 h (Fig. 6). The pseudo-order-one rate constant k_{-a} , that reflects the change from the aggregated form to the free form, as shown in the inset of Fig. 6, increases from 0.8 to 2 s^{-1} . Since $[t-A]_{\text{free}}$ remains constant, the evolution of k_{-a} reflects the evolution of k_{-1} . During this time, the pseudo-order-one rate constant k_a that reflects the inverse exchange from the free form to the aggregated state remains stable at 1.3 s^{-1} . Since $[t-A]_{\text{aggregate}}$ decreases all along the time, it suggests that, as expected, k_1 increases as k_{-1} until a metastable equilibrium is reached, in which the three forms of *t*-A coexist.

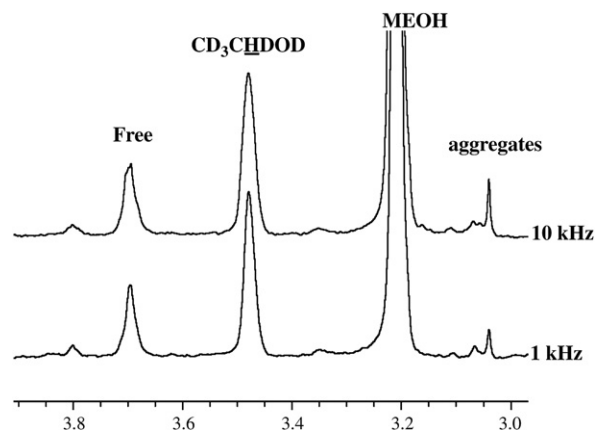


Fig. 5. HR-MAS spectra recorded at 1 and 10 kHz of an hydroalcoholic *t*-A (4 mM) solution mixture: expansion showing the Me 10 region.

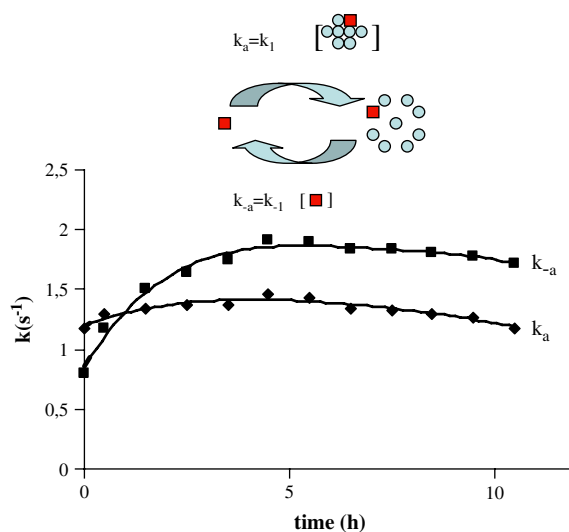
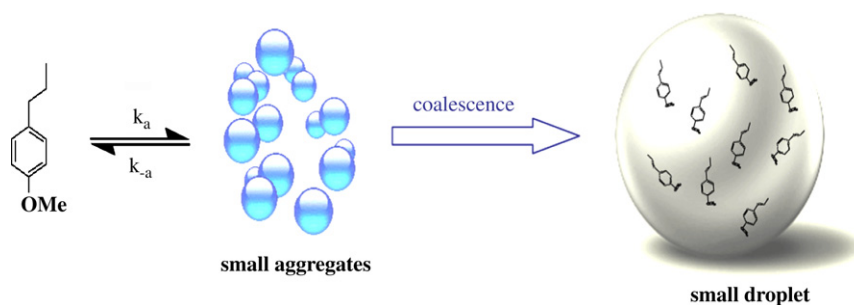


Fig. 6. k_a and k_{-a} evolution over the time course measured from EXSY experiments.

Scheme 2. Proposed mechanism of *t*-A aggregation process.

4. Conclusion

The results we reported herein clearly demonstrate that the spontaneous emulsification of *t*-A in water proceeds via a molecular maturation process that can be probed using NMR spectroscopy (Scheme 2): *t*-A molecules are able to give small aggregates of around 2-nm diameter in which all *t*-A molecules are sufficiently mobile to be observed by classical liquid NMR experiments. An exchange is still occurring all along the time course between both free and aggregated form. Simultaneously, a coalescence process of the small aggregates is taking place to give micrometer-size droplets; *t*-A embedded into these droplets are not visible using liquid NMR techniques, but its behaviour can be indirectly followed and evolved with respect to an Ostwald ripening process.

Acknowledgment

Financial support from the Région Aquitaine, ANRT and the CNRS are gratefully acknowledged.

References

- [1] S.A. Vitale, J.L. Katz, *Langmuir* 19 (2003) 4105.
- [2] F. Ganachaud, J.L. Katz, *ChemPhysChem* 6 (2005) 209.
- [3] D. Carteau, I. Pianet, P. Brunerie, B. Guillemat, D. Bassani, *Langmuir* 33 (2007) 3561.
- [4] D. Carteau, P. Brunerie, B. Guillemat, D. Bassani, *Photochem. Photobiol. Sci.* 6 (2007) 423.
- [5] J. Jeener, B.H. Meir, P. Bachmann, J. Ernst, *J. Chem. Phys.* 71 (1979) 4546.
- [6] D. Marion, K. Wüthrich, *Biochem. Biophys. Res. Commun.* 113 (1983) 967.
- [7] C.L. Perrin, R.K. Gipe, *J. Am. Chem. Soc.* 106 (1984) 4036.
- [8] C.L. Perrin, T.J. Dwyer, *Chem. Rev.* 90 (1990) 935.
- [9] I. Pianet, F. Fouquet, M. Pereyre, M. Gielen, F. Kayser, M. Biesmans, R. Willem, *Magn. Reson. Chem.* 32 (1994) 617.
- [10] W.S. Price, *Concepts Magn. Reson.* 9 (1997) 299.
- [11] E.J. Cabrita, S. Berger, P. Braüer, J. Käger, *J. Magn. Reson.* 157 (2002) 124.
- [12] D. Carteau, thèse de l'université Bordeaux 1, École doctorale des sciences chimiques, N° d'ordre 3255.
- [13] I. Grillo, *Colloids Surf., A: Physicochem. Eng. Aspects* 225 (2003) 153.
- [14] G. Lippens, M. Bourdonneau, C. Dhalluin, R. Warrass, T. Richert, C. Seetharaman, C. Boutillon, M. Piotto, *Curr. Org. Chem.* 3 (1999) 147.