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Syntheses of the tedanolides and myriaporones

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Abstract

This review covers the synthetic contributions leading to the syntheses of the polyketides tedanolide, 13-deoxytedanolide and the structurally related myriaporones. Fragment syntheses that lead to valuable insight into the subtle synthetic problems of the tedanolides are also presented. *To cite this article: N. Schübel et al., C. R. Chimie 11 (2008).* © 2008 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

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1. Introduction

In 1984, Schmitz isolated a novel compound, tedanolide (1) from the Caribbean sponge *Tedania ignis* [1]. The crystalline nature of the compound enabled determination of the structure by X-ray crystallography. Five other related natural products forming the tedanolide family have been isolated from various organisms, namely tedanolide (1), 13-deoxytedanolide (2), tedanolide C (3) and candidaspongiolides (4, 5) (Fig. 1).

The tedanolides' complex architecture, in addition to their high cytotoxicity, attracted considerable interest within the synthetic community. Initial biological evaluation showed an arrest of P338 tumor cell growth in S-phase [1]. Seven years after, Fusetani et al. reported the isolation of a deoxygenated analogue, 13-deoxytedanolide (2), from the sponge *Mycale adhaerens* [2]. The amount isolated was

sufficient for only limited studies on the biological profile in the course of which a unique binding to the 60S ribosome subunit [3b,4] was demonstrated. As a result of this binding, peptide elongation in eukaryotic cells was efficiently inhibited. While several compounds exhibiting specificity for prokaryotes were identified, it is noteworthy that 13-deoxytedanolide (2)is the first known macrolide to be selective for eukaryotes [3]. This selectivity for eukaryotes and archaea was rationalized by Moore et al. who successfully co-crystallized 13-deoxytedanolide (2) with the 50S ribosome subunit of *Haloarcula marismortui* [4]. The binding location was identified as the E site of the ribosome. More recently, Ireland isolated tedanolide C (3) from the marine sponge Ircinia sp. [5] in 2005 and McKee et al. isolated the group of candidaspongiolides (4, 5) from the sponge of the genus *Candidaspongia* in 2007 [6]. Degradation studies were also performed on 13-deoxytedanolide by Fusetani to probe the structure-activity relationships [3]. The first functionality targeted was the epoxide. Unfortunately, attempts to remove it only resulted in a skeleton rearrangement

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Fig. 1. The family of tedanolides.

and the formation of a furan (Scheme 1), and thus, left open the question of its significance for the biological activity.

One striking feature of this family resides in their structure, raising some questions on their biosynthesis. Indeed, to the best of our knowledge, a macrolactone with a primary linkage was unprecedented. Thus, the hypothesis of a different lactone as the direct PKS product was envisioned. On the synthesis perspective, tedanolide is prone to retro-aldol degradations due to the numerous β -hydroxyketone moieties. In addition, the epoxide containing side chain is acid sensitive.



Scheme 1. Fragmentation of 13-deoxytedanolide under acidic conditions.

Since the initial isolation report, tremendous efforts were devoted to the synthesis of tedanolide. These efforts culminated with the first total synthesis of 13-deoxytedanolide (2) by Smith et al. [7] in 2004, followed three years later by the synthesis of tedanolide (1). Their strategy was designed to give access to both compounds in a unified approach. In 2005, a second synthesis of 13-deoxytedanolide was reported by the Roush group [8]. The first total synthesis of tedanolide was achieved by the Kalesse group [9a] in 2006. Fragment syntheses that lead to valuable insight into the subtle synthetic problems of this class of compounds will be covered as well.

2. Results and discussion

2.1. The common approach to the tedanolides by Smith

As already stated, Smith developed a unified strategy to synthetize both tedanolide (1) and 13-deoxytedanolide (2) from a common late stage intermediate. Retrosynthetic disconnections at the macrolactone linkage and at the C11–C12 bond gave rise to the northern and the southern fragments. While the northern fragment is identical for both compounds, simple variation of the southern fragment was required. Coupling by opening an epoxide or by displacement of an iodide would generate tedanolide (1) or 13-deoxytedanolide (2), respectively (Fig. 2). The use of a dithiane anion reversed the reactivity of the aldehyde for the coupling to proceed and protected the carbonyl from possible hemiketal formation. This hemiketal was indeed detrimental as it prevented Roush from completing the synthesis of tedanolide. Two different routes were reported by Smith for the southern fragment. The first generation [6] disclosed for the synthesis of 13-deoxytedanolide (2) was linear and closely similar to the one developed by Roush for his synthesis [8]. The second generation [7c] developed for tedanolide (1) was more convergent and more efficient (Scheme 2).

One of the main challenges encountered during the syntheses was the protecting group strategy. The change from DEIPS to SEM for C17 allylic alcohol between the two routes is noteworthy. The SEM was easily deprotected selectively in presence of a TIPS and TBS and was also more stable than the DEIPS.

2.2. Synthesis of the first-generation southern fragment

Starting from alcohol 6 standard transformations generated acetate 8. Next the terminal double bond was



Fig. 2. Common strategy for the synthesis of tedanolide and 13-deoxytedanolide.



Scheme 2. First-generation approach of the synthesis of the southern fragment of 13-deoxytedanolide.

selectively oxidized and subjected to a Roush crotylation which proved to be more appropriate for this transformation than the Brown protocol. Indeed, the boronate was more labile and thus provided higher yields in the hydrolytic work up. After protecting group manipulations and oxidation, the Z-alkene could be introduced via Wittig olefination. Finally, saponification and iodination provided the southern segment.



Scheme 3. Second-generation approach of the synthesis of the southern fragment of tedanolide.



Scheme 4. Synthesis of the northern fragment of tedanolide.

2.3. Synthesis of the second-generation southern fragment

The second-generation synthesis disconnected the southern fragment at the C17–C18 bond. The coupling proceeded accordingly to Yonemitsu's study of this transformation. Starting from the protected Roche aldehyde, an alkyne was generated via the Corey–Fuchs protocol. Regioselective *E*-vinyl iodide formation was achieved by stannyl cupration to provide **13**. Several conditions were investigated in order to obtain only the desired isomer (Schemes 3 and 4).

The C12–C17 aldehyde synthesis featured a Brown crotylation and an Evans aldol condensation. The subsequent coupling directed by the inherent properties of the aldehyde yielded **14** in a 6:1 diastereoselectivity. The southern fragment **17** was then completed by a *Z*-selective Wittig olefination and a Fraser-Reid epoxide formation.

2.4. Synthesis of the northern fragment

In a similar fashion to the southern fragment, the northern fragment **24** [7] was assembled by the addition of a vinyl iodide to an aldehyde. The vinyl iodide **21** was generated as previously from an alkyne and a hydrostannylation. Iterative Evans aldol reactions provided the C1–C7 aldehyde intermediate **22**.

2.5. Coupling of the northern and southern segments for the tedanolides

The endgame strategy for 13-deoxytedanolide (Scheme 5) and tedanolide (Scheme 6) are closely related and only one will be described herein. The

dianion of the northern fragment 24 (dithiane and free primary alcohol) was used for the key coupling which proved to be high yielding (80% BORSM). As previously stated, the dithiane was seen as a protecting group of the carbonyl to prevent hemiketalization. The downside of this strategy was the dithiane sensitivity to common oxidation methods. This issue was addressed by the development of Tishchenko oxidation. This specific oxidation promoted by samarium^{II} relied on an internaloxidation-reduction in the presence of a βhydroxyketone (26). The resulting ester 27 was hydrolyzed and subsequent macrolactonization of the triol (C15, C17 and C29) selectively closed on the less hindered primary alcohol to provide 28. Diastereoselective epoxidation of 30 directed by the C17 allylic alcohol (Henbest effect) and global deprotection finalized the synthesis. The longer linear sequence was of 31 steps with an overall yield of 0.31% (Scheme 7).

2.6. The Roush synthesis of 13-deoxytedanolide

Roush envisioned a disconnection between C12 and C13 (Fig. 3). This approach should allow access to both tedanolide and 13-deoxytedanolide from the same precursor [8]. The configuration at C15 was chosen to be anti with respect to the configuration at C14 in order to maximize the Felkin enhancing effect of both stereocenters. Additionally, they decided to carry the functionality at C5 as the ketone through the synthesis. Having recognized that mild deprotection conditions were needed for the ester moiety, they employed an allyl group and the Alloc protecting group for the C29 primary alcohol. Thus, both could be removed simultaneously (Fig. 4).



Scheme 5. Endgame of the Smith's 13-deoxytedanolide synthesis.

2.7. Synthesis of the C1–C12 segment

The Roush synthesis of the C1–C12 segment began with a Noyori reduction of keto ester **38** and subsequent methylation. A sequence of reduction, acetylation and regioselective reductive acetal opening provided after oxidation compound **42**. Allyl stannylation, methylation and oxidative cleavage of the double bond provided aldehyde **44** which was transformed to allyl ester **45** through Pinnack oxidation, Mitsunobu esterification and oxidation of C11 to



Scheme 6. Endgame of Smith's tedanolide synthesis.

the ketone. The key aldol coupling proceeded in high yield to provide the northern fragment **36**.

2.8. Synthesis of the C13–C23 segment

One of the key issues in this segment synthesis was the appropriate introduction of the protecting groups. In particular the introduction of the Alloc group was achieved at a relative late stage since it would interfere with oxidative transformation employed in this segment synthesis. Therefore, the BEC group was exploited as a transient protecting group for the primary alcohol.

Starting from the commercially available Roche ester (Scheme 8), Wittig olefination followed by Dibal-H



Scheme 7. Synthesis of the C1-C12 segment of tedanolide.

reduction generated the allylic alcohol **6** required for the stereoselective introduction of the epoxide. An Evans aldol reaction using (S)-**56** established olefin **51** which could be transformed to the double acetate



Fig. 3. Retrosynthetic disconnection of tedanolide in the Roush synthesis.

protected intermediate **53**. Introduction of the C12–C22 double bond was performed with standard conditions. Finally, the pivotal introduction of the Alloc group was achieved by treatment of **54** with allyl alcohol and MeMgBr. This simultaneously liberated diol was then transformed to the corresponding aldehyde through $Pd(OAc)_4$ cleavage.

2.9. Endgame of the synthesis

The aldol coupling was effectively achieved using LiHMDS in THF (Scheme 9). Upon deprotection of the C15 hydroxyl group, intermediate **59** spontaneously cyclized to form an undesired and stable



Fig. 4. Retrosynthesis of tedanolide.

hemiketal. The deoxygenation of C13 allowed the equilibrium with the acyclic ketone to exist, and thus, to be further processed. However, this prevented the synthesis of tedanolide (1). Oxidation of the open form drove the equilibrating mixture to the triketone **60**. Simultaneous allyl ester and Alloc deprotection to provide the seco acid which was followed by a modified Yamaguchi macrolactonization yielded **61** in 31% over two steps. Finally, global sillyl deprotection provided (+)-13-deoxytedanolide (**2**).

2.10. The Kalesse synthesis of tedanolide

As a consequence of the labile epoxide moiety in tedanolide (1) the Kalesse group decided to introduce this functional group at the last step of the synthesis.

As in all other syntheses of the tedanolides an essential issue was the appropriate choice of orthogonal protecting groups for the C29 alcohol and the carboxylate. After substantial variations, the monomethoxytrityl group (MMTr) was identified to be an ideal choice for C29 hydroxyl since its installation and chemical stability were compatible with the operations employed. Additionally, the very mild conditions for its removal, namely treatment with hexafluoro isopropanol allowed removal even in the presence of unprotected β -hydroxyketones.

2.11. Synthesis of aldehyde 65

The originally published route had its drawbacks due to a relatively greater number of steps and occasionally uncontrolled migration of protecting groups. These disadvantages led to an improved synthesis of the C13–C23 aldehyde **65**, using a more convergent aldol coupling between C16 and C17 that paralleled the fragment synthesis proposed by Loh. The synthesis commenced with trityl protected Roche aldehyde **66** and subsequent olefination to yield alkene **67**. The acid catalyzed cleavage of trityl ether and subsequent Swern oxidation provided an aldehyde which was directly subjected to Wittig olefination providing the unsaturated ester **68**. Dibal-H reduction and MnO₂ oxidation provided aldehyde **69** (Scheme 10).

Ketone **72** was synthesized in four steps according to the route described by Loh. An *anti*-selective aldol addition between **72** and **69** set the desired *anti*-relation between the C17 hydroxy and the C16 hydroxymethyl group. The diastereomeric ratio, however, was only of 2:1 in favor of the desired isomer **73**. Dibal-H reduction provided exclusively *syn*-diol **74** (dr > 95:5). The TBS group was removed with TBAF. The three alcohol moieties were selectively protected with an MMTr group on the primary alcohol, a TBS group on the allylic alcohol and the closure of the PMP acetal on the third. Reductive opening of this acetal with Dibal-H



Scheme 8. Synthesis of the C13-C23 segment.

followed by oxidation with TPAP/NMO provided the southern fragment **65** (Scheme 10).

Aldol reaction. Screening of different bases for the enolization of **64** was needed for the pivotal aldol coupling with aldehyde **65**. KHMDS was identified to provide the highest selectivities for obtaining the Felkin product **77** (Scheme 11). To liberate the seco acid the monomethoxytrityl group was removed with hexa-fluoro isopropanol and the allyl group by palladium. The Yamaguchi protocol which was previously employed in both syntheses of 13-deoxytedanolide (**2**) provided 10-35% of the undesired 14-membered macrolactone **79** [9e].

To avoid the undesired lactonization the secondary alcohol at C13 was protected with TBS triflate. The moderate yield of 55% in the conversion of 77-80 could be attributed to considerable retro aldol of 77.

After cleavage of the allyl ester, the free carboxylic acid was subjected to different lactonization methods. The Mitsunobu protocol led to the desired macrolactone **81** in good yields (Scheme 12).

The oxidation state manipulation required deprotection of the PMB and TES groups followed by oxidation. Global silyl deprotection with $3HF \cdot Et_3N$ provided **82** after five days (Scheme 12). The final epoxidation could be achieved with substoichiometric amounts of *m*-CPBA at -45 °C, taking full advantage of the directing Henbest effect.

2.12. The Yonemitsu approach

Early contributions by Yonemitsu were based on computational analyses of the seco acid **84** and unraveled the cyclic acetals to induce the desired conformation for the



Scheme 9. Endgame of Roush's synthesis of 13-deoxytedanolide.

lactonization. The aldol reaction between **86** and **87** was an additional example of how subtle differences can dominate the outcome of synthetic strategies. Indeed, the aldol reaction using LiHMDS provided a 1:1.2-1:1.9 mixture of diastereoisomers in favor of the undesired isomer. Fortunately, the isomers could be separated by chromatography and macrolactonization was achieved by using the Yamaguchi protocol in 87% yield (Fig. 5).

Different disconnections can be envisioned to generate **86**. Initially, **86** was generated from the coupling of known vinyl iodide **89** and aldehyde **88** (Fig. 6). The coupling, however, suffered from low selectivity and irreproducibility. The synthesis of the northern fragment was then reviewed, this time highlighted by a tin triflate based *syn*-aldol pioneered by Paterson between **90** and **91**.

The pivotal Paterson aldol provided the adduct in 80% yield with a diastereoselectivity greater than

20:1 (Scheme 13). After protecting group manipulations and oxidation of **93**, a Wittig homologation provided the carbon skeleton of the C1–C11 fragment. Subsequent dihydroxylation followed by DDQ treatment provided the first acetal unit. Finally, the methyl ketone was introduced through methyl lithium addition and oxidation.

The C13–C23 fragment was further disconnected at the C17–C18 bond with the addition of vinyl lithium **97** to aldehyde **96** (Scheme 14). The synthesis of **96** began with a TiCl₄-mediated addition of oxazolidinone (*R*)-**56** to the Roche ester-derived aldehyde **98**. Reduction and acetalization provided **100**. Reductive opening of the acetal followed by TBSCl protection and oxidative cleavage of the double bond provided aldehyde **96**.

In contrast to the previously discussed syntheses Jung, Miyashita and Loh mainly focused on providing



Scheme 10. Synthesis of fragment 65.

segment syntheses and new approaches toward tedanolide (1). The groups of Jung and Miyashita synthesized the northern and southern fragments, respectively. Loh et al. reported the synthesis of the northern and the southern hemispheres as well as the fragment coupling via an aldol reaction to provide the corresponding seco acid **126**.

Additionally, the approaches of Jung and Miyashita also aimed on developing an efficient synthetic route flexible enough to provide access to 13-deoxytedano-lide (2) and tedanolide (1).

2.13. The Miyashita approach

Miyashita et al. published the stereoselective synthesis of the C13-C23 segment in 2005 [11a]. Their retrosynthetic analysis (Fig. 7) paralleled the one presented by Roush and Lane [8], Hassfeld and Kalesse [9] and Yonemitsu et al. [10]. The first retrosynthetic disconnection was also made at the macrolactone linkage. Further disconnection divided the seco acid into the C13–C23 fragment **102**, already containing the labile epoxide functionality, and the C1–C12 segment **103**. Both segments were envisioned to be assembled by an aldol under Felkin–Anh control [11a].

Miyashita et al. designed a synthetic route for the C13–C23 segment **102** starting from the chiral unsaturated ester **105** in 23 linear steps. The main drawback was the linearity of the synthesis. However, it is interesting to note that the stereoselective synthesis was achieved without the use of any aldol reaction. Instead, the strategy is highlighted by two stereoselective



Scheme 11. Aldol coupling and lactonization.

epoxidation reactions of trisubstituted olefins (epoxidation/epoxide opening) and a stereospecific $S_N 2'$ methylation reaction of a *trans*- γ , δ -epoxy-*cis*- α , β unsaturated ester **104** (Scheme 15).

The C13–C21 portion was constructed from the chiral unsaturated ester **105** via Sharpless asymmetric epoxidation followed by epoxide opening using

vinylmagnesium chloride and copper(I)bromide dimethyl sulphide complex. The *E*-configured double bond of the C18–C19 moiety was installed via Wittig reaction. To introduce an α -secondary methyl group at the C20 position stereoselectively, the use of the stereospecific S_N2' methylation reaction developed by Miyashita et al. was applied [11b]. For this purpose,



Scheme 12. Cyclization of hydroxy acid 80.



Fig. 5. Lactonization by Yonemitsu.

the requisite ester **104** was synthesized via the Horner–Wadsworth–Emmons reaction with the Ando reagent (Scheme 15). This reaction not only installed the methyl group at the C20 but also opened the epoxide at the same time to obtain the C17 hydroxyl stereoselectively.

To obtain the southern fragment **102**, the C13–C21 polypropionate chain was extended via a Z-selective Wittig reaction. After several protecting group manipulations and stereoselective epoxidation of the C18–C19 alkene with *m*-CPBA, a protection–oxidation sequence yielded **102**.

2.14. The Loh approach

Loh and Feng [12] reported on the synthesis of the northern hemisphere **114** and the southern hemisphere **113** of tedanolide (1) as well as the fragment coupling via aldol reaction to complete the carbon backbone. However, the successful total synthesis of tedanolide (1) has not been reported yet.

The retrosynthetic analysis of tedanolide (1) is outlined in Fig. 8. Disconnection at the C29 lactone and the C12, C13 aldol units created the two subunits **113** C13–C23 and **114** C1–C12.



Fig. 6. Retrosynthetic disconnection of tedanolide according to Yonemitsu.



Scheme 13. Synthesis of the C1-C12 segment.



Scheme 14. Synthesis of the C13-C23 segment.



Fig. 7. Retrosynthetic analysis of tedanolide by Miyashita et al.

The key step of the southern fragment **113** [12] synthesis displayed a boron-mediated aldol reaction developed by Paterson and Tillyer [13] between the α , β -unsaturated aldehyde **115** and ketone **72** in very good yield and selectivity (95% yield, major:minor = 85:15). It is interesting to note that Kalesse et al. followed the same strategy using the same ketone **72** to assemble the carbon backbone of the C13–C23 fragment. However, in the case of the Kalesse synthesis, another aldehyde was used and lower yields and stereoselectivity were obtained (68%, major:minor = 2:1).

Ketone **72** could be easily synthesized from the commercially available Roche ester via aldol reaction with methyl acetate and several oxidation and protecting group manipulations. Aldehyde **115** was synthesized from the readily available (*S*)-methyl-3-hydroxy-2methylpropionate as reported by Paterson and Tillyer [13] (Scheme 16).

Further elaboration of fragment C13-C23 was effectuated with a Wittig reaction to install the C21-

C22 Z-olefin. After protecting group transformations, the C18–C19 epoxide was generated with *m*-CPBA and **121** was obtained in 54% as a single diastereomer (Scheme 17). It is the first example of a late stage epoxidation of these substrates. Indeed, it was initially expected to give a mixture of diastereomers or even the undesired isomer [19]. Gratifyingly, the desired product was formed exclusively.

The C1–C12 subunit [12] was generated from the intermediate **122** which could be obtained from the coupling of the two precursors **117** and **118** (Scheme 18). The key feature also presents a *syn*–*syn* boron-mediated aldol reaction developed by Paterson and Tillyer [13].

Ketone **117** was synthesized from the Roche ester via Wittig reaction and Sharpless asymmetric dihydroxylation to incorporate the alcohol functionalities at C2 and C3. The *t*-butyl ester was selected due to its stability toward nucleophilic attack (Scheme 19).

The coupling of the northern and southern fragments was achieved via an aldol reaction. Optimization



Scheme 15. Synthesis of the C13-C23 fragment of tedanolide by Miyashita et al.

of this pivotal transformation identified $(c-hex)_2BCl$ as a Lewis acid to provide the highest yields and diastereoselectivities.

2.15. The Jung approach

Jung et al. proposed in their approach of tedanolides 1 and 2 the preparation of the C1–C12 fragment [14] via a stereoselective non-aldol aldol process, which was developed in their laboratories. The reaction involves the internal hydride migration from an alcohol or silyl protected alcohol to open an epoxide with inversion of configuration at the epoxide center. As the product is already protected during the course of this transformation, further modifications are not required in contrast to standard aldol reactions.

The retrosynthetic disconnection of tedanolide's backbone began with the cleavage at the lactone and scission at the C12–C13 bond to generate the common precursors **126** C1–C12 fragment and **125** C13–C23 fragment (Fig. 9). These intermediates should have been coupled either by an aldol reaction of the

aldehyde **125** (for tedanolide (1)) or an alkylation of the tosylate derived from **125** (for 13-deoxytedanolide (2)).

The northern fragment **126** [14] was generated from the aldol coupling of the two intermediates **127** and **128** with TiCl₄ and Hunig's base (Scheme 20). It is worth mentioning that only the desired Felkin product was obtained in good yield. Other coupling reagents were unsuccessful.

After protection of the free hydroxyl group and reduction of the ketone by L-Selectride compound 132 was obtained containing all the required chiral centers of the northern fragment of the tedanolides 1 and 2 (Scheme 21).

The precursor of **127**, namely ester **129** was prepared from commercially available L-ascorbic acid in three steps via diol protection with 2,2 dimethoxypropane, cleavage of the alkene and esterification with ethyl iodide [14]. The desired ketone **127** was obtained through reduction, epoxidation and regioselective epoxide opening with methyl magnesium bromide and copper iodide (Scheme 22).



Fig. 8. Retrosynthesis of tedanolide by Loh et al.

(Scheme 23) The synthesis of aldehyde **128**, already containing the required stereogenic centers from C5 to C11, started from the commercially available Roche ester **131** and was completed in 13 steps. *E*-selective Wittig reaction was applied to install the C18–C19 olefin. NBS-promoted cyclization of the free homoallylic alcohol to the double bond generated the bromotetrahydrofuran **136** (which demonstrated the utility of

this protecting group as a masked homoallylic ketone). The protection of the alkene functionality was necessary to permit the application of the non-aldol aldol process. Thereby problems associated with allylic epoxide rearrangements were avoided. A second *E*-selective Wittig olefination, reduction to the allylic alcohol and Sharpless asymmetric epoxidation led to intermediate **130**. The crucial non-aldol aldol



Scheme 16. Boron-mediated aldol reaction.



 $\begin{array}{c} TBSO \\ \hline t - BuO_2C \\ \hline \overline{O}Me \\ \hline TBSO \\ \hline TESO \\ \hline 0 \\ 123 \\ \hline TESO \\ \hline 0 \\ PMB \\ \hline TESO \\ \hline 124 \\ \hline TESO \\ \hline 0 \\ PMB \\ \hline TESO \\ \hline 0 \\ TESO \\ \hline 0 \\ PMB \\ \hline TESO \\ \hline 0 \\ TESO \\ \hline 0 \\$

Scheme 19. Fragments coupling by Loh et al.

rearrangement of the epoxide provided the aldol product **137** as a single diastereomer.

2.16. The Iqbal approach

A different synthetic plan was presented by Iqbal et al. [15] who reported a general approach for the synthesis of the C3–C16 segment of tedanolide (1) and 13-deoxytedanolide (2). The central idea was to introduce a C12–C13 double bond of **138** through metathesis and to transform it depending on the natural product to be synthesized, either to saturated hydrocarbon **139** for 13-deoxytedanolide (**2**) or the corresponding epoxide **140** in the case of tedanolide (Fig. 10).

The synthesis started with the reductive transformation of **141** and subsequent acetalization. Reductive opening, oxidation and Wittig olefination provided the corresponding extended ester **144**. A second reduction/oxidation sequence finally provided the corresponding aldehyde which was subjected to an Evans aldol reaction (Scheme 24).

Liberation of the aldehyde **146** and a second Evans aldol reaction provided stereoselectively the



Fig. 9. Retrosynthesis of the tedanolides by Jung et al.



Scheme 20. Aldol coupling of 127 and 128 with TiCl₄ and Hunig's base.



Scheme 21. Protection of the hydroxyl group and reduction with L-Selectride.



Scheme 22. Synthesis of ketone 127.

intermediate 147 (Scheme 25). A sequence of PMPacetalization/Dibal-H opening provided the precursor 149 for the pivotal cross metathesis reaction. During the course of their investigations it appeared that the free C5 allylic alcohol was necessary for the reaction. The final transformations for the different intermediates could be achieved through either Dess-Martin oxidation and NiCl₂-catalyzed reduction (139) or epoxidation of the allylic alcohol (140).

2.17. Myriaporones

Ten years after the discovery of the first tedanolide (1), Rinehart isolated from *Myriapora truncata* a closely related family of natural products, the myriaporones [16]. It appears that myriaporone 1 (151) and 2 (152) are isolation by-products of the natural compound, myriaporone 3/4 (153 and 154). Myriaporone 2 (152) arises from the opening of the epoxide by the C7 carbonyl in myriaporone 1 (151). Myriaporone 3/4 exists as a dynamic equilibrium between a closed and an open form due to the lability of the hemiketal moiety. The small amount isolated did not allow the determination of the stereochemistry at C5 and C6 at

the time and they were elucidated by total synthesis. Their interesting biological activity (IC₅₀ value of 100 ng/mL against L-1210 murine leukemia cells) as well as the similarities of their structure to the C10–C23 portion of tedanolide (1) suggests that the myriaporones are in fact naturally occurring analogues. The myriaporones would then share the same mode of action as tedanolide and their simpler structure renders them more attractive as drug candidates. Recently, a study established that like tedanolide, myriaporone 3/4 is a potent protein synthesis inhibitor selective for eukaryotes [17]. Surprisingly, despite the numerous methodologies developed for the synthesis of tedanolide, few groups applied them for the synthesis of myriaporone (Fig. 11).

2.18. The Yonemitsu approach

As previously mentioned, the stereochemistry at C5 and C6 was not assigned at the time of the isolation. In light of the structural similarities between myriaporone and tedanolide, one could have assumed the stereochemistry of these two centers to be identical to the corresponding carbons of tedanolide (C13



Scheme 23. Synthesis of aldehyde 128.



Fig. 10. The Iqbal approach.

and C14). Yonemitsu, however, hypothesized the opposite configuration for C6 (C14 of tedanolide) [10i]. He also altered his route to the tedanolide and developed a new synthesis toward myriaporone 3/4. His retrosynthesis was based on the coupling of known vinyl iodide **157** and aldehyde **156**. The C5–C9

aldehyde was generated from intermediate **158** which was developed for his synthesis of erythronolide A [10i]. The starting material for this synthesis was D-glucose (Fig. 12).

The key methodology applied in this study was the regioselective opening of MP acetal 160 with MgBr₂



Scheme 24. Synthesis of 145.



Scheme 25. Synthesis of 139 and 140.

and SnBu₃H [10i]. The opening selectivity is governed by a five-membered ring chelation of the substrate, and thus opening occurs at the more hindered carbon. This is a good complement to the Dibal-H opening of acetals (Scheme 26).

The total synthesis was not completed and only the intermediate **155** was generated. As it is now proven that his assumption of the stereochemistry was not

correct at C6, his synthesis will need to be revised to generate the natural product.

2.19. The Taylor approach

The first approach envisioned for the introduction of C5–C9 hydroxypropionate unit was two successive stereocontrolled allylations (Fig. 13) [17a]. The known



Fig. 11. The family of myriaporones.



Fig. 12. The Yonemitsu approach of myriaporone 3/4.



Scheme 26. Opening of MP acetal 160.

intermediate **162** was in turn generated via an alternative approach using a thionyl chloride rearrangement (Scheme 27). The epoxide was chosen to be introduced prior to C9 installation even though such an early epoxidation was expected to render the synthetic intermediates more sensitive. This choice was motivated by the report depicting *anti*-selectivity for epoxidation of secondary allylic alcohols by Sharpless et al. [18].

Although the thionyl chloride rearrangement mechanism was already known, its application in synthesis was not common [17b]. The key reaction depended upon an intramolecular cyclic rearrangement to provide the primary allylic chloride selectively. The zirconium-mediated allylation, however, was not selective due to the steric hindrance generated by the α -methyl of the aldehyde.

Despite the lower activity of myriaporone 1 (**151**) compared to myriaporone 3/4, Taylor also decided to target the former [17c]. With C6 being sp² hybridized in myriaporone 1, only the stereochemistry at C5 remained unknown, and thus, only two diastereomers needed to be synthesized (Fig. 14).



Fig. 15. Retrosynthesis of myriaporone 3/4.

The C5–C7 β -hydroxyketone was masked as an isoxazoline. Formation of the C6–C7 bond was envisioned to be made by a homoallenylation.

The main drawback of this route was the sensitivity of aldehyde **167**. It was generated via an Evans aldol reaction using a chiral auxiliary which proved to be selective and high yielding. C6–C7 bond formation was attempted using Nozaki–Hiyama–Kishi coupling and several homoallenylation conditions. The best results were obtained using Brown conditions giving only a moderate yield. The isoxazoline in **166** was then introduced in a 3:1 selectivity via an inverse dipolar cycloaddition.

The amount of material generated was not sufficient to finish the synthesis. Myriaporone 1 was unexpectedly generated during the synthesis of myriaporone 3/4 (vide infra) [17d] (Fig. 15).

Due to the excessive sensitivity of the epoxyaldehydes and encouraged by the promising epoxidation of Loh [12a], the synthetic plan for myriaporone 3/4 was revised as well as the timing of epoxidation.



Scheme 28. Toward myriaporone 1.

The C5–C9 hydroxypropionate unit of **171** was installed by two iterative Evans aldol reactions. The isoxazoline was then introduced without any diastereo-selectivity. At the time of the synthesis, the stereo-chemistry at C5 was still unknown and both isomers were needed for spectroscopic comparison with the natural product. In later work, the selectivity was increased to 3:1 toward the desired isomer. The Z-olefin was installed via selective Wittig reaction to provide **173** and after protecting group manipulations, the iso-xazoline was reductively cleaved to unmask the β -hydroxyketone. The final key epoxidation was highly

selective according to Yonemitsu's precedence. Myriaporone 3/4 (**153/154**) was thus completed in 2.1% over 25 linear steps.

When acetate-protecting groups were used instead of TBS, the final deprotection led to an elimination reaction, thus providing myriaporone 1 (151) (Schemes 28-30).

2.20. The Cuevas approach

Concomitantly with the Taylor synthesis, Cuevas et al. published their own synthesis in the same journal



Scheme 29. Taylor synthesis of myriaporone 3/4.



Scheme 31. Cuevas synthesis of myriaporone 3/4.

issue [19]. The synthesis developed was also linear and its beginning was similar to the Taylor synthesis. The main difference resided in the choice of an early epoxidation and the installation of the C1–C5 β -hydroxyketone moiety (Scheme 31).

The aldol reaction to form the C4–C5 bond gave **177** with a selectivity of 1:2.5 in favor of the unnatural isomer at C5. This is thus not a valuable selectivity a posteriori. Selective monoprotection discriminating between two secondary alcohols was performed to subsequently oxidize only C7. The carbon backbone was then completed via an Z-selective Wittig olefination to provide **178** and a Grignard addition to the Weinreb amide. Global deprotection gave myriaporone 3/4. The synthesis was completed in 24 linear steps (one step shorter than the Taylor synthesis) but with only 0.3% yield. Partial dehydration on silica gel (not observed by the Taylor group) and subsequent acetate protection of the dehydrated by-product provided them with myriaporone 1. The facility of elimination tends to prove that myriaporone 1 is indeed an isolation by-product of myriaporone 3/4.

3. Conclusion

In summary, the tedanolides are fascinating compounds due to their unprecedented biosynthetic origin, complex architecture and promising biological activity. For more than 20 years, a variety of new strategies to address the challenges offered by tedanolide were developed, completing the established set of tools available for polyketide synthesis.

Additionally, completing the total synthesis of tedanolide opens the door to structure—activity relationship studies, providing analogues that cannot be accessed from natural sources or degradation experiments. *A fortiori*, myriaporone 3/4 appears to be a simpler version of tedanolide that retains the latter's potency, thus suggesting that myriaporone 3/4 is a naturally occurring analogue of tedanolide. Both families represent exciting new leads for therapeutic development which will attract the attention of medicinal chemistry in the future.

References

- F.J. Schmitz, S.P. Gunasekera, G. Yalamanchili, M.B. Hossain, D. van der Helm, J. Am. Chem. Soc. 106 (1984) 7251.
- [2] N. Fusetani, T. Sugawara, S. Matsunaga, H. Hirota, J. Org. Chem. 56 (1991) 4971.
- [3] (a) S. Nishimura, S. Matsunaga, M. Yoshida, H. Hirota, S. Yokoyama, N. Fusetani, Bioorg. Med. Chem. 13 (2005) 449;
 (b) S. Nishimura, S. Matsunaga, S. Yoshida, Y. Nakao, H. Hirota, N. Fusetani, Bioorg. Med. Chem. 13 (2005) 455.
- [4] S.J. Schroeder, G. Blaha, J. Tirado-Rives, T.A. Steitz, P.B. Moore, J. Mol. Biol. 367 (2007) 1471.
- [5] C. Chevallier, T.S. Bugni, X. Feng, M.K. Harper, A.M. Orendt, C.M. Ireland, J. Org. Chem. 71 (2006) 2510.
- [6] T.L. Meragelman, R.H. Willis, G.M. Woldemichael, A. Heaton, P.T. Murphy, K.M. Snader, D.J. Newman, R. van Soest, M.R. Boyd, J.H. Cardellina, T. McKee, J. Nat. Prod. 70 (2007) 1133.
- [7] (a) A.B. Smith III, C.M. Adams, S.A. Lodise Barbosa, A.P. Degnan, J. Am. Chem. Soc. (2003) 350;
 (b) A.B. Smith III, C.M. Adams, S.A.L. Barbosa, A.P. Degnan, Proc. Natl. Acad. Sci. U S A 101 (2004) 12042;
 (c) A.B. Smith III, D. Lee, J. Am. Chem. Soc. 129 (2007) 10957.
- [8] (a) W.R. Roush, G.C. Lane, Org. Lett. 1 (1999) 95;
 (b) W.R. Roush, J.S. Newcom, Org. Lett. 4 (2002) 4739;
 (c) L.D. Julian, J.S. Newcom, W.R. Roush, J. Am. Chem. Soc. 127 (2005) 6186.
- [9] (a) J. Hassfeld, M. Kalesse, Synlett (2002) 2007;
 (b) J. Hassfeld, U. Eggert, M. Kalesse, Synthesis (2005) 1183;
 (c) G. Ehrlich, M. Kalesse, Synlett (2005) 655;
 (d) G. Ehrlich, J. Hassfeld, U. Eggert, M. Kalesse, J. Am. Chem. Soc. 128 (2006) 14038;
 (e) G. Ehrlich, J. Hassfeld, U. Eggert, M. Kalesse, Chem. Eur. J.

(2008) 2232.

- [10] (a) Y. Oikawa, T. Nishi, O. Yonemitsu, J. Chem. Soc., Perkin Trans. 1 (1985).
 - (b) O. Yonemitsu, J. Synth. Org. Chem. 52 (1994) 946;

(c) T. Matsushima, K. Horita, N. Nakajima, O. Yonemitsu, Tetrahedron Lett. 37 (1996) 385;

(d) T. Matsushima, M. Mori, N. Nakajima, H. Maeda, J. Uenishi, O. Yonemitsu, Chem. Pharm. Bull. 46 (1998) 1335;
(e) T. Matsushima, M. Mori, B.-Z. Zheng, H. Maeda, N. Nakajima, J.-I. Uenishi, O. Yonemitsu, Chem. Pharm. Bull. 47 (1999) 308;

(f) T. Matsushima, B.-Z. Zheng, H. Maeda, N. Nakajima, J. Uenishi, O. Yonemitsu, Synlett (1999) 780;

(g) B.-Z. Zheng, H. Maeda, M. Mori, S. Kusak, O. Yonemitsu, T. Matsushima, N. Nakajima, J.-I. Uenishi, Chem. Pharm. Bull. 47 (1999) 1288;

(h) H. Matsushima, H. Nakajima, B.-Z. Zheng, O. Yonemitsu, Chem. Pharm. Bull. 48 (2000) 855;

(i) B.-Z. Zheng, M. Yamauchi, H. Dei, S. Kusaka, K. Matsui,O. Yonemitsu, Tetrahedron Lett. 41 (2000) 6441;

(j) B.-Z. Zheng, M. Yamauchi, H. Dei, O. Yonemitsu, Chem. Pharm. Bull. 48 (2000) 1761;
(k) K. Matsui, B.-Z. Zheng, S.-I. Kusaka, M. Kuroda,

(k) K. Indisut, D. Z. Zheng, S. I. Rusaki, M. Rubaki,K. Yoshimoto, H. Yamada, O. Yonemitsu, Eur. J. Org. Chem. 9 (2001) 3615.

- [11] (a) A. Hirai, A. Matsui, K. Komatsu, K. Tanino, M. Miyashita, Chem. Commun. (2002) 1970;
- (b) Y. Iwata, K. Tanino, M. Miyashita, Org. Lett. 7 (2005) 2341.
 [12] (a) T.-P. Loh, L.-C. Feng, Tetrahedron Lett. 42 (2001) 3223;
 (b) C.-M. Wong, T.-P. Loh, Tetrahedron Lett. 47 (2006) 4485;
- (c) T.-P. Loh, L.-C. Feng, Tetrahedron Lett. 42 (2001) 6001.[13] I. Paterson, R.D. Tillyer, J. Org. Chem. 58 (1993) 4182.
- [14] (a) M.E. Jung, U. Karama, R. Marquez, J. Org. Chem. 64 (1999) 663;
 (b) M.E. Jung, R. Marquez, Tetrahedron Lett. 40 (1999) 3129;
 (c) M.E. Jung, R. Marquez, Org. Lett. 2 (2000) 1669;
 (d) M.E. Jung, C.P. Lee, Tetrahedron Lett. 41 (2000) 9719;
 (e) M.E. Jung, C.P. Lee, Org. Lett. 3 (2001) 333;
 (f) H. Jung, M.E. Yoo, Org. Lett. 9 (2007) 3543;
 (g) M.E. Jung, T. Zhang, Org. Lett. 10 (2008) 137.
- [15] (a) V.K. Nyavanandi, S. Nanduri, R.V. Dev, A. Naidu, J. Iqbal, Tetrahedron Lett. 47 (2006) 6667;
 (b) V.K. Nyavanandi, P. Nadipalli, S. Nanduri, A. Naidu, J. Iqbal, Tetrahedron Lett. 48 (2007) 6905.
- [16] (a) K.L. Rinehart, K. Tachibana, J. Nat. Prod. 58 (1995) 344;
 (b) J.-F. Cheng, J.-S. Lee, R. Sakai, E.A. Jares-Erijman, M.V. Silva, K.L. Rinehart, J. Nat. Prod. 70 (2007) 332.
- [17] (a) R.E. Taylor, J.P. Ciavarri, B.R. Hearn, Tetrahedron Lett. 39 (1998) 936;
 (b) B.R. Hearn, J.P. Ciavarri, R.E. Taylor, Org. Lett. 4 (2002) 2953;
 (c) K.N. Fleming, R.E. Taylor, Angew. Chem. Int. Ed. 43

(c) K.N. Fleming, K.E. Taylor, Angew. Chem. Int. Ed. 45 (2004) 1728;

(d) J. Hines, M. Roy, H. Cheng, C.M. Agapakis, R.E. Taylor, C.M. Crews, Mol. Biosyst. 2 (2006) 371.

- [18] B.E. Rossiter, T.R. Verhoeven, K.B. Sharpless, Tetrahedron Lett. 20 (1979) 4733.
- [19] M. Perez, C. del Pozo, F. Reyes, A. Rodriguez, A. Francesch, A.M. Echavarren, C. Cuevas, Angew. Chem. Int. Ed. 43 (2004) 1724.