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Synthesis of non glycosidic nucleobase-sugar mimetics

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ABSTRACT

Biologically active organic molecules acting as nucleoside mimics are frequently encountered in pharmaceutical research. They are either synthetic heterocycles, which miss the sugar-derived interactions with the active site of the nucleoside-binding protein, or natural products containing a glycosidic linkage, which may cause bioavailability and metabolic stability problems. We report here the concept of synthetic full nucleoside mimics, including both a N-containing nucleobase-like portion and a sugar-like moiety, where the latter consists of 5- and 6-membered carbocycles connected by a more stable and drug-like C–N bond to the nucleobase mimic. Compounds **14**, **16** (indolinones), **21** and **23** (benzimidazolones) have been prepared as model compounds.

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1. Introduction

Mimicry of nucleosides is an active research field, because of the many potential applications in chemical biology and drug discovery. Probably the most popular application consists of ATP-competitive kinase inhibitors [1]. Naturally occurring ATP-competitive inhibitors are usually composed of a heterocyclic, nucleobase-like part connected to a sugar ribose via a glycosidic bond. Apart from several specificity and toxicity issues, the glycosidic bond and the carbohydrate moiety often create stability and bioavailability problems in pharmacologically relevant conditions [2]. Many of the known synthetic ATP-competitive kinase inhibitors consist of heterocyclic structures capable of replacing the nucleobase in the conserved ATP-binding site of the enzymes [3]. The affinity and possibly the specificity of such molecules could be improved by sugar-like appendages capable of engaging the ribose-binding portion of the active site, while being devoid of the above-mentioned stability and bioavailabili-

ty issues connected to glycosidic linkages and sugars. This concept is supported by the synthetic indolocarbazole SRN-003-556 (**1**) (compare with the naturally occurring, glycosidic bond-containing K252a (**2**), Fig. 1), which is an in vivo active, potent kinase inhibitor [4].

Here, we report on the general synthetic approaches capable of affording two classes of molecular structures, bearing 5- and 6-membered carbocyclic polyols of the general formula **A** or **B** shown in Chart 1. These examples contain indolinone (Z=CH₂) or benzimidazolone (Z=NH) rings as the nucleobase decoy. Such heterocycles have been used previously in the synthesis of ATP-competitive kinase inhibitors [5] and represent therefore an interesting model system.

2. Results and discussion

2.1. Epoxide opening

The envisioned modular strategy (Chart 1), allows the access to chemical diversity via the opening of two epoxides, enantiomerically pure **3** [6a] and racemic **4** (The epoxide **4** was obtained by epoxidation of trans 4-*tert*-butyl-dimethyl-silanyloxy)-cyclopent-2-enol [7] (see the

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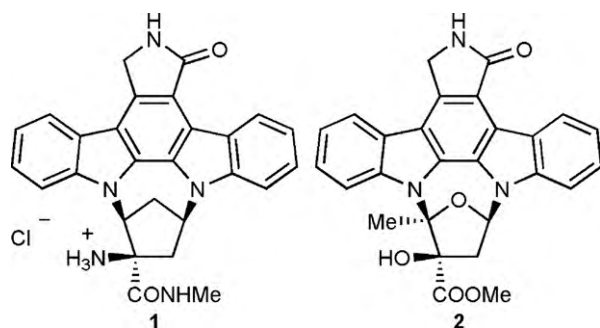


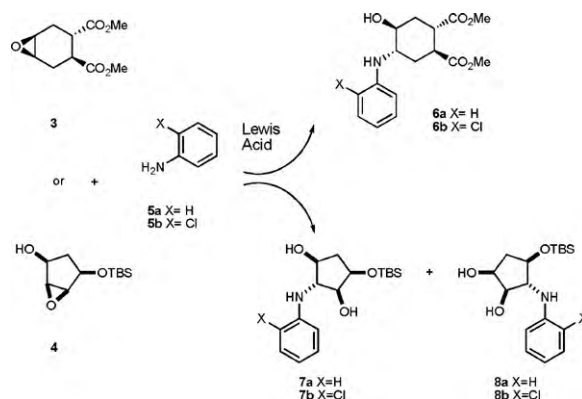
Fig. 1. Structure of SRN-003-556 (**1**) and K252a (**2**).

experimental section) with different anilines. Further N-functionalization and Pd-catalyzed ring formation yield the target nucleobase-sugar mimics. Known epoxide **3** was chosen because it is easily available in enantiopure form and has been used for the synthesis of sugar mimics [6]. Easily available epoxide **4** was introduced as a closer mimic of naturally occurring sugars.

The opening of epoxides with anilines in the presence of Lewis acid promoters is amply documented [8a–g], and a brief screening was performed to identify the best reaction conditions for substrates **3** and **4**, using aniline **5a** or 2-chloroaniline **5b** as nucleophiles (Scheme 1). Results are detailed in Table 1.

Both regioisomers **7** and **8** (structures assigned by nOe experiments) were formed with epoxide **4**, whereas the C2' symmetry axis of compound **3** led only to isomer **6**. Somewhat surprisingly, yields for the reaction with epoxide **3** were consistently higher with 2-chloroaniline **5b** than with **5a**. Both epoxides **3** and **4** (the latter requiring stronger reaction conditions) reacted with good yields in presence of InBr_3 [8a] (entries 3, 7, 8, 9, 13, Table 1) or $\text{Yb}(\text{OTf})_3$ [8b], (entries 2, 6, 12, Table 1). Bi-based catalysts [8c,8d] were less performing under all the examined reaction conditions (entries 1, 4, 5, 10, 11, Table 1).

Eventually, InBr_3 was selected as the catalyst for further experiments. Although its performance was overall similar



Scheme 1. Opening of epoxides **3** and **4** with anilines **5a,b**.

Table 1
Opening of epoxides **3**^[a] and **4**^[b] with anilines **5a,b**.

Entry	Epoxide	Aniline	Promoter	Reaction time (h)	Compound/yield (%) ^[c]
1	3	5a	$\text{Bi}(\text{OTf})_3$	18	6a /55
2	3	5a	$\text{Yb}(\text{OTf})_3$	18	6a /77
3	3	5a	InBr_3	2.5	6a /94
4	3	5b	BiCl_3	72	6b /47
5	3	5b	$\text{Bi}(\text{OTf})_3$	18	6b /65
6	3	5b	$\text{Yb}(\text{OTf})_3$	18	6b /83
7	3	5b	InBr_3	2.5	6b /85
8	3	5b ^[d]	InBr_3	2.5	6b /91
9	4	5a	InBr_3	8	7a /54 8a /22
10	4	5a	$\text{Bi}(\text{OTf})_3$	48	7a /n.i. ^[e] 8a /n.i. ^[e]
11	4	5a	BiCl_3	48	7a /27 ^[f] 8a /23 ^[f]
12	4	5a	$\text{Yb}(\text{OTf})_3$	8	7a /67 ^[f] 8a /15 ^[f]
13	4	5b	InBr_3	8	7b /50 8b /25

[a] Unless otherwise noted, reactions were performed with 0.1 eq. promoter and 1.1 eq. aniline in DCM at RT.

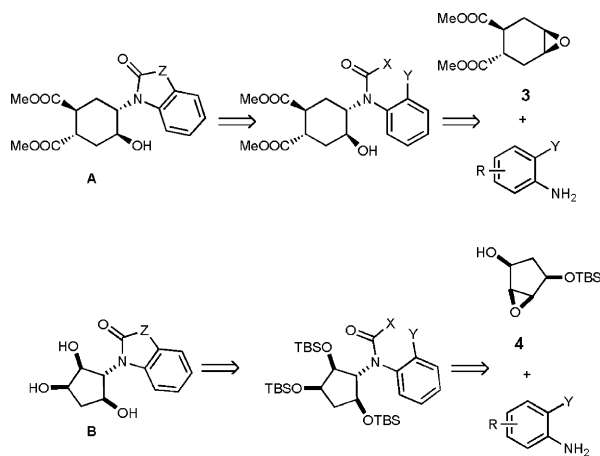
[b] Reactions were performed with 0.5 eq. promoter and 1.5 eq. aniline in CH_2Cl_2 at reflux.

[c] Isolated yields after chromatography, unless otherwise noted.

[d] 1.5 eq of aniline were used.

[e] Not isolated.

[f] NMR yields estimated from crude reaction mixtures.

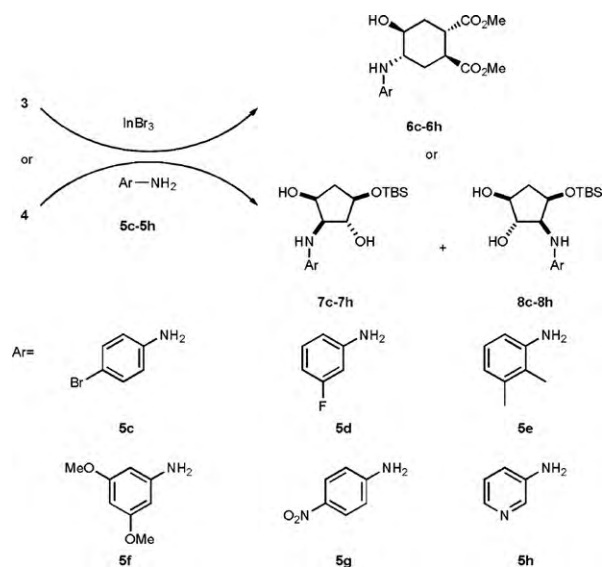


X = CH_2Cl , NH_2 ; Z = CH_2 , NH ; Y = H, Cl;
R = *p*-Br, *m*-F, 2,3-di-Me, 3,5-di-OMe, *p*-NO₂

Chart 1. General structures of the targets and retrosynthetic approaches.

to $\text{Yb}(\text{OTf})_3$, we were often able with InBr_3 to significantly reduce reaction times (compare entries 2 and 3, Table 1). Anilines **5c–h** were then used with the same experimental protocols to test the versatility of our strategy (Scheme 2).

Using epoxide **3**, yields were good to excellent with anilines **5c–f** (entries 1–4), i.e. comparable to those obtained with **5a,b**. Anilines **5g** and **5h** gave moderate yields (entries 5–6), probably due to the poor nucleophilicity of their nitrogen atom. Epoxide **4** confirmed its overall lower activity, even under stronger reaction conditions, and did not react with deactivated anilines **5g** and **5h** (entries 11–12). Yields obtained with anilines **5c–f** were nevertheless good (entries 7–10). Results are reported in Table 2.

Scheme 2. Opening of epoxides **3** and **4** with anilines **5c-h**.

2.2. Synthesis of indolinones **14** and **16**

We then proceeded to the synthesis of indolinones **14** and **16** starting respectively from **6a** and a mixture of compounds **7a** and **8a**. It must be noted that the formation of both regioisomers from the opening of epoxide **4** (Scheme 1) is of no relevance from now on, as the two regioisomers are converted into a single racemate after the first reaction step (step a, Scheme 3).

Sequential protection of the free hydroxyl groups (step a, Scheme 3) and chloroacetylation of the fully O-protected aniline (step b, Scheme 3) afforded respectively **9** and **10** in good to excellent yields. Both compounds could be used without purification in the subsequent cyclization to the indolinone ring using typical Buchwald's conditions [9] (step c, Scheme 3). O-protected indolinones **11** and **12**

Table 2
Opening of epoxides **3**^[a] and **4**^[b] with anilines **5c-h** (Scheme 2).

Entry	Epoxide	Aniline	Reaction time (h)	Compound/yield (%) ^[c]
1	3	5c	15	6c /82
2	3	5d	4	6d /98
3	3	5e	15	6e /97
4	3	5f	5	6f /80
5	3	5g	72	6g /43
6	3	5h	48	6h /40
7	4	5c	48	7c /40 8c /30
8	4	5d	24	7d /64 8d /21
9	4	5e	30	7e /52 8e /14
10	4	5f	24	7f /58 8f /22
11	4	5g	X	7g /n.i. ^d 8g /n.i. ^d
12	4	5h	X	7h /n.i. ^d 8h /n.i. ^d

[a] Reactions were performed with 0.1 eq. promoter and 1.1 eq. aniline in DCM at RT.

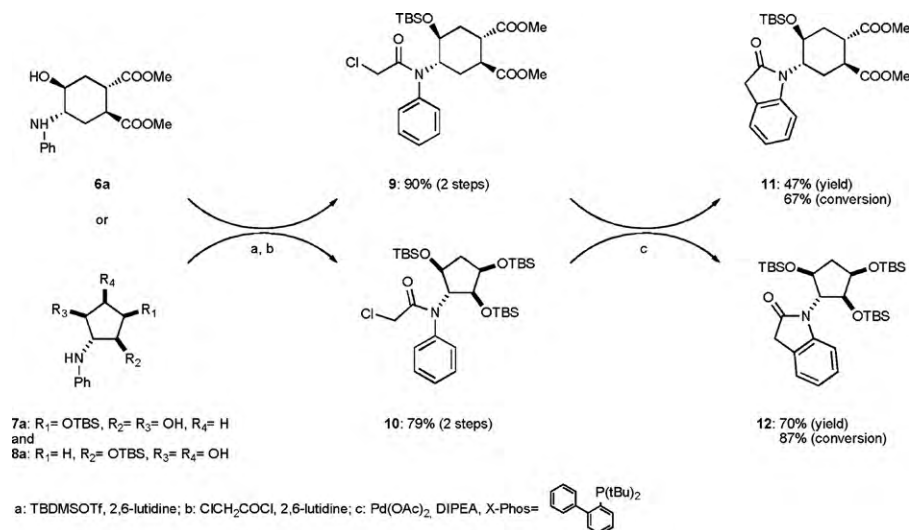
[b] Reactions were performed with 0.5 eq. promoter and 1.5 eq. aniline in CH₂Cl at reflux.

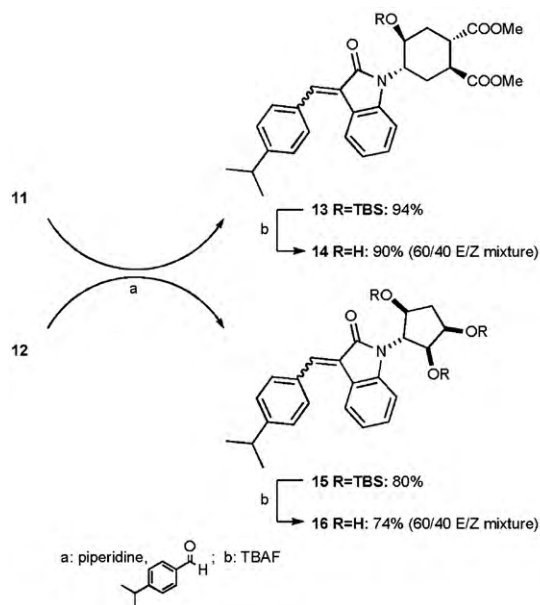
[c] Isolated yields, after chromatographic purification.

[d] Not isolated.

were isolated respectively in 47 and 70% unoptimized yield. With indolinone **11**, 30% of starting material was recovered and could be recycled.

With protected compounds **11** and **12** in hands, we decided to perform an aldolisation on the free CH₂ position of the heterocycle in order to obtain 3-alkylidene products, as similar heterocycles are known to be active on kinases in the literature [10]. Therefore, we performed an aldol condensation (step a, Scheme 4) according to established protocols [10]. 3-(Benzylidene)indolin-2-ones **14** and **16**

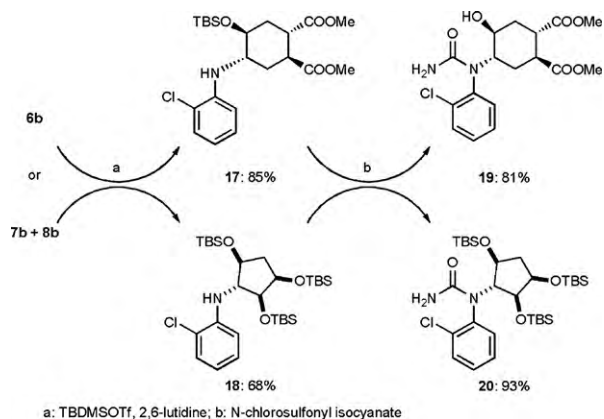
Scheme 3. Synthesis of indolinones **11** and **12**.

Scheme 4. Synthesis of compound **14** and **16**.

were obtained in an overall 85 and 59% yield respectively, and in a $\approx 60/40$ E/Z ratio, after deprotection (step b, Scheme 4). Unfortunately, but not unexpectedly [10], single geometric isomers isolated by chromatography quickly equilibrated to the original E/Z ratio in solution.

2.3. Synthesis of benzimidazolones **21** and **23**

The synthesis of benzimidazolones **21** and **23** (Scheme 6) entailed a key Pd-catalyzed cyclization of intermediate ureas **19** and **20**. Protection of **6b** or of a mixture of compounds **7b** and **8b** (step a, Scheme 5), followed by urea

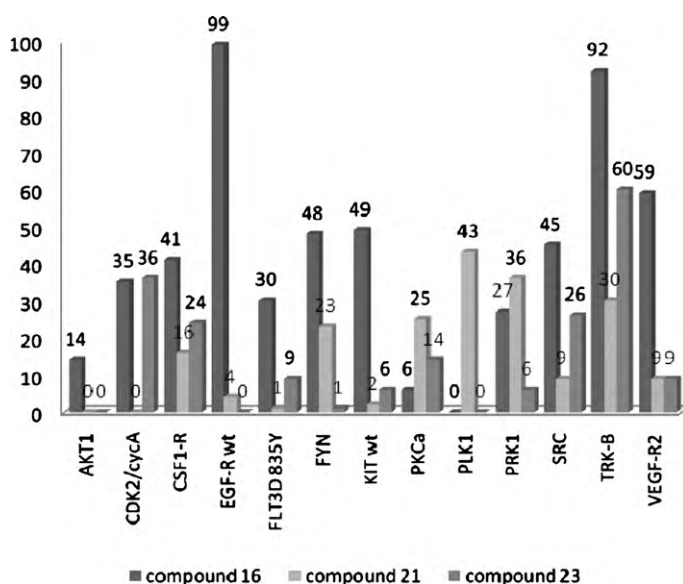
Scheme 5. Synthesis of ureas **19** and **20**.

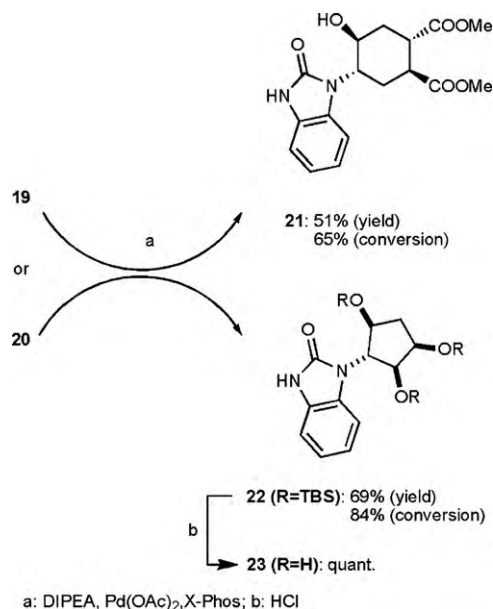
formation with chlorosulfonylisocyanate (step b, Scheme 5) yielded respectively **19** and **20** in good overall yields. It has to be noted that O-deprotection occurred during urea formation from compound **17**, while the same Si-protecting group was not removed from compound **18**.

Cyclization of **19** and **20** was achieved by a slight modification of a strategy, recently proposed by McLaughlin et al. [11], using Pd(OAc)₂ and Buchwald's X-Phos ligand (step a, Scheme 6) [12]. Benzimidazolones **21** and **22** were isolated in good yield and some starting material was recovered to be recycled. Final O-deprotection of compound **22** (step b, Scheme 6) yielded benzimidazolone **23** in quantitative yield.

3. Biology

We have tested compounds **16**, **21** and **23** on a panel of 48 kinases (single point determination, 20 μ M). Best results on selected 12 kinases are reported on Chart 2.

Chart 2. Percentage inhibition of compounds **16**, **21** and **23** on kinases.

Scheme 6. Synthesis of benzimidazolones **21** and **23**.

In particular, compound **16** shows promising results on several kinases (TRK-B and VEGF-R2 in particular). Consequently, we measured the IC₅₀ of compound **16** on several kinases, which obtained a 24 μM IC₅₀ value for both TRK-B and VEGF-R2, and a 50 μM IC₅₀ value for IKK-2.

4. Conclusions

We have devised a practical strategy to tether a 5- and a 6-membered carbocyclic polyol to a heterocyclic structure with the function of a nucleobase decoy. We have shown that this strategy could be extended using different anilines to perform epoxide opening reactions.

The molecules described here, although not yet structurally optimized, are expected to lead to potent and selective ATP-competitive kinase inhibitors, since 3-substituted indolin-2-ones and *N*-monosubstituted benzimidazolones represent a well-established class of tyrosine kinase inhibitors, which exhibit selectivity toward different receptor tyrosine kinases [5].

Indeed, a preliminary set of tests allowed to determine a medium/micromolar activity on several therapeutically relevant kinases for compound **16**, thus providing proof of principle for our mimic concept. The synthesis of a larger set of aryl functionalized indolinones and benzimidazolones bearing variously substituted 5- and 6-membered carbocyclic sugar mimics is ongoing and their test on a panel of different kinases and other nucleotide-binding proteins will be reported in due course.

5. Experimental section

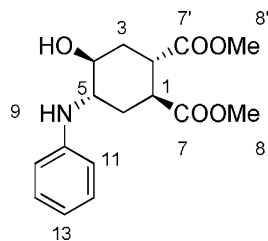
Solvents were dried by standard procedures. Dichloromethane, toluene, methanol, *N,N*-diisopropylethylamine,

piperidine and triethylamine were dried over calcium hydride; *i*PrOH and 2,6-lutidine were dried over activated molecular sieves. Reactions requiring anhydrous conditions were performed under nitrogen or argon. Thin-layer chromatography (TLC) was carried out with precoated silica gel plates. Purifications were performed by flash chromatography carried out with silica gel (230–400 mesh) or by Biotage SP1™ Purification System. Mass spectra were obtained with an ESI apparatus Bruker Esquire 3000 plus. Optical rotations [α]_D were measured in a cell of 1 dm pathlength and 1 mL capacity. ¹H and ¹³C spectra were recorded at 300 K on a 400 MHz spectrometer. Chemical shifts δ for ¹H and ¹³C are expressed in ppm relative to internal Me₄Si as standard. Signals were abbreviated as s: singlet; bs: broad singlet; d: doublet; t: triplet; q: quartet; sp: septuplet; m: multiplet.

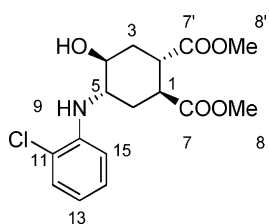
Each molecule was numbered without following the IUPAC numeration, and the numeration we used is reported on schemes for all the new compounds synthesized.

5.1. Epoxide opening: general procedure for compound **3**

To a 0.8 M solution of **3**^[6a] (1 mol equiv.) in CH₂Cl₂ (technical grade) aniline **5a–h** (1.5 mol equiv.) and InBr₃ (0.1 mol equiv.) were added. The reaction mixture was stirred for 2.5 h at room temperature, monitoring the reaction progression by TLC (6/4 petroleum ether/EtOAc). After reaction completion the solution was evaporated and all the compounds were isolated by flash chromatography or by Biotage SP1™.

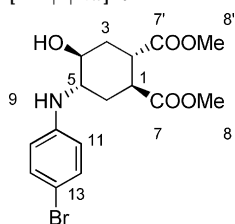
5.2. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-hydroxy-5-(phenylamino)cyclohexane-1,2-dicarboxylate **6a** (Fig. 2. – S1)

Starting from 240 mg (1.14 mmol) of epoxide **3**, we obtained a crude reaction product which was purified by flash chromatography using petroleum ether/EtOAc (60/40) to afford **6a** as a colorless oil (329 mg, 94%). R_f = 0.2 (DCM/EtOAc 65/35). ¹H-NMR (400 MHz, CDCl₃): 1.65–1.72 (m, 1 H, 6ax-H); 1.81–1.89 (m, 1 H, 3ax-H); 2.21–2.30 (m, 1 H, 3eq-H); 2.38–2.43 (m, 1 H, 6eq-H); 3.09–3.16 (m, 1 H, 1-H); 3.25–3.32 (m, 1 H, 2-H); 3.46–3.52 (m, 1 H, 5-H); 3.73 (s, 3 H, 8-H or 8'-H); 3.75 (s, 3 H, 8-H or 8'-H); 3.75–3.81 (m, 1 H, 4-H); 6.73 (d, J_{11–12} = 7.4 Hz, 2 H, 11-H); 6.78 (t, J_{12–13} = 7.4 Hz, 1 H, 13-H); 7.21 (t, J_{11–12} = J_{12–13} = 7.4 Hz, 2 H, 12-H). ¹³C-NMR (100.61 MHz, CDCl₃): 28.3 (6-C); 31.1 (3-C); 39.9 (2-C); 40.3 (1-C); 52.2 (8-C + 8'-C); 54.4 (5-C); 68.6 (4-C); 114.0 (11-C); 118.6 (13-C); 129.5 (12-C); 146.5 (10-C); 174.1 (7-C or 7'-C); 174.5 (7-C or 7'-C). MS (ESI⁺): 308.0 [M + H⁺] [α]_D = +40.0 [c = 0.1, MeOH].



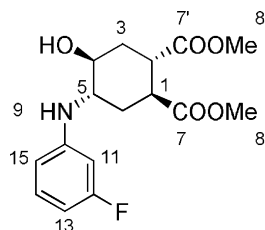
5.3. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(2-chlorophenylamino)-5-hydroxycyclohexane-1,2-dicarboxylate **6b** (Fig. 2. – S2)

Starting from 357 mg (1.67 mmol) of epoxide **3**, we obtained a crude reaction product which was purified by flash chromatography using n-hexane/EtOAc (60/40) to afford **6b** as a colorless oil (520 mg, 91%). $R_f = 0.2$ (CHCl₃/EtOAc 9/1). ¹H-NMR (400 MHz, CDCl₃): 1.68–1.77 (m, 1 H, 3ax-H); 1.80–1.88 (m, 1 H, 6ax-H); 2.29–2.42 (m, 2 H, 3eq-H + 6eq-H); 3.16–3.20 (m, 1 H, 2-H); 3.30–3.33 (m, 1 H, 1-H); 3.53–3.58 (m, 1 H, 4-H); 3.70 (s, 3 H, 8-H or 8'-H); 3.75 (s, 3 H, 8-H or 8'-H); 3.77–3.88 (m, 1 H, 5-H); 6.73 (t, $J = 7.6$ Hz, 1 H, 13-H); 6.88 (d, $J = 8.0$ Hz, 1 H, 15-H); 7.18 (t, $J = 7.6$ Hz, 1 H, 14-H); 7.26–7.31 (m, 1 H, 12-H). ¹³C-NMR (100.61 MHz, CDCl₃): 28.2 (6-C); 31.1 (3-C); 40.2 (2-C); 40.6 (1-C); 52.3 (8-C + 8'-C); 55.5 (5-C); 68.8 (4-C); 112.8 (15-C); 118.8 (13-C); 120.9 (11-C); 127.7 (14-C); 129.8 (12-C); 142.1 (10-C); 173.9 (7-C or 7'-C); 174.1 (7-C or 7'-C). MS (ESI⁺): 705.2 [2M + Na]⁺.



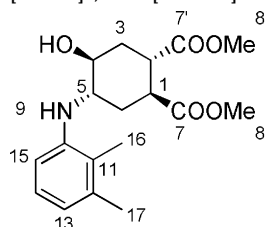
5.4. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(4-bromophenylamino)-5-hydroxycyclohexane-1,2-dicarboxylate **6c** (Fig. 2. – S3)

Starting from 103 mg (0.48 mmol) of epoxide **3**, we obtained a crude reaction product which was purified by Biotage SP1TM using n-hexane/AcOEt (from 100/0 to 20/80) to afford **6c** as a colorless oil (152 mg, 82%). $R_f = 0.3$ (Hexane/EtOAc 7/3). ¹H-NMR (400 MHz, CDCl₃): 1.64 (ddd, $J_{gem} = 13.5$ Hz, $J = 7.2$ Hz, $J = 4.3$ Hz, 1 H, 3ax-H); 1.81 (ddd, $J_{gem} = 13.8$ Hz, $J = 7.4$ Hz, $J = 4.6$ Hz, 1 H, 6ax-H); 2.14 (ddd, $J_{gem} = 13.8$ Hz, $J = 7.8$ Hz, $J = 3.1$ Hz, 1 H, 6eq-H); 2.30 (ddd, $J_{gem} = 13.5$ Hz, $J = 7.8$ Hz, $J = 3.9$ Hz, 1 H, 3eq-H); 2.83 (bs, 1 H, 4-OH); 2.96–3.10 (m, 1 H, 2-H); 3.18–3.26 (m, 1 H, 1-H); 3.32–3.47 (m, 1 H, 4-H); 3.68 (s, 3 H, 8-H or 8'-H); 3.70 (s, 3 H, 8-H or 8'-H); 3.70–3.83 (bs, 1 H, 5-H); 6.53 (d, $J = 8.9$ Hz, 2 H, 10-H); 7.21 (d, $J = 8.9$ Hz, 2 H, 11-H). ¹³C-NMR (100.61 MHz, CDCl₃): 28.2 (3-C); 31.2 (6-C); 39.8 (1-C); 40.2 (2-C); 52.4 (8-C + 8'-C); 53.9 (4-C); 68.3 (5-C); 109.6 (10-C); 115.2 (11-C); 132.2 (12-C); 146.1 (13-C); 174.3 (7-C or 7'-C); 174.8 (7-C or 7'-C). MS (ESI⁺): 386 [M + H]⁺.



5.5. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(3-fluorophenylamino)-5-hydroxycyclohexane-1,2-dicarboxylate **6d** (Fig. 2. – S4)

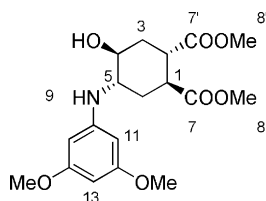
Starting from 100 mg (0.47 mmol) of epoxide **3**, we obtained a crude reaction product which was purified by flash chromatography using n-hexane/AcOEt (from 100/0 to 50/50) to afford **6d** as a colorless oil (151 mg, 98%). $R_f = 0.3$ (Hexane/EtOAc 6/4). ¹H-NMR (400 MHz, CDCl₃): 1.68 (ddd, $J_{gem} = 13.8$ Hz, $J = 7.1$ Hz, $J = 4.3$ Hz, 1 H, 3ax-H); 1.86 (ddd, $J_{gem} = 13.7$ Hz, $J = 7.3$ Hz, $J = 4.6$ Hz, 1 H, 6ax-H); 2.17 (ddd, $J_{gem} = 13.7$ Hz, $J = 8.1$ Hz, $J = 3.8$ Hz, 1 H, 6eq-H); 2.36 (ddd, $J_{gem} = 13.8$ Hz, $J = 8.1$ Hz, $J = 3.8$ Hz, 1 H, 3eq-H); 2.90 (bs, 1 H, OH); 3.08 (ddd, $J = 7.6$ Hz, $J = 7.3$ Hz, $J = 4.4$ Hz, 1 H, 2-H); 3.26 (ddd, $J = 7.4$ Hz, $J = 7.3$ Hz, $J = 4.8$ Hz, 1 H, 2-H); 3.95 (dd, $J = 10.3$ Hz, $J = 6.6$ Hz, 1 H, 4-H); 3.71 (s, 3 H, 8-H or 8'-H); 3.74 (s, 3 H, 8-H or 8'-H); 3.78–3.85 (bs, 1 H, 5-H); 6.36–6.46 (m, 3 H, 11-H + 13-H + 15-H); 7.05–7.12 (m, 1 H, 14-H). ¹³C-NMR (100.61 MHz, CDCl₃): 28.2 (3-C); 31.1 (6-C); 39.7 (1-C); 40.1 (2-C); 52.2 (8-C + 8'-C); 53.8 (4-C); 68.1 (5-C); 100.0 + 104.3 + 109.3 (11-C + 13-C + 15-C); 130.4 (14-C); 148.9 (10-C); 164.2 (d, $J = 244$ Hz, 12-C); 174.2 (7-C or 7'-C); 174.8 (7-C or 7'-C). MS (ESI⁺): 326 [M + H]⁺, 348 [M + Na]⁺.



5.6. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(2,3-dimethylphenylamino)-5-hydroxycyclohexane-1,2-dicarboxylate **6e** (Fig. 2. – S5)

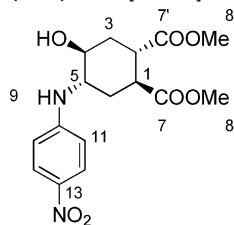
Starting from 107 mg (0.5 mmol) of epoxide **3**, we obtained a crude reaction product which was purified by flash chromatography using n-hexane/AcOEt (from 100/0 to 60/40) to afford **6e** as a brown oil (163 mg, 97%). $R_f = 0.25$ (Hexane/EtOAc 7/3). ¹H-NMR (400 MHz, CDCl₃): 1.71 (ddd, $J_{gem} = 13.6$ Hz, $J = 6.8$ Hz, $J = 4.3$ Hz, 1 H, 3ax-H); 1.88 (ddd, $J_{gem} = 13.8$ Hz, $J = 7.1$ Hz, $J = 4.6$ Hz, 1 H, 6ax-H); 2.05 (s, 3 H, 17-H); 2.13–2.24 (m, 1 H, 6eq-H); 2.28 (s, 3 H, 16-H); 2.38 (ddd, $J_{gem} = 13.6$ Hz, $J = 8.2$ Hz, $J = 3.7$ Hz, 1 H, 3eq-H); 3.04–3.13 (m, 1 H, 2-H); 3.23–3.31 (m, 1 H, 1-H); 3.48–3.57 (m, 1 H, 4-H); 3.72 (s, 3 H, 8-H or 8'-H); 3.74 (s, 3 H, 8-H or 8'-H); 3.80–3.85 (bs, 1 H, 5-H); 6.60–6.68 (m, 2 H, 13-H + 15-H); 7.02 (t, $J = 7.8$ Hz, 1 H, 14-H). ¹³C-NMR (100.61 MHz, CDCl₃): 12.7 (17-C); 20.8 (16-C); 28.6 (3-C); 31.1 (6-C); 39.7 (1-C); 40.3 (2-C); 52.1 (8-C + 8'-C); 53.8 (4-C); 68.3 (5-

C); 108.9 (15-C); 120.1 (13-C); 121.2 (11-C); 126.3 (14-C); 136.9 (12-C); 144.7 (10-C); 174.3 (7-C or 7'-C); 174.8 (7-C or 7'-C). MS (ESI⁺): 336 [M + H]⁺, 358 [M + Na]⁺.



5.7. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(3,5-dimethoxyphenylamino)-5-hydroxycyclohexane-1,2-dicarboxylate **6f** (Fig. 2. – S6)

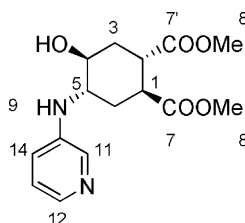
Starting from 120 mg (0.56 mmol) of epoxide **3**, we obtained a crude reaction product which was purified by Biotage SP1TM using n-hexane/AcOEt (from 100/0 to 20/80) to afford **6f** as a colorless oil (164 mg, 80%). *R*_f = 0.3 (Hexane/EtOAc 7/3). ¹H-NMR (400 MHz, CDCl₃): 1.64 (ddd, *J*_{gem} = 12.9 Hz, *J* = 8.1 Hz, *J* = 4.4 Hz, 1 H, 3ax-H); 1.81 (ddd, *J*_{gem} = 13.9 Hz, *J* = 8.1 Hz, *J* = 4.8 Hz, 1 H, 6ax-H); 2.16 (bs, 1 H, 4-OH); 2.24 (ddd, *J*_{gem} = 13.9 Hz, *J* = 6.8 Hz, *J* = 3.6 Hz, 1 H, 6eq-H); 2.41 (ddd, *J*_{gem} = 13.5 Hz, *J* = 6.9 Hz, *J* = 3.6 Hz, 1 H, 3eq-H); 3.01 (dd, *J* = 11.0 Hz, *J* = 6.1 Hz, 1 H, 2-H); 3.28 (dd, *J* = 11.6 Hz, *J* = 5.6 Hz, 1 H, 1-H); 3.43 (ddd, *J* = 7.7 Hz, *J* = 7.6 Hz, *J* = 4.0 Hz, 1 H, 4-H); 3.52 (bs, 1 H, 5-H); 3.72 (s, 3 H, 8-H or 8'-H); 3.75 (s, 3 H, 8-H or 8'-H); 3.76 (s, 6 H, 14-H); 5.89 (d, *J* = 2.1 Hz, 2 H, 11-H); 5.90–5.93 (m, 1 H, 13-H). ¹³C-NMR (100.61 MHz, CDCl₃): 28.3 (6-C); 31.1 (3-C); 39.7 (1-C); 40.2 (2-C); 52.2 (8-C + 8'-C); 53.7 (4-C); 55.2 (11-C); 68.1 (5-C); 90.3 (13-C); 92.3 (11-C); 149.0 (10-C); 161.8 (12-C); 174.3 (7-C or 7'-C); 174.9 (7-C or 7'-C). MS (ESI⁺): 368 [M + H]⁺, 390 [M + Na]⁺.



5.8. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-hydroxy-5-(4-nitrophenylamino)cyclohexane-1,2-dicarboxylate **6g** (Fig. 2. – S6)

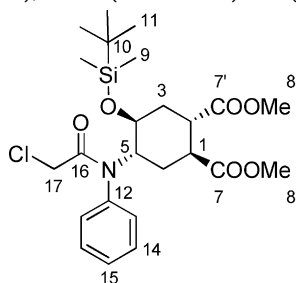
Starting from 98 mg (457 μmol) of epoxide **3**, we obtained a crude reaction product which was purified by flash chromatography using n-hexane/AcOEt (from 100/0 to 50/50) to afford **6g** as a colorless oil (69 mg, 43%). *R*_f = 0.3 (Hexane/EtOAc 6/4). ¹H-NMR (400 MHz, CDCl₃): 1.65 (ddd, *J*_{gem} = 12.6 Hz, *J* = 7.7 Hz, *J* = 4.4 Hz, 1 H, 3ax-H); 1.77 (ddd, *J*_{gem} = 12.9 Hz, *J* = 7.6 Hz, *J* = 4.7 Hz, 1 H, 6ax-H); 2.13 (ddd, *J*_{gem} = 12.9 Hz, *J* = 6.9 Hz, *J* = 3.0 Hz, 1 H, 6eq-H); 2.31 (ddd, *J*_{gem} = 12.6 Hz, *J* = 7.7 Hz, *J* = 4.1 Hz, 1 H, 3eq-H); 2.78 (bs, 1 H, OH); 3.05 (dd, *J* = 11.0 Hz, *J* = 6.8 Hz, 1 H, 2-H); 3.22 (dd, *J* = 11.4 Hz, *J* = 6.5 Hz, 1 H, 2-H); 3.54 (dt, *J* = 11.1 Hz, *J* = 7.2 Hz, 1 H, 4-H); 3.63 (s, 3 H, 8-H or 8'-H); 3.68 (s, 3 H, 8-H or 8'-H); 3.80 (ddd, *J* = 7.3 Hz, *J* = 7.1 Hz, *J* = 3.3 Hz, 1

H, 5-H); 4.93 (d, *J* = 7.5 Hz, 1 H, 9-H); 6.56 (d, *J* = 9.2 Hz, 2 H, 11-H); 7.94 (d, *J* = 9.2 Hz, 2 H, 12-H). ¹³C-NMR (100.61 MHz, CDCl₃): 28.1 (3-C); 31.4 (6-C); 39.9 (1-C); 40.1 (2-C); 52.3 (8-C or 8'-C); 52.4 (8-C or 8'-C); 53.7 (4-C); 68.0 (5-C); 111.6 (11-C); 126.5 (12-C); 138.0 (13-C); 153.0 (9-C); 174.0 (7-C or 7'-C); 174.5 (7-C or 7'-C). MS (ESI⁺): 353 [M + H]⁺, 375 [M + Na]⁺.



5.9. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-hydroxy-5-(pyridin-3-ylamino)cyclohexane-1,2-dicarboxylate **6h** (Fig. 2. – S8)

Starting from 120 mg (0.56 mmol) of epoxide **3**, we obtained a crude reaction product which was purified with Biotage SP1TM using CH₂Cl₂/MeOH (from 90/10 to 80/20) to afford **6h** as a white solid (69 mg, 40%). *R*_f = 0.1 (DCM/MeOH 9/1). ¹H-NMR (400 MHz, CD₃OD): 1.77 (ddd, *J*_{gem} = 13.9 Hz, *J* = 11.3 Hz, *J* = 5.2 Hz, 1 H, 3ax-H); 2.34 (dt, *J*_{gem} = 13.3 Hz, *J* = 5.1 Hz, 1 H, 6ax-H); 2.52–2.64 (m, 2 H, 3eq-H + 6eq-H); 3.42–3.50 (m, 1 H, 2-H); 3.50–3.56 (m, 1 H, 1-H); 3.84 (s, 3 H, 8-H or 8'-H); 3.85 (s, 3 H, 8-H or 8'-H); 4.05 (dt, *J* = 10.9 Hz, *J* = 4.6 Hz, 1 H, 4-H); 4.42 (ddd, *J* = 12.9 Hz, *J* = 9.6 Hz, *J* = 3.5 Hz, 1 H, 5-H); 7.66–7.73 (m, 2 H, 13-H + 14-H); 8.06–8.16 (m, 1 H, 12-H); 8.16–8.26 (m, 1 H, 11-H). ¹³C-NMR (100.61 MHz, CD₃OD): 30.6 (6-C); 33.6 (3-C); 41.9 (1-C); 42.1 (2-C); 52.2 (8-C + 8'-C); 69.5 (4-C); 75.2 (5-C); 128.4 (11-C); 129.0 (13-C or 14-C); 129.4 (13-C or 14-C); 131.5 (12-C); 150.5 (10-C); 174.1 (7-C or 7'-C); 174.4 (7-C or 7'-C). MS (ESI⁺): 309 [M + H]⁺.



5.10. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(tert-butyldimethylsilyloxy)-5-(2-chloro-N-phenylacetamido)cyclohexane-1,2-dicarboxylate **6i** (Fig. 2. – S9)

To a solution of compound **6a** (273 mg, 0.89 mmol) and dry 2,6-lutidine (190 mg, 1.78 mmol) in dry CH₂Cl₂ (4.4 mL), at 0 °C and under N₂, TBDMSOTf (398 mg, 1.51 mmol) was added. The reaction was stirred at room temperature for 2.5 h and monitored by TLC (9/1 CH₂Cl₂/EtOAc). After reaction completion, the solvent was evaporated under reduced pressure and the crude was taken up with Et₂O, washed with H₂O and dried with

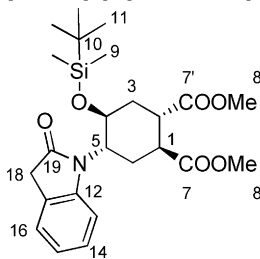
Na₂SO₄. The solvent was evaporated under reduced pressure and the resulting crude silylether intermediate (345 mg, 92%) was used for the following reaction without any further purification. R_f = 0.1 (DCM/EtOAc 9/1). MS (ESI⁺): 422.2 [M + H⁺].

To a solution of the silylether intermediate (390 mg, 0.93 mmol) and dry 2,6-lutidine (222 μg, 1.87 mmol) in dry toluene (2 mL), chloroacetylchloride (181 mg, 1.6 mmol) was added under nitrogen and at room temperature. The reaction was stirred while monitoring by TLC (8/2 petroleum ether/EtOAc), then the solution was diluted with Et₂O and the organic phase was washed with water. The solvent was dried with Na₂SO₄ and evaporated under reduced pressure, to yield a crude that was purified by flash chromatography (95/5 CH₂Cl₂/AcOEt), affording compound **9** (452 mg, 98%). R_f = 0.4 (Hexane/EtOAc 8/2). ¹H-NMR (400 MHz, CDCl₃): 0.16 (s, 3 H, 9-H); 0.18 (s, 3 H, 9-H); 0.95 (s, 9H, 11-H); 1.61 (m, 1 H, 6ax-H); 2.25–2.38 (m, 3 H, 3eq-H + 3ax-H + 6eq-H); 3.05–3.11 (m, 1 H, 2-H); 3.20–3.26 (m, 1 H, 1-H); 3.65 (s, 3 H, 8-H or 8'-H); 3.72 (s, 3 H, 8-H or 8'-H); 3.75 (m, 3 H, 4-H + 2 × 17-H); 4.41–4.51 (m, 1 H, 5-H); 7.27–7.38 (m, 2 H, Ar-H); 7.40–7.50 (m, 3 H, Ar-H). ¹³C-NMR (100.61 MHz, CDCl₃): -4.3 (9-C); -4.2 (9-C); 18.0 (10-C); 25.9 (11-C); 26.4 (3-C); 33.5 (6-C); 40.6 (1-C + 2-C); 43.3 (17-C); 52.2 (8-C or 8'-C); 64.1 (4-C); 128.8 (Ar-C); 129.9 (Ar-C); 141.4 (12-C); 165.8 (16-C); 173.4 (7-C or 7'-C); 173.7 (7-C or 7'-C). MS (ESI⁺): 498.2 [M + H⁺]. [α]_D = -174.0 [MeOH, c = 1.0].

5.11. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(*tert*-butyldimethylsilyloxy)-5-(2-oxoindolin-1-yl)cyclohexane-1,2-dicarboxylate **11** (Fig. 2. – S10)

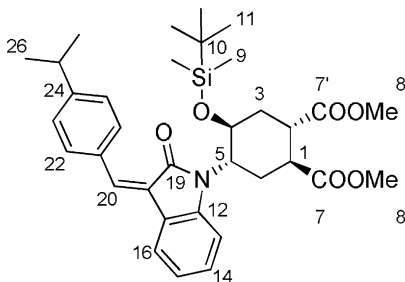
In a Schlenk reactor, under argon atmosphere and at room temperature, **9** (109 mg, 0.22 mmol), Pd(OAc)₂ (5 mg, 0.022 mmol) and di-*tert*-butyl-diphenyl-phosphine (14 mg, 0.48 mmol) were dissolved in dry toluene (240 μL), and dry DIPEA (63 μL, 0.36 mmol) was added. The reaction mixture was stirred at 100 °C for ca. 6 h, monitoring by TLC (95/5 CH₂Cl₂/AcOH). After reaction completion, the reaction mixture was taken up with EtOAc (2 mL), filtered on a pad of celite, and the organic phase was evaporated under reduced pressure. The crude was purified by flash chromatography (95/5 CH₂Cl₂/AcOH) affording compound **11** (48 mg, 47%). Starting material **9** (33 mg, 68% conversion) was also recovered, and could be recycled. R_f = 0.4 (Hexane/EtOAc 8/2). ¹H-NMR (400 MHz, CDCl₃): -0.30 (s, 3 H, 9-H); -0.01 (s, 3 H, 9-H); 0.66 (s, 9H, 11-H); 1.65 (dt, J_{gem} = J_{1-6ax} = 15.6 Hz, J_{5-6ax} = 4.8 Hz, 1 H, 6ax-H); 2.21 (td, J_{gem} = 12.0 Hz, J_{2-3eq} = J_{3eq-4} = 2.0 Hz, 1 H, 3eq-H); 2.44 (td, J_{gem} = 15.6 Hz, J_{1-6eq} = J_{5-6eq} = 2.0 Hz, 1 H, 6eq-H); 2.72 (dt, J_{gem} = J_{3ax-2} = 13.5 Hz, J_{3ax-4} = 4.6 Hz, 1 H, 3ax-H); 3.40 (bs, 2 H, 1-H + 2-H); 3.46 (s, 2 H, 18-H); 3.82 (s, 3 H, 8-H or 8'-H); 3.83 (s, 3 H, 8-H or 8'-H); 3.98–4.07 (m, 1 H, 5-H); 4.54 (dt, J = 11.0 Hz, J_{3ax-4} = 4.6 Hz, 1 H, 4-H); 6.93 (d, J₁₅₋₁₆ = 7.8 Hz, 1 H, 16-H); 7.00 (t, J₁₃₋₁₄ = J₁₄₋₁₅ = 7.8 Hz, 1 H, 14-H); 7.22 (t, J₁₄₋₁₅ = J₁₅₋₁₆ = 7.8 Hz, 1 H, 15-H); 7.18–7.28 (m, 1 H, 13-H). ¹³C-NMR (100.61 MHz, CDCl₃): -5.5 (9-C); -4.7 (9-C); 17.5 (10-C); 25.3 (3-C); 25.4 (11-C); 33.6 (6-C); 36.3 (18-C); 41.0 (2-C); 41.2 (1-C); 52.3 (8-C or 8'-C); 52.4 (8-C or 8'-C); 55.5 (5-C); 66.3 (4-C); 109.2 (16-C); 121.7 (14-C); 124.2 (15-C); 124.4 (17-C); 127.6 (13-C); 145.2 (12-C); 173.4 (19-C);

173.5 (7-C or 7'-C); 175.2 (7-C or 7'-C). MS (ESI⁺): 462.0 [M + H⁺]. [α]_D = +175.0 [c = 1.5, MeOH].



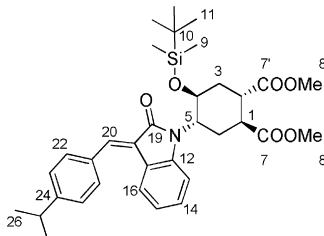
5.12. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(*tert*-butyldimethylsilyloxy)-5-((*E* and *Z*)-3-(4-isopropylbenzylidene)-2-oxoindolin-1-yl)cyclohexane-1,2-dicarboxylate **13**

To a solution of **11** (67 mg, 0.164 mmol) in dry methanol (290 μL), under nitrogen atmosphere and at room temperature, 4-isopropyl-benzaldehyde (27 μL, 0.175 mmol) and piperidine (6 μL, 0.06 mmol) were added. The reaction mixture was stirred at 65 °C monitoring the reaction progression by TLC (85/15 petroleum ether/EtOAc). After reaction completion, the solvent was evaporated under reduced pressure and the silylether **13** was isolated by flash chromatography (85/15 petroleum ether/EtOAc) as a *E/Z* isomeric mixture (251 mg, 94%).



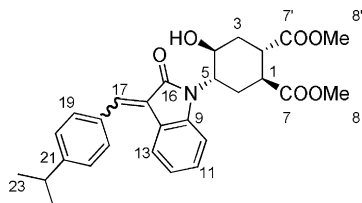
5.12.1. *Z* isomer (Fig. 2. – S11)

R_f = 0.34 (Hexane/EtOAc 9/1). ¹H-NMR (400 MHz, CDCl₃): -0.33 (s, 3 H, 9-H); -0.03 (s, 3 H, 9-H); 0.66 (s, 9 H, 11-H); 1.30 (d, J₂₅₋₂₆ = 7.2 Hz, 6 H, 26-H); 1.60–1.72 (m, 1 H, 3ax-H); 2.20–2.30 (m, 1 H, 6eq-H); 2.42–2.51 (m, 1 H, 3eq-H); 2.79–2.90 (m, 1 H, 6ax-H); 2.92–3.01 (sp, J₂₅₋₂₆ = 7.2 Hz, 1 H, 25-H); 3.37–3.45 (m, 2 H, 1-H + 2-H); 3.82 (s, 3 H, 8-H or 8'-H); 3.84 (s, 3 H, 8-H or 8'-H); 4.08–4.02 (m, 1 H, 5-H); 4.53–4.60 (m, 1 H, 4-H); 6.92 (d, J₁₅₋₁₆ = 8.0 Hz, 1 H, 16-H); 7.01 (dt, J = 7.6 Hz, J = 0.9 Hz, 1 H, 14-H); 7.24 (dt, J = 7.8 Hz, J = 1.2 Hz, 1 H, 15-H); 7.32 (d, J₂₂₋₂₃ = 8.0 Hz, 2 H, 23-H); 7.50 (s, 1 H, 20-H); 8.22 (d, J₂₂₋₂₃ = 8.0 Hz, 2 H, 22-H). MS (ESI⁺): 592 [M + H⁺].

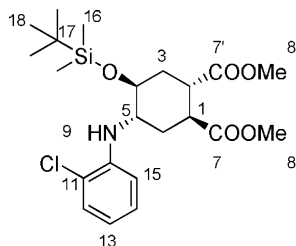


5.12.2. *E* isomer (Fig. 2. – S12)

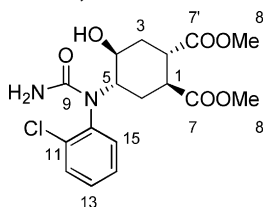
$R_f = 0.34$ (Hexane/EtOAc 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): -0.28 (s, 3 H, 3-H); 0.06 (s, 3 H, 9-H); 0.66 (s, 9 H, 11-H); 1.32 (d, $J_{25-26} = 6.8$ Hz, 6 H, 26-H); 1.58 – 1.71 (m, 1 H, 3ax-H); 2.21 – 2.30 (m, 1 H, 6eq-H); 2.44 – 2.51 (m, 1 H, 3eq-H); 2.79 – 2.89 (m, 1 H, 6'-H); 2.98 (sp, $J_{25-26} = 6.8$ Hz, 1 H, 25-H); 3.40 (bs, 1 H, 2-H); 3.44 (bs, 1 H, 1-H); 3.83 (s, 3 H, 8-H or 8'-H); 3.85 (s, 3 H, 8-H or 8'-H); 3.97 – 4.06 (m, 1 H, 5-H); 4.51 – 4.62 (m, 1 H, 4-H); 6.84 – 6.89 (m, 1 H, 14-H); 6.92 – 6.97 (m, 1 H, 16-H); 7.21 – 7.28 (m, 1 H, 15-H); 7.34 (d, $J_{22-23} = 8.0$ Hz, 2 H, 23-H); 7.60 (d, $J_{22-23} = 8.0$ Hz, 2 H, 22-H); 7.72 (d, $J_{13-14} = 7.2$ Hz, 1 H, 13-H); 7.75 (s, 1 H, 20-H). MS (ESI⁺): 592 [M + H⁺].

5.13. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-hydroxy-5-(3-(4-isopropylbenzylidene)-2-oxoindolin-1-yl)cyclohexane-1,2-dicarboxylate **14** (Fig. 2. – S13)

A 1 M solution of TBAF in THF (100 μL , 0.84 mmol) was added to intermediate silylether **13** (14.5 mg, 0.21 mmol). The reaction mixture was stirred at room temperature for 16 h, monitoring the reaction progression by TLC (85/15 $\text{CH}_2\text{Cl}_2/\text{EtOAc}$). After reaction completion the mixture was taken up with Et_2O and washed with saturated NH_4Cl . The organic phase, dried with Na_2SO_4 , was evaporated under reduced pressure and the crude product was purified by flash chromatography (85/15 $\text{CH}_2\text{Cl}_2/\text{EtOAc}$) affording compound **14** (10.5 mg, 90%) as a $\approx 40/60$ (Z/E) isomeric mixture of Z and E isomers which reequilibrated after any purification attempt. $R_f = 0.2 + 0.4$ (DCM/EtOAc 85/15). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 1.33 (d, $J_{22-23} = 6.8$ Hz, 6 H, 23-H); 1.62 – 1.78 (m, 1 H, 3eq-H); 2.15 – 2.40 (m, 2 H, 6ax-H + 6eq-H); 2.58 – 2.66 (m, 1 H, 3ax-H); 3.32 (bs, 1 H, 1-H); 3.36 (bs, 1 H, 2-H); 3.82 (s, 3 H, 8-H or 8'-H); 3.84 (s, 3 H, 8-H or 8'-H); 3.95 – 4.01 (m, 1 H, 5-H); 4.70 – 4.86 (m, 1 H, 4-H); 6.43 – 6.49 (m, Ar-H, Z isomer); 6.67 – 6.74 (m, Ar-H, Z isomer); 6.77 – 6.90 (m, Ar-H, E and Z isomers); 6.95 – 7.15 (m, Ar-H, E and Z isomers); 7.19 – 7.28 (m, Ar-H, E and Z isomers); 7.31 (m, Ar-H, Z isomer); 7.46 (m, Ar-H, Z isomer); 7.52 (m, Ar-H, E isomer); 7.67 (m, Ar-H, E isomer); 7.70 (m, Ar-H, E isomer); 8.10 (m, Ar-H, Z isomer). MS (ESI⁺): 478.0 [M + H⁺].

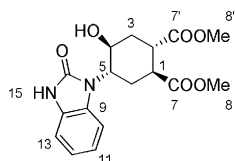
5.14. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(tert-butylidimethylsilyloxy)-5-(2-chlorophenylamino)cyclohexane-1,2-dicarboxylate **17** (Fig. 2. – S14)

To a solution of compound **6b** (205 mg, 0.6 mmol) and 2,6-lutidine (140 μL , 1.2 mmol) in dry CH_2Cl_2 (2.4 mL), at 0°C and under N_2 , TBDMSOTf (330 μL , 1.44 mmol) was added. The reaction mixture was stirred at room temperature for 2.5 h, monitoring the reaction progression by TLC (9/1 $\text{CH}_2\text{Cl}_2/\text{EtOAc}$). After reaction completion, the solvent was evaporated under reduced pressure, the crude was taken up with Et_2O , washed with H_2O and dried with Na_2SO_4 . The solvent was evaporated under reduced pressure. The crude was purified by flash chromatography on silica gel (9/1 petroleum ether/EtOAc), affording **17** (232 mg, 85%) that was used as such. A sample of the silylether was further purified by flash chromatography (94/6 petroleum ether/EtOAc) for analytical characterization. $R_f = 0.2$ (Hexane/EtOAc 6/4). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.00 (s, 6 H, 16-H); 0.80 (s, 9 H, 18-H); 1.70 – 1.85 (m, 3 H, 3ax-H + 6eq-H + 6ax-H); 2.10 (m, 1 H, 3eq-H); 2.82 – 2.89 (m, 1 H, 2-H); 3.02 – 3.09 (m, 1 H, 1-H); 3.36 – 3.42 (m, 1 H, 4-H); 3.60 (s, 6 H, 8-H and 8'-H); 3.82 – 3.89 (m, 1 H, 5-H); 6.60 (t, $J_{12-13} = J_{13-14} = 8.0$ Hz, 1 H, 13-H); 6.68 (d, $J_{14-15} = 8.0$ Hz, 1 H, 15-H); 7.05 (t, $J_{13-14} = J_{14-15} = 8.0$ Hz, 1 H, 14-H); 7.20 (m, 1 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.7 (16-C); -4.6 (16-C); 17.9 (17-C); 25.8 (18-C); 27.7 (6-C); 31.5 (3-C); 39.0 (2-C); 39.6 (1-C); 52.1 (8-C + 8'-C); 53.0 (5-C); 67.8 (4-C); 112.5 (15-C); 119.0 (13-C); 127.9 (14-C); 129.5 (12-C); 142.1 (10-C); 173.9 (7-C or 7'-C); 174.1 (7-C or 7'-C).

5.15. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-(1-(2-chlorophenyl)ureido)-5-hydroxy-cyclohexane-1,2-dicarboxylate **18** (Fig. 2. – S15)

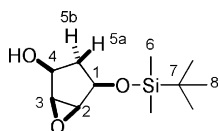
A solution of chlorosulfonyl isocyanate (70 μL , 0.81 mmol) in dry THF (560 μL), under N_2 , was cooled to -10°C and added dropwise over 30 minutes (using a syringe pump) to a solution of the **17** (232 mg, 0.51 mmol) in dry THF (1.2 mL). After 20 minutes a second portion of chlorosulfonyl isocyanate (30 μL , 0.34 mmol) was added dropwise. The reaction progression was monitored by TLC (95/5 $\text{CH}_2\text{Cl}_2/\text{MeOH}$). After reaction completion, water (120 μL) was added, and the mixture stirred for 30 minutes at room temperature. The solution was treated with 3 M NaOH until pH 8–9, and the organic phase washed twice with brine and dried with Na_2SO_4 . The solvent was evaporated under reduced pressure and the crude was purified by flash chromatography on silica gel (93/7 $\text{CH}_2\text{Cl}_2/\text{MeOH}$), affording **18** (158 mg, 81%). $R_f = 0.3$ (DCM/MeOH 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 1.23 (td, $J_{\text{gem}} = J_{5-6\text{ax}} = 13.0$ Hz, $J_{1-6\text{ax}} = 4.0$ Hz, 1 H, 6ax-H); 1.77 (td, $J_{\text{gem}} = J_{3\text{ax}-4} = 13.6$ Hz, $J_{2-3\text{ax}} = 5.6$ Hz, 1 H, 3ax-H); 2.38 –

2.52 (m, 2 H, 3eq-H + 6eq-H); 3.12 (bs, 1 H, 1-H); 3.27 (bs, 1 H, 2-H); 3.70–3.55 (m, 4H, 4-H + 8-H or 8'-H); 3.77 (s, 3 H, 8-H or 8'-H); 4.43–4.52 (m, 1 H, 5-H); 7.38 (m, 2 H, 13-H + 15-H); 7.46 (m, 1 H, 14-H); 7.55 (m, 1 H, 12-H). ¹³C-NMR (100.61 MHz, CDCl₃): 26.5 (6-C); 33.20 (3-C); 40.5 (2-C); 40.7 (1-C); 52.1 (8-C or 8'-C); 52.5 (8-C or 8'-C); 58.4 (5-C); 70.0 (4-C); 128.6 (15-C); 130.4 (13-C); 131.1 (12-C); 132.3 (14-C); 135.3 (11-C); 135.7 (10-C); 159.3 (9-C); 173.3 (7-C or 7'-C); 173.6 (7-C or 7'-C). MS (ESI⁺): 791.2 [2M + Na⁺]. [α]_D = -26.7 [c = 1.0, CHCl₃]. HRMS: C₁₇H₂₁N₂O₆ClNa: calcd. 407.09804; found 407.09781.



5.16. (1*S*,2*S*,4*S*,5*S*)-dimethyl-4-hydroxy-5-(2-oxo-2,3-dihydro-1*H*-benzo[d]imidazol-1-yl)cyclohexane-1,2-dicarboxylate **21** (Fig. 2. – S16)

A solution of **18** (157.9 mg, 0.41 mmol) and *i*Pr₂EtN (215 μL, 1.23 mmol) in dry *i*PrOH (1.8 mL) (dried on molecular sieves and purged with N₂) was stirred under N₂ atmosphere. The reaction mixture was purged with N₂ for 15 minutes and then a first aliquot of X-Phos [9] (2-dicyclohexylphosphino-2',4',6'-triisopropylbiphenyl) (24.4 mg, 0.06 mmol) and Pd(OAc)₂ (4.6 mg, 0.02 mmol) were added. The reaction mixture was purged with N₂ for further 15 minutes and the solution was heated to 83 °C. After 4 h a second aliquot of X-Phos (24.4 mg, 0.06 mmol) and Pd(OAc)₂ (4.6 mg, 0.02 mmol) in *i*PrOH (300 μL) was added. The reaction was stirred at 83 °C for 24 h, monitoring its progression by TLC (EtOAc). The solvent was evaporated under reduced pressure, the residue taken up with CH₂Cl₂, washed once with H₂O and dried with Na₂SO₄. The crude was purified by flash chromatography on silica gel (EtOAc), affording **21** (74 mg, 51%) and recovered starting material **18** (34 mg, 65% conversion). R_f = 0.2 (DCM/MeOH 9/1). ¹H-NMR (400 MHz, CDCl₃): 1.78 (td, J_{gem} = J_{3ax-4} = 13.2 Hz, J_{2-6ax} = 5.0 Hz, 1 H, 3ax-H); 2.41 (m, 2 H, 6eq-H + 6ax-H); 2.62 (dt, J_{gem} = 13.7 Hz, J_{3eq-4} = J_{2-3eq} = 2.0 Hz, 1 H, 3eq-H); 3.40 (bs, 1 H, 2-H); 3.47 (bs, 1 H, 1-H); 3.83 (s, 3 H, 8-H or 8'-H); 3.84 (s, 3 H, 8-H or 8'-H); 4.18 (m, 1 H, 4-H); 4.65 (dt, J₄₋₅ = J_{3eq-5} = 11.2 Hz, J_{3ax-5} = 4.3 Hz, 1 H, 5-H); 7.01–7.12 (m, 3 H, 10-H + 11-H + 13-H); 7.23 (d, J = 7.4 Hz, 1 H, 10-H). 9.10 (bs, 1 H, 15-H). ¹³C-NMR (Hecor, CDCl₃, 400 MHz): 26.8 (6-C); 32.4 (3-C); 40.7 (2-C); 41.1 (1-C); 52.4 (8-C and 8'-C); 56.9 (5-C); 65.9 (4-C); 108.9 (15-C); 109.5 (13-C); 121.5 (12-C). MS (ESI⁺): 719.8 [2M + Na⁺]. [α]_D = +21.0 [c = 1.5, CHCl₃]. HRMS: C₁₇H₂₀N₂O₆Na: calcd. 371.12136; found 371.12189.



5.17. (±)-(1*S*,2*S*,4*R*,5*S*)-4-(*tert*-butyldimethylsilyloxy)-6-oxabicyclo[3.1.0]hexan-2-ol **4** (Fig. 2. – S17)

To a stirred solution of commercially available (±)-(1*S*,4*R*)-4-(*tert*-butyldimethylsilyloxy)cyclopent-2-enol (3.5 g, 16.33 mmol) in CH₂Cl₂ (40 mL) 3-chloroperbenzoic acid (5.63 g, 32.7 mmol) was added and the reaction was stirred for 3 h at room temperature. The reaction mixture was diluted with CH₂Cl₂ (40 mL) and quenched by saturated Na₂CO₃. The organic layer was washed (2 × 60 mL) with saturated Na₂CO₃, then with brine (2 × 60 mL), was dried on Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (n-hexane/EtOAc from 20/80 to 50/50) to afford epoxide **4** (3.21 g, 85%) as a white solid. R_f = 0.5 (Hexane/EtOAc 6/4). ¹H-NMR (400 MHz, CDCl₃): 0.08 (s, 3 H, 3 × 6-H); 0.10 (s, 3 H, 3 × 6-H); 0.91 (s, 9 H, 9 × 8-H); 1.30 (dt, J_{gem} = 12.6 Hz, J_{1-5a} = J_{4-5a} = 8.4 Hz, 1 H, 5a-H); 2.19 (dt, J_{gem} = 12.6 Hz, J_{1-5b} = J_{4-5b} = 7.6 Hz, 1 H, 5b-H); 3.44 (dd, J₁₋₂ = 1.4 Hz, J₂₋₃ = 2.9 Hz, 1 H, 2-H); 3.48 (dd, J₃₋₄ = 1.4 Hz, J₂₋₃ = 2.9 Hz, 1 H, 3-H); 4.02–4.08 (m, 1 H, 4-H); 4.10–4.16 (m, 1 H, 1-H). ¹³C-NMR (100.61 MHz, CDCl₃): -4.8 (6-C); -4.7 (6-C); 18.0 (7-C); 25.7 (8-C); 34.4 (5-C); 57.3 (3-C); 58.3 (2-C); 69.9 (4-C); 70.6 (1-C). MS (ESI⁺): 253 [M + Na]⁺; 231 [M + H]⁺.

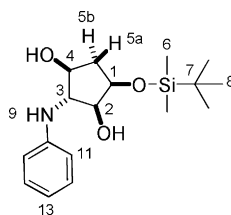
5.18. Epoxide opening: general procedure for compound **4**

To a 0.061 M solution of **4** [14] (1 mol equiv.) in CHCl₃ anilines **5a–h** (1.5 mol equiv.) and InBr₃ (0.5 mol equiv.) were added. The reaction mixture was stirred between 10 h to 48 h at reflux, monitoring the reaction progression by TLC (6/4 petroleum ether/EtOAc). After reaction completion the solution was evaporated and all the compounds were isolated by flash chromatography or with Biotage SP1TM.

5.19. (±)-(1*S*,2*R*,3*S*,4*R*)-4-(*tert*-butyldimethylsilyloxy)-2-(phenylamino)cyclopentane-1,3-diol **7a** and (1*S*,2*R*,3*R*,4*R*)-4-(*tert*-butyldimethylsilyloxy)-3-(phenylamino)cyclopentane-1,2-diol **8a**

Starting from 500 mg (2.17 mmol) of epoxide **4**, we obtained a crude reaction product which was purified by Biotage SP1TM (n-hexane/EtOAc from 90/10 to 0/100; SNAP 100 g column) to give **7a** (377 mg, 54%) and **8a** (155 mg, 22%) as brown oils.

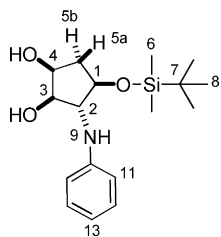
5.19.1. **7a** (Fig. 2. – S18)



R_f = 0.5 (Hexane/EtOAc 5/5). ¹H-NMR (400 MHz, CDCl₃): 0.14 (s, 3 H, 3 × 6-H); 0.15 (s, 3 H, 3 × 6-H); 0.94 (s, 9 H, 9 × 8-H); 1.87 (ddt, J_{gem} = 14.3 Hz, J = 4.3 Hz, J = 1.1 Hz, 1 H,

5a-H); 2.26 (ddd, $J_{\text{gem}} = 14.3 \text{ Hz}$, $J = 6.5 \text{ Hz}$, $J = 5.7 \text{ Hz}$, 1 H, 5b-H); 2.74–2.75 ($2 \times \text{bs}$, 2 H, 2-OH+4-OH); 3.61 (t, $J = 3.5 \text{ Hz}$, 1 H, 3-H); 3.71–3.80 (dd + bs, $J = 10.3 \text{ Hz}$, $J = 4.8 \text{ Hz}$, 2 H, 9-NH + 2-H); 3.90 (bs, 1 H, 4-H); 4.24 (dd, $J = 9.8 \text{ Hz}$, $J = 4.8 \text{ Hz}$, 1 H, 1-H); 6.75 (d, $J_{12-13} = 7.3 \text{ Hz}$, 2 H, 13-H); 6.79 (d, $J_{11-12} = 8.7 \text{ Hz}$, 2 H, 11-H); 7.20 (dd, $J_{11-12} = 8.7 \text{ Hz}$, $J_{12-13} = 7.3 \text{ Hz}$, 1 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.4 (7-C); 26.1 (8-C); 40.2 (5-C); 69.2 (3-C); 73.5 (1-C); 76.8 (4-C); 79.1 (2-C); 114.1 (11-C); 118.5 (13-C); 129.6 (12-C); 147.6 (10-C). MS (ESI⁺): 324 [M + H]⁺; 346 [M + Na]⁺.

5.19.2. 8a (Fig. 2. – S19)

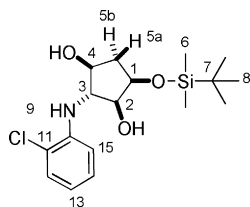


$R_f = 0.2$ (Hexane/EtOAc 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.08 (s, 3 H, 3 \times 6-H); 0.09 (s, 3 H, 3 \times 6-H); 0.91 (s, 9 H, 9 \times 8-H); 1.91 (ddd, $J_{\text{gem}} = 14.6 \text{ Hz}$, $J_{4-5a} = 4.3 \text{ Hz}$, $J_{1-5a} = 2.8 \text{ Hz}$, 1 H, 5a-H); 2.17 (ddd, $J_{\text{gem}} = 14.3 \text{ Hz}$, $J_{1-5b} = J_{4-5b} = 5.6 \text{ Hz}$, 1 H, 5b-H); 3.10 (bs, 1 H, OH); 3.20 (bs, 1 H, OH); 3.65–3.70 (m, 1 H, 2-H); 3.77 (t, $J_{2-3} = J_{3-4} = 4.9 \text{ Hz}$, 1 H, 3-H); 4.00 (ddd, $J_{1-5b} = 4.6 \text{ Hz}$, $J_{1-5a} = J_{1-2} = 2.8 \text{ Hz}$, 1 H, 1-H); 4.13–4.20 (m, 1 H, 4-H); 6.72 (d, $J_{11-12} = 8.6 \text{ Hz}$, 1 H, 11-H); 6.76 (d, $J_{12-13} = 7.3 \text{ Hz}$, 1 H, 13-H); 7.18 (dd, $J_{11-12} = 8.6 \text{ Hz}$, $J_{12-13} = 7.3 \text{ Hz}$, 1 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.2 (7-C); 26.1 (8-C); 39.8 (5-C); 68.7 (2-C); 72.8 (4-C); 78.0 (1-C); 79.6 (3-C); 114.1 (11-C); 118.5 (13-C); 129.6 (12-C); 148.0 (10-C). MS (ESI⁺): 324 [M + H]⁺; 346 [M + Na]⁺.

5.20. (\pm)-(1S,2S,4R,5S)-4-(tert-butyl dimethylsilyloxy)-2-(2-chlorophenylamino)cyclopentane-1,3-diol 7b and (\pm)-(1S,2R,3R,4R)-4-(tert-butyl dimethylsilyloxy)-3-(2-chlorophenylamino)cyclopentane-1,2-diol 8b

Starting from 500 mg (2.17 mmol) of epoxide **4**, we obtained a crude reaction product which was purified by Biotage SP1TM ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ from 98/2 to 80/20; SNAP 50 g column) to give **7b** (388 mg, 50%) and **8b** (195 mg, 25%) as colorless oils.

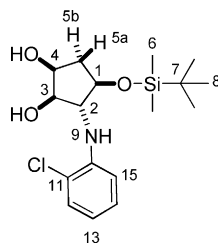
5.20.1. 7b (Fig. 2. – S20)



$R_f = 0.9$ (DCM/MeOH 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.15 (s, 3 H, 3 \times 6-H); 0.16 (s, 3 H, 3 \times 6-H); 0.94 (s, 9 H,

9 \times 8-H); 1.90 (ddd, $J_{\text{gem}} = 14.3 \text{ Hz}$, $J = 4.2 \text{ Hz}$, $J = 1.3 \text{ Hz}$, 1 H, 5a-H); 2.28 (ddd, $J_{\text{gem}} = 14.3 \text{ Hz}$, $J_{1-5b} = 6.5 \text{ Hz}$, $J_{4-5b} = 5.5 \text{ Hz}$, 1 H, 5b-H); 2.66 (d, $J = 9.2 \text{ Hz}$, 1 H, 4-OH); 2.74 (d, $J = 6.8 \text{ Hz}$, 1 H, 2-OH); 3.59–3.68 (m, 1 H, 3-H); 3.83 (dt, 1 H, 2-H, $J_{1-2} = J_{2-3} = 4.7 \text{ Hz}$, $J_{2-\text{OH}} = 6.8 \text{ Hz}$); 3.95 (ddd, $J = 9.8 \text{ Hz}$, $J = 7.1 \text{ Hz}$, $J = 3.7 \text{ Hz}$, 1 H, 4-H); 4.27 (dd, $J = 9.6 \text{ Hz}$, $J = 4.7 \text{ Hz}$, 1 H, 1-H); 4.32 (d, $J = 4.3 \text{ Hz}$, 1 H, 9-NH); 6.68 (ddd, $J = 7.8 \text{ Hz}$, $J = 7.4 \text{ Hz}$, $J = 1.5 \text{ Hz}$, 1 H, 13-H); 7.01 (dd, $J = 8.2 \text{ Hz}$, $J = 1.4 \text{ Hz}$, 1 H, 15-H); 7.15–7.21 (m, 1 H, 14-H); 7.26 (dd, $J = 7.8 \text{ Hz}$, $J = 1.4 \text{ Hz}$, 1 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.4 (7-C); 26.1 (8-C); 40.1 (5-C); 69.4 (3-C); 73.5 (1-C); 76.6 (4-C); 79.1 (2-C); 113.5 (15-C); 118.7 (13-C); 120.0 (11-C); 128.4 (14-C); 129.5 (12-C); 147.0 (10-C). MS (ESI⁺): 358 [M + H]⁺.

5.20.2. 8b (Fig. 2. – S21)

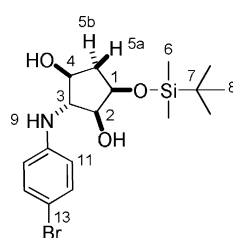


$R_f = 0.5$ (DCM/MeOH 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.14 (s, 3 H, 3 \times 6-H); 0.15 (s, 3 H, 3 \times 6-H); 0.90 (s, 9 H, 9 \times 8-H); 1.90–1.98 (m, 1 H, 5a-H); 2.22 (dt, $J_{\text{gem}} = 14.6 \text{ Hz}$, $J = 5.7 \text{ Hz}$, 1 H, 5b-H); 2.74 (d, $J = 6.8 \text{ Hz}$, 1 H, 2-OH); 3.06 (d, $J = 8.0 \text{ Hz}$, 1 H, 4-OH); 3.68–3.76 (m, 1 H, 3-H); 3.79–3.88 (m, 1 H, 2-H); 4.00–4.05 (m, 1 H, 4-H); 4.11–4.34 (m, 1 H, 1-H); 6.62–6.72 (m, 1 H, Ar-H); 6.91–6.95 (m, 1 H, Ar-H); 7.09–7.17 (m, 1 H); 7.24 (dd, $J = 7.9 \text{ Hz}$, $J = 1.5 \text{ Hz}$, 1 H, Ar-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.8 (6-C); -4.5 (6-C); 18.1 (7-C); 25.9 (8-C); 39.7 (5-C); 68.3 (3-C); 72.4 (1-C); 77.8 (4-C); 79.2 (2-C); 113.1 (15-C); 118.2 (13-C); 119.6 (11-C); 128.0 (14-C); 129.3 (12-C); 143.5 (10-C). MS (ESI⁺): 358 [M + H]⁺.

5.21. (\pm)-(1S,2R,4S,5R)-2-(4-bromophenylamino)-4-(tert-butyl dimethylsilyloxy)cyclopentane-1,3-diol 7c and (\pm)-(1S,2R,4S,5R)-3-(4-bromophenylamino)-4-(tert-butyl dimethylsilyloxy)cyclopentane-1,2-diol 8c

Starting from 70 mg (0.304 mmol) of epoxide **4**, we obtained a crude reaction product which was purified by Biotage SP1TM ($\text{CH}_2\text{Cl}_2/\text{EtOAc}$ from 100/0 to 20/80; SNAP 50 g column) to give a mixture of **7c** (49 mg, 40%) and **8c** (36 mg, 30%) as brown oils.

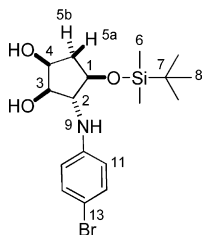
5.21.1. 7c (Fig. 2. – S22)



0.15 (s, 3 H, 3 \times 6-H); 0.16 (s, 3 H, 3 \times 6-H); 0.94 (s, 9 H,

$R_f = 0.9$ (DCM/MeOH 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.14 (s, 3 H, 3 \times 6-H); 0.15 (s, 3 H, 3 \times 6-H); 0.93 (s, 9 H, 9 \times 8-H); 1.86 (ddd, $J_{\text{gem}} = 14.3$ Hz, $J = 4.3$ Hz, $J = 1.3$ Hz, 1 H, 5a-H); 2.23 (ddd, $J_{\text{gem}} = 14.3$ Hz, $J = 6.6$ Hz, $J_{4-5b} = 5.5$ Hz, 1 H, 5b-H); 3.53 (dd, $J_{2-3} = J_{3-4} = 3.7$ Hz, 1 H, 3-H); 3.73 (t, $J_{1-2} = J_{2-3} = 4.8$ Hz, 1 H, 2-H); 3.82–3.90 (m, 1 H, 4-H); 4.21 (dd, $J = 4.8$ Hz, $J = 9.5$ Hz, 1 H, 1-H); 6.67 (d, $J_{11-12} = 8.9$ Hz, 2 H, 11-H); 7.26 (d, $J_{11-12} = 8.9$ Hz, 2 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.4 (7-C); 26.1 (8-C); 40.1 (5-C); 69.4 (3-C); 73.4 (1-C); 76.5 (4-C); 79.1 (2-C); 110.2 (13-C); 115.7 (11-C); 132.3 (12-C); 147.0 (10-C). MS (ESI $^+$): 402 [M + H] $^+$; 424 [M + Na] $^+$.

5.21.2. 8c (Fig. 2. – S23)

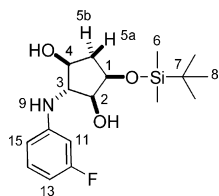


$R_f = 0.5$ (DCM/MeOH 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.06 (s, 3 H, 3 \times 6-H); 0.08 (s, 3 H, 3 \times 6-H); 0.90 (s, 9 H, 9 \times 8-H); 1.87–1.96 (m, 1 H, 5a-H); 2.17 (dt, $J_{\text{gem}} = 14.6$ Hz, $J_{1-5b} = J_{4-5b} = 5.5$ Hz, 1 H, 5b-H); 3.58–3.64 (m, 1 H, 2-H); 3.75 (t, $J_{1-2} = J_{2-3} = 5.03$ Hz, 1 H, 3-H); 3.94–3.99 (m, 1 H, 1-H); 4.13–4.19 (m, 1 H, 4-H); 6.61 (d, $J_{11-12} = 8.8$ Hz, 2 H, 11-H); 7.25 (d, $J_{11-12} = 8.8$ Hz, 2 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.2 (7-C); 26.1 (8-C); 39.8 (5-C); 68.9 (2-C); 72.7 (4-C); 77.6 (1-C); 79.3 (3-C); 101.1 (d, $J = 25.5$ Hz, Ar-C); 105.1 (d, $J = 21.5$ Hz, Ar-C); 110.1 (d, $J = 2.3$ Hz, Ar-C); 130.6 (d, $J = 0.1$ Hz, 15-C); 149.1 (d, $J = 10.4$ Hz, 10-C); 164.3 (d, $J = 243.3$ Hz, 12-C). MS (ESI $^+$): 342 [M + H] $^+$; 364 [M + Na] $^+$.

5.22. (\pm)-(1S,2R,3S,4R)-4-(tert-butyl dimethylsilyloxy)-2-(3-fluorophenylamino)cyclopentane-1,3-diol 7d and (\pm)-(1S,2R,3R,4R)-4-(tert-butyl dimethylsilyloxy)-3-(3-fluorophenylamino)cyclopentane-1,2-diol 8d

Starting from 70 mg (0.304 mmol) of epoxide **4**, we obtained a crude reaction product which was purified by flash chromatography (first $\text{CH}_2\text{Cl}_2/\text{n-hexane}$ 1/1, then $\text{CH}_2\text{Cl}_2/\text{AcOEt}$ from 95/5 to 70/30) to give **7d** (66 mg, 64%) and **8d** (22 mg, 21%) as a brown oil.

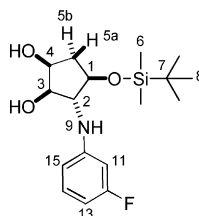
5.22.1. 7d (Fig. 2. – S24)



$R_f = 0.5$ (DCM/Hex 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.14 (s, 3 H, 3 \times 6-H); 0.15 (s, 3 H, 3 \times 6-H); 0.93 (s, 9 H, 9 \times 8-H); 1.87 (ddd, $J_{\text{gem}} = 14.3$ Hz, $J = 4.3$ Hz, $J = 1.2$ Hz, 1 H, 5a-H); 2.27 (ddd, $J_{\text{gem}} = 14.3$ Hz, $J = 6.9$ Hz, $J = 5.5$ Hz, 1 H, 5b-H); 2.74 (d, $J = 7.1$ Hz, 1 H, 2-OH); 3.58 (t, $J_{2-3} = J_{3-4} = 4.2$ Hz, 1 H,

3-H); 3.79–3.86 (m, 1 H, 2-H); 3.93–4.02 (m, 1 H, 4-H); 4.23 (dd, $J = 4.8$ Hz, $J = 9.5$ Hz, 1 H, 1-H); 6.54 (dt, $J = 1.5$ Hz, $J = 8.3$ Hz, 1 H, Ar-H); 6.61–6.70 (m, 2 H, Ar-H + 11-H); 7.11–7.22 (m, 1 H, Ar-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.4 (7-C); 26.1 (8-C); 40.1 (5-C); 69.3 (3-C); 73.2 (1-C); 76.4 (4-C); 78.8 (2-C); 101.0 (d, $J = 25.8$ Hz, Ar-C); 104.8 (d, $J = 21.8$ Hz, Ar-C); 109.9 (d, $J = 2.4$ Hz, Ar-C); 130.7 (d, $J = 0.1$ Hz, 15-C); 149.8 (d, $J = 10.9$ Hz, 10-C); 164.3 (d, $J = 245.4$ Hz, 12-C). MS (ESI $^+$): 342 [M + H] $^+$; 364 [M + Na] $^+$.

5.22.2. 8d (Fig. 2. – S25)

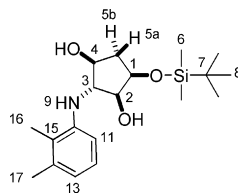


$R_f = 0.2$ (DCM/Hex 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.07 (s, 3 H, 3 \times 6-H); 0.08 (s, 3 H, 3 \times 6-H); 0.90 (s, 9 H, 9 \times 8-H); 1.92 (ddd, $J_{\text{gem}} = 14.7$ Hz, $J = 4.34$ Hz, $J = 2.81$ Hz, 1 H, 5a-H); 2.17 (dt, $J_{\text{gem}} = 14.7$ Hz, $J_{1-5b} = J_{4-5b} = 5.6$ Hz, 1 H, 5b-H); 2.58–3.49 (bs, 2 H, 4-OH + 3-OH); 3.61–3.67 (m, 1 H, 2-H); 3.78 (t, $J_{1-2} = J_{2-3} = 5.11$ Hz, 1 H, 3-H); 3.88–3.95 (m, 1 H, 1-H); 4.11–4.19 (m, 1 H, 4-H); 6.37–6.52 (m, 3 H, Ar-H); 7.10 (dt, $J = 8.2$ Hz, $J = 6.7$ Hz, 1 H, 14-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.7 (6-C); -4.3 (6-C); 18.2 (7-C); 26.1 (8-C); 39.8 (5-C); 68.9 (2-C); 72.7 (4-C); 77.6 (1-C); 79.3 (3-C); 101.1 (d, $J = 25.5$ Hz, Ar-C); 105.1 (d, $J = 21.5$ Hz, Ar-C); 110.1 (d, $J = 2.3$ Hz, Ar-C); 130.6 (d, $J = 0.1$ Hz, 15-C); 149.1 (d, $J = 10.4$ Hz, 10-C); 164.3 (d, $J = 243.3$ Hz, 12-C). MS (ESI $^+$): 342 [M + H] $^+$; 364 [M + Na] $^+$.

5.23. (\pm)-(1S,2R,3S,4R)-4-(tert-butyl dimethylsilyloxy)-2-(2,3-dimethylphenylamino)cyclopentane-1,3-diol 7e and (\pm)-(1S,2R,3R,4R)-4-(tert-butyl dimethylsilyloxy)-3-(2,3-dimethylphenylamino)cyclopentane-1,2-diol 8e

Starting from 50 mg (0.217 mmol) of epoxide **4**, we obtained a crude reaction product, which was purified with Biotage SP1 $^{\text{TM}}$ (n-hexane/AcOEt from 90/10 to 0/100; SNAP 10 g column) to give **7e** (40 mg, 52%) and **8e** (11 mg, 14%) as a brown oils.

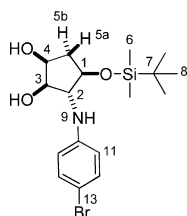
5.23.1. 7e (Fig. 2. – S26)



$R_f = 0.6$ (DCM/Hex 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.14 (s, 3 H, 3 \times 6-H); 0.15 (s, 3 H, 3 \times 6-H); 0.94 (s, 9 H, 9 \times 8-H); 1.89 (dt, $J_{\text{gem}} = 14.3$ Hz, $J = 3.96$ Hz, 1 H, 5a-H); 2.05 (s, 3 H, 3 \times 16-H); 2.28 (s + m, 4H, 5b-H + 3 \times 17-H); 2.66 (d, $J = 9.4$ Hz, 1 H, 4-OH); 2.71 (d, $J = 6.7$ Hz, 1 H, 2-OH); 3.42–3.59 (m, 1 H, 3-H); 3.59–3.67 (m, 1 H, 2-H); 3.83 (dt,

$J = 4.5$ Hz, $J = 6.6$ Hz, 1 H, 4-H); 3.88–3.98 (m, 1 H, 1-H); 6.65 (d, $J_{11-12} = 7.5$ Hz, 1 H, 11-H); 6.84 (d, $J_{12-13} = 8.1$ Hz, 1 H, 13-H); 7.07 (dd, $J_{11-12} = 7.5$ Hz, $J_{12-13} = 8.1$ Hz, 1 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 12.9 (16-C); 18.4 (7-C); 21.0 (17-C); 26.1 (8-C); 40.1 (5-C); 69.4 (3-C); 73.6 (1-C); 77.0 (4-C); 79.3 (2-C); 110.1 (13-C); 120.5 (11-C); 121.1 (14-C); 126.7 (12-C); 136.8 (15-C); 145.9 (10-C). MS (ESI⁺): 352 [M + H]⁺; 374 [M + Na]⁺.

5.23.2. **8e** (Fig. 2. – S27)

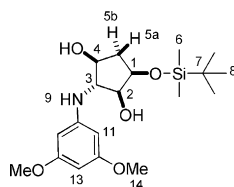


$R_f = 0.3$ (DCM/Hex 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.10 (s, 3 H, 3 × 6-H); 0.11 (s, 3 H, 3 × 6-H); 0.92 (s, 9 H, 9 × 8-H); 1.95 (m, 1 H, 5a-H); 2.03 (s, 3 H, 3 × 16-H); 2.17 (dt, $J_{\text{gem}} = 14.5$ Hz, $J_{1-5b} = J_{4-5b} = 5.3$ Hz, 1 H, 5b-H); 2.28 (s, 3 H, 3 × 17-H); 3.14 (bs, 3 H, 2-OH + 4-OH + 9-NH); 3.65–3.71 (m, 1 H, 2-H); 3.84 (t, 1 H, 3-H, $J_{2-3} = J_{3-4} = 4.5$ Hz); 4.02–4.08 (m, 1 H, 1-H); 4.17–4.24 (m, 1 H, 4-H); 6.64 (d, $J_{11-12} = 7.5$ Hz, 1 H, 11-H); 6.70 (d, $J_{12-13} = 8.0$ Hz, 1 H, 13-H); 7.02 (dd, $J_{11-12} = 7.5$ Hz, $J_{12-13} = 8.0$ Hz, 1 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 12.9 (16-C); 18.2 (7-C); 21.0 (17-C); 26.1 (8-C); 39.7 (5-C); 69.3 (2-C); 73.4 (4-C); 78.3 (1-C); 80.3 (3-C); 110.1 (13-C); 120.5 (11-C); 121.1 (14-C); 126.5 (12-C); 137.0 (15-C); 145.4 (10-C). MS (ESI⁺): 352 [M + H]⁺; 374 [M + Na]⁺.

5.24. (\pm)-(1*S*,2*R*,3*S*,4*R*)-4-(*tert*-butyldimethylsilyloxy)-2-(3,5-dimethoxyphenylamino)cyclopentane-1,3-diol **7f** and (\pm)-(1*S*,2*R*,3*R*,4*R*)-4-(*tert*-butyldimethylsilyloxy)-3-(3,5-dimethoxyphenylamino)cyclopentane-1,2-diol **8f**

Starting from 100 mg (0.434 mmol) of epoxide **2**, we obtained a crude reaction product which was purified with Biotage SP1TM (n-hexane/AcOEt from 90/10 to 0/100; SNAP 50 g column) to give a mixture of **7f** (96 mg, 58%) and **8f** (36 mg, 22%) as brown oils.

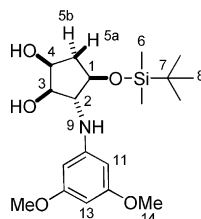
5.24.1. **7f** (Fig. 2. – S28)



$R_f = 0.6$ (Hexane/EtOAc 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.13 (s, 3 H, 3 × 6-H); 0.14 (s, 3 H, 3 × 6-H); 0.93 (s, 9 H, 9 × 8-H); 1.86 (ddd, $J_{\text{gem}} = 14.2$ Hz, $J = 4.4$ Hz, $J = 1.1$ Hz, 1 H, 5a-H); 2.26 (ddd, $J_{\text{gem}} = 14.2$ Hz, $J = 6.6$ Hz, $J = 5.5$ Hz, 1 H, 5b-H); 3.55 (t, $J = 3.8$ Hz, 1 H, 3-H); 3.74 (s, 3 H, 14-H); 3.75 (s, 3 H, 14-H); 3.76–3.80 (m, 1 H, 2-H); 3.88–3.93 (m, 1 H,

4-H); 4.21 (dd, $J = 9.8$ Hz, $J = 4.8$ Hz, 1 H, 1-H); 5.91–5.95 (t, $J_{11-13} = 2.1$ Hz, 1 H, 13-H); 6.03 (d, $J_{11-13} = 2.1$ Hz, 2 H, 11-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -4.6 (6-C); -4.3 (6-C); 18.4 (7-C); 26.1 (8-C); 40.1 (5-C); 55.5 (14-C); 69.6 (3-C); 73.3 (1-C); 76.5 (4-C); 78.9 (2-C); 91.4 (13-C); 94.2 (11-C); 148.5 (10-C); 162.1 (12-C). MS (ESI⁺): 384 [M + H]⁺.

5.24.2. **8f** (Fig. 2. – S29)

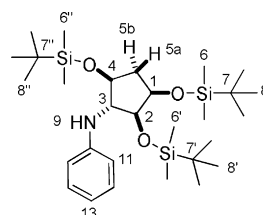


$r_f = 0.3$ (Hexane/EtOAc 5/5). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.06 (s, 3 H, 3 × 6-H); 0.07 (s, 3 H, 3 × 6-H); 0.88 (s, 9 H, 9 × 8-H); 1.90 (ddd, $J_{\text{gem}} = 14.6$ Hz, $J_{4-5a} = 4.3$ Hz, $J_{1-5a} = 2.9$ Hz, 1 H, 5a-H); 2.17 (dt, $J_{\text{gem}} = 14.7$ Hz, $J_{1-5b} = J_{4-5b} = 5.6$ Hz, 1 H, 5b-H); 3.63–3.68 (m, 1 H, 2-H); 3.75 (s, 3 H, 14-H); 3.83 (t, $J_{2-3} = J_{3-4} = 4.9$ Hz, 1 H, 3-H); 4.04 (dt, $J_{1-5b} = 5.6$ Hz, $J_{1-5a} = J_{1-2} = 2.9$ Hz, 1 H, 1-H); 4.13–4.18 (m, 1 H, 4-H); 5.96 (t, $J_{11-13} = 2.1$ Hz, 1 H, 13-H); 5.99 (d, $J_{11-13} = 2.1$ Hz, 2 H, 11-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): -5.0 (6-C); -4.7 (6-C); 17.8 (7-C); 25.7 (8-C); 39.5 (5-C); 55.2 (14-C); 69.0 (2-C); 72.4 (4-C); 77.2 (1-C); 78.8 (3-C); 91.2 (13-C); 93.4 (11-C); 148.1 (10-C); 161.8 (12-C). MS (ESI⁺): 384 [M + H]⁺.

5.25. (\pm)-2-chloro-*N*-phenyl-*N*-((1*R*,2*S*,3*R*,5*S*)-2,3,5-tris(*tert*-butyldimethylsilyloxy)cyclopentyl) acetamide **10**

2,6-Lutidine (0.224 mL, 1.920 mmol) and TBDMSOTf (372 mg, 1.408 mmol) were added at 0 °C to a stirred solution of **7a** (207 mg, 0.640 mmol) and **8a** (207 mg, 0.640 mmol) in CH_2Cl_2 (4 mL). The mixture was stirred for 1 h at room temperature, and then the reaction was quenched with a saturated NH_4Cl solution. The organic layer was washed once with a saturated NH_4Cl solution and twice with brine, dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude product was purified by flash chromatography (n-hexane/ CH_2Cl_2 from 100/0 to 88/12) to give pure silylether intermediate (577 mg, 82%) as a colorless oil.

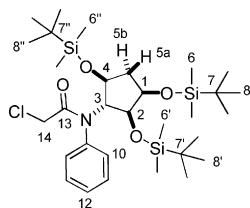
5.26. Silylether intermediate (Fig. 2. – S30)



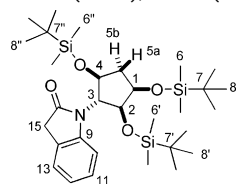
$R_f = 0.9$ (Hexane/EtOAc 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): -0.05 + 0.00 + 0.04 + 0.06 + 0.07 + 0.08 (6 s, 6 × 3 H, 6 × 6-H + 6 × 6'-H + 6 × 6''-H); 0.87 + 0.88 + 0.92 (3 s, 3 × 9 H, 9 × 8-H + 9 × 8'-H + 9 × 8''-H); 1.81 (ddd, $J_{\text{gem}} = 13.4$ Hz, J_{1-5a}

$5_a = J_{4-5a} = 5.7$ Hz, 1 H, 5a-H); 2.20 (ddd, $J_{gem} = 13.4$ Hz, $J = 8.2$ Hz, $J = 5.6$ Hz, 1 H, 5b-H); 3.64–3.70 (m, 1 H, 2-H); 3.73 (dd, $J_{2-3} = J_{3-4} = 5.0$ Hz, 1 H, 3-H); 3.81–3.89 (m, 1 H, 4-H); 3.93–4.02 (m, 1 H, 1-H); 6.66–6.82 (m, 3 H, 2 × 11-H + 13-H); 7.15 (dd, $J = 8.7$ Hz, $J = 7.3$ Hz, 2 H, 12-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-4.6 + -4.4 + -4.2 + -4.1 + -3.9$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 18.2 + 18.48 + 18.49 ($7\text{-C} + 7'\text{-C} + 7''\text{-C}$); 26.1 + 26.2 + 26.3 ($8\text{-C} + 8'\text{-C} + 8''\text{-C}$); 40.8 (5-C); 67.6 (3-C, only on HSQC); 73.0 (1-C); 76.6 (4-C, only on HSQC); 78.5 (2-C, only on HSQC); 114.4 (11-C, only on HSQC); 117.9 (13-C, only on HSQC); 129.5 (12-C, only on HSQC); (10-C, not visible on HSQC). MS (ESI⁺): 552 [M + H]⁺.

Chloroacetyl chloride (145 μL , 1.812 mmol) and 2,6-lutidine (264 μL , 2.264 mmol) were added to a stirred solution of silylether intermediate (500 mg, 0.906 mmol) in dry toluene (2 mL) under N_2 atmosphere. The reaction was stirred for 45 min at room temperature, then was diluted with CH_2Cl_2 and washed three times with water. The organic layer was dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude residue was purified with Biotage SP1TM (n-hexane/EtOAc from 98/2 to 80/20; SNAP 50 g column) to give pure **10** (545 mg, 96%) as a white solid.



$R_f = 0.64$ (Hexane/EtOAc 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): 0.03 + 0.04 + 0.05 + 0.07 (6 s, 6 × 3 H, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.84 + 0.90 (3 s, 3 × 9 H, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.52–1.67 (m, 1 H, 5a-H); 2.24 (ddd, 1 H, 5b-H, $J_{gem} = 13.5$ Hz, $J = 8.4$ Hz, $J = 4.7$ Hz); 3.81 (d, 1 H, 17a-H, $J_{gem} = 13.2$ Hz); 3.88 (d, 1 H, 17b-H, $J_{gem} = 13.2$ Hz); 3.93 (dd, 1 H, 3-H, $J_{2-3} = 6.7$ Hz, $J_{3-4} = 8.9$ Hz); 4.03 (dd, 1 H, 1-H, $J = 7.3$ Hz, $J_{3-4} = 4.3$ Hz); 4.73 (dd, 1 H, 2-H, $J = 8.9$ Hz, $J_{3-4} = 4.1$ Hz); 4.81–4.89 (m, 1 H, 4-H); 7.28–7.43 (m, 5H, Ar-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-4.5 + -4.3 + -4.2 + -4.0 + -3.9 + -3.8$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 18.1 + 18.3 + 18.4 ($7\text{-C} + 7'\text{-C} + 7''\text{-C}$); 26.1 + 26.3 + 26.4 ($8\text{-C} + 8'\text{-C} + 8''\text{-C}$); 40.9 (5-C); 43.3 (14-C); 69.2 (4-C); 73.0 (1-C); 76.6 (2-C); 78.5 (3-C); 127.9 (Ar-C); 128.2 (Ar-C); 129.7 (Ar-C); 144.6 (10-C); 166.5 (13-C). MS (ESI⁺): 650 [M + Na]⁺.



5.27. (\pm)-1-((1R,2S,3R,5S)-2,3,5-tris(tert-butyl dimethylsilyloxy)cyclopentyl)indolin-2-one **12** (Fig. 2. – S32)

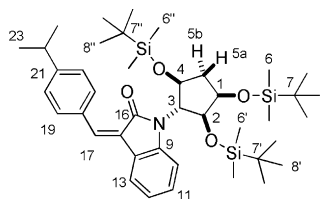
Palladium(II) acetate (18.86 mg, 0.084 mmol) was added under N_2 atmosphere to a stirred solution of **10** (264 mg,

0.420 mmol), di-tert-butyl(naphthalen-1-yl)phosphine (252 mg, 0.924 mmol) in toluene (1 mL). The mixture was stirred for 6 h at 100 °C, then filtered through a celite pad and evaporated under reduced pressure. The residue was purified by flash chromatography (n-hexane/ CH_2Cl_2 from 95/5 to 40/60) to give **12** (174 mg, 70%) as a white solid. $R_f = 0.3$ (Hexane/EtOAc 93/7). $^1\text{H-NMR}$ (400 MHz, CDCl_3): $-0.27 + -0.25 + -0.12 + -0.07 + 0.09 + 0.10$ (6 s, 6×3 H, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.69 + 0.74 + 0.97 (3 s, 3×9 H, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.66–1.75 (m, 1 H, 5a-H); 2.29 (ddd, 1 H, 5b-H, $J_{gem} = 14.3$ Hz, $J = 8.6$ Hz, $J = 4.4$ Hz); 3.42 (d, 1 H, 17a-H, $J_{gem} = 22.4$ Hz); 3.48 (d, 1 H, 17b-H, $J_{gem} = 22.4$ Hz); 4.01–4.08 (m, 1 H, 1-H); 4.39 (dd, 1 H, 3-H, $J_{2-3} = 9.3$ Hz, $J_{3-4} = 6.3$ Hz); 4.57 (dd, 1 H, 2-H, $J_{2-3} = 9.3$ Hz, $J_{1-2} = 4.1$ Hz); 4.79–4.86 (m, 1 H, 4-H); 6.89 (d, 1 H, 13-H, $J = 8.0$ Hz); 6.96 (t, 1 H, 12-H, $J = 7.5$ Hz); 7.18 (d, 1 H, 10-H, $J = 7.3$ Hz); 7.20–7.26 (m, 1 H, 11-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-5.0 + -4.9 + -4.7 + -4.2 + -4.3 + -4.0$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 18.1 + 18.5 ($7\text{-C} + 7'\text{-C} + 7''\text{-C}$); 25.9 + 26.0 + 26.3 ($8\text{-C} + 8'\text{-C} + 8''\text{-C}$); 36.7 (15-C); 40.9 (5-C); 67.7 (3-C); 69.1 (4-C); 72.6 (1-C); 73.5 (2-C); 109.3 (13-C); 122.1 (12-C); 124.3 (14-C); 124.4 (10-C); 128.1 (11-C); 146.5 (9-C); 176.1 (16-C). MS (ESI⁺): 614 [M + Na]⁺. HRMS: $\text{C}_{31}\text{H}_{57}\text{NO}_4\text{Si}_3\text{Na}$: calcd. 614.34876; found 614.34797.

5.28. (\pm)-(*Z*)-3-(4-isopropylbenzylidene)-1-((1R,2S,3R,5S)-2,3,5-tris(tert-butyl dimethylsilyloxy)cyclopentyl)indolin-2-one **15Z** and (\pm)-(*E*)-3-(4-isopropylbenzylidene)-1-((1R,2S,3R,5S)-2,3,5-tris(tert-butyl dimethylsilyloxy)cyclopentyl)indolin-2-one **15E**

4-Isopropylbenzaldehyde (14 μL , 0.093 mmol) and piperidine (0.8 μL , 8.45 μmol) were added to a stirred solution of **12** (50 mg, 0.084 mmol) in MeOH (5 mL). The reaction was stirred at room temperature for 2 h to give a yellow solution. After reaction completion, the solvent was evaporated under reduced pressure and the residue was purified with Biotage SP1TM (n-hexane/ CH_2Cl_2 from 90/10 to 0/100; SNAP 10 g column) to give **15Z** (10 mg, 16%) and **15E** (39 mg, 64%), as yellow solids.

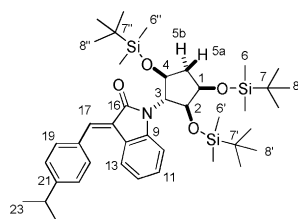
5.28.1. **15Z** (Fig. 2. – S33)



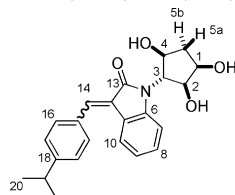
$R_f = 0.7$ (Hexane/DCM 6/4). $^1\text{H-NMR}$ (400 MHz, CDCl_3): $-0.28 + -0.26 + -0.13 + -0.07 + 0.10 + 0.11$ (6 s, 6×3 H, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.68 + 0.74 + 0.98 (3 s, 3×9 H, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.28 (d, 6 H, 23-H, $J = 6.9$ Hz); 1.71 (ddd, 1 H, 5a-H, $J_{gem} = 14.3$ Hz, $J_{4-5a} = 2.6$ Hz, $J_{1-5a} = 2.0$ Hz); 2.33 (ddd, 1 H, 5b-H, $J_{gem} = 14.3$ Hz, $J_{4-5b} = 8.5$ Hz, $J_{1-5b} = 4.3$ Hz); 2.96 (sp, 1 H, 22-H, $J = 6.9$ Hz); 4.08 (ddd, 1 H, 1-H, $J_{1-5b} = 4.1$ Hz, $J_{1-2} = 4.0$ Hz, $J_{1-5a} = 2.0$ Hz); 4.38–4.52 (m, 1 H, 3-H); 4.69 (dd, 1 H, 2-H, $J_{2-3} = 9.4$ Hz, $J_{1-2} = 4.0$ Hz); 4.88 (ddd, 1 H, 4-H, J_{4-}

$5b = 8.5$ Hz, $J_{3-4} = 6.3$ Hz, $J_{4-5a} = 2.6$ Hz); 6.88 (d, 1 H, 13-H, $J = 7.9$ Hz); 6.98 (dt, 1 H, 11-H, $J = 7.6$ Hz, $J = 0.9$ Hz); 7.22 (dt, 1 H, 12-H, $J = 7.8$ Hz, $J = 1.2$ Hz); 7.32 (d, 2 H, 20-H, $J = 8.3$ Hz); 7.47 (dd, 1 H, 10-H, $J = 7.6$ Hz, $J = 0.7$ Hz); 7.50 (s, 1 H, 17-H) 8.20 (d, 1 H, 19-H, $J = 8.3$ Hz). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-4.9 + -4.8 + -4.7 + -4.3 + -4.2 + -4.0$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 18.1 + 18.5 (7-C + 7'-C + 7''-C); 24.0 + 24.1 + 24.2 (8-C + 8'-C + 8''-C); 40.9 (5-C); 67.8 (3-C); 69.3 (4-C); 72.7 (1-C); 73.4 (2-C); 109.0 (13-C); 118.9 (10-C); 121.8 (12-C); 127.0 (20-C); 129.9 (11-C); 132.1 (9-C); 132.3 (19-C); 137.0 (17-C); 152.0 (15-C); 169.4(16-C). MS (ESI⁺): 744 [M + Na]⁺.

5.28.2. 15E (Fig. 2. – S34)



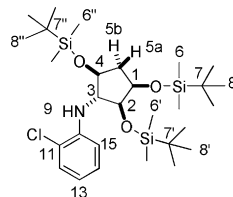
$R_f = 0.4$ (Hexane/EtOAc 6/4). $^1\text{H-NMR}$ (400 MHz, CDCl_3): $-0.26 + -0.25 + -0.11 + -0.06 + 0.10 + 0.12$ (6 s, 6×3 H, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.69 + 0.76 + 0.98 (3 s, 3×9 H, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.30 (d, 6 H, 23-H, $J = 6.9$ Hz); 1.73 (ddd, 1 H, 5a-H, $J_{\text{gem}} = 14.1$ Hz, $J_{4-5a} = 2.9$ Hz, $J_{1-5a} = 2.6$ Hz); 2.32 (ddd, 1 H, 5b-H, $J_{\text{gem}} = 14.1$ Hz, $J_{4-5b} = 8.6$ Hz, $J_{1-5b} = 4.4$ Hz); 2.98 (sp, 1 H, 22-H, $J = 6.9$ Hz); 4.07–4.11 (m, 1 H, 1-H); 4.42 (dd, 1 H, 3-H, $J_{2-3} = 9.0$ Hz, $J_{3-4} = 6.3$ Hz); 4.62 (dd, 1 H, 2-H, $J_{2-3} = 9.0$ Hz, $J_{1-2} = 3.9$ Hz); 4.88 (ddd, 1 H, 4-H, $J_{4-5b} = 8.6$ Hz, $J_{3-4} = 6.3$ Hz, $J_{4-5a} = 2.9$ Hz); 6.84 (dt, 1 H, 11-H, $J = 7.6$ Hz, $J = 1.0$ Hz); 6.91 (d, 1 H, 13-H, $J = 8.1$ Hz); 7.22 (dt, 1 H, 12-H, $J = 7.8$ Hz, $J = 1.2$ Hz); 7.32 (d, 2 H, 20-H, $J = 8.4$ Hz); 7.63 (d, 1 H, 19-H, $J = 7.9$ Hz); 7.71 (d, 1 H, 10-H, $J = 6.6$ Hz); 7.74 (s, 1 H, 17-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-4.9 + -4.8 + -4.7 + -4.3 + -4.2 + -4.0$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 18.1 + 18.5 (7-C + 7'-C + 7''-C); 25.9 + 26.0 + 26.3 (8-C + 8'-C + 8''-C); 41.0 (5-C); 67.9 (3-C); 69.7 (4-C); 72.6 (1-C); 73.9 (2-C); 109.4 (13-C); 121.5 (12-C); 122.9 (10-C); 127.0 (20-C); 129.9 (11-C); 130.0 (19-C); 132.9 (9-C); 136.9 (17-C); 151.2 (15-C); 169.4(16-C). MS (ESI⁺): 744 [M + Na]⁺.



5.29. (±)-(Z and E)-3-(4-isopropylbenzylidene)-1-((1R,2S,3R,5S)-2,3,5-trihydroxycyclopentyl)indolin-2-one 16

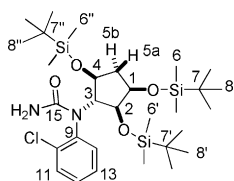
TBAF (0.127 mL, 0.127 mmol, 1 M solution in THF) was added to a stirred solution of **15Z** and **15E** mixture (30 mg, 0.041 mmol). The solvent was evaporated under reduced pressure and taken up with CH_2Cl_2 . The organic layer was washed four times with water, the combined organic

layers were then evaporated under reduced pressure. The residue was purified with Biotage SP1TM (n-hexane/acetone from 88/12 to 0/100; SNAP 10 g column) to give **16** in a Z/E 40/60 mixture (11.6 mg, 74%) as a yellow solid. $R_f = 0.3 + 0.2$ (Hexane/Acetone 5/5). $^1\text{H-NMR}$ (400 MHz, CD_3OD): 1.46 (d, 6 H, 23-H Z-isomer, $J = 6.9$ Hz); 1.48 (d, 6 H, 23-H, Z-isomer, $J = 6.9$ Hz); 1.94–2.02 (m, 1 H, 5a-H); 2.69 (ddd, 1 H, 5b-H, $J_{\text{gem}} = 14.2$ Hz, $J = 8.4$ Hz, $J = 5.6$ Hz); 3.15 (sp, 1 H, 19-H, $J = 6.9$ Hz); 4.31–4.38 (m, 1 H, 1-H); 4.64 (dd, 1 H, 3-H, $J = 15.5$ Hz, $J = 7.0$ Hz); 4.81 (dd, 1 H, 2-H, $J = 8.8$ Hz, $J = 4.0$ Hz); 4.85–4.93 (m, 1 H, 4-H); 7.03–7.12 (m, 1 H, 9-H Z-isomer); 7.18–7.26 (m, 2 H, 9-H Z-isomer + 10-H E-isomer); 7.25–7.31 (d, 1 H, 10-H E-isomer, $J = 8.0$ Hz); 7.40–7.51 (m, 2 H, Ar-H); 7.52–7.60 (m, 1 H, Ar-H); 7.76–7.92 (m, 3 H, Ar-H); 8.44 (d, 1 H, 14-H Z-isomer, $J = 8.3$ Hz). $^{13}\text{C-NMR}$ (100.61 MHz, CD_3OD): 24.1 + 24.2 (20-C); 35.4 + 35.5 (19-C); 40.2 (5-C); 67.8 + 67.8 (3-C); 69.8 (4-C); 71.0 + 71.1 (1-C); 73.1 (2-C); 109.9 + 110.6 (10-C); 118.9 (10-C); 129.7 (Ar-CH); 130.7 + 130.8 (Ar-CH); 133.3 (C^{IV}); 133.5 (Ar-CH); 133.6 (C^{IV}); 138.3 (Ar-CH); 138.5 (Ar-CH); 143.6 + 145.6 (C^{IV}); 152.6 + 153.2 (12-C); 168.3 + 170.7 (16-C). MS (ESI⁺): 744 [M + Na]⁺. HRMS: $\text{C}_{23}\text{H}_{25}\text{NO}_4\text{Na}$: calcd. 402.16758; found 402.16725.



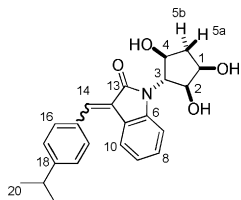
5.30. (±)-2-chloro-N-((1R,2S,3R,5S)-2,3,5-tris(tert-butyl dimethylsilyloxy)cyclopentyl)aniline 19 (Fig. 2. – S36)

TBDMSOTf (302 μL , 1.314 mmol) was added at 0 °C to a solution of 2,6-lutidine (218 μL , 1.877 mmol), **7b** (224 mg, 0.626 mmol) and **8b** (224 mg, 0.626 mmol) in dry THF (4 mL). The solution was stirred for 2 h, the solvent was removed under reduced pressure and the residue was taken up in Et_2O (4 mL). The organic layer was washed two times with water, dried over Na_2SO_4 , filtered and evaporated under reduced pressure. The residue was purified by flash chromatography on silica gel (n-hexane/ CH_2Cl_2 from 100/0 to 85/15) to give pure **19** (493 mg, 68%) as a white solid. $R_f = 0.5$ (Hexane/DCM 9/1). $^1\text{H-NMR}$ (400 MHz, CDCl_3): $-0.09 + -0.01 + -0.01 + 0.05 + 0.08$ (5 s, 6×3 H, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.85 + 0.86 + 0.93 (3 s, 3×9 H, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.75 (dt, 1 H, 5a-H, $J_{\text{gem}} = 13.8$ Hz, $J = 4.4$ Hz); 2.15 (ddd, 1 H, 5b-H, $J_{\text{gem}} = 13.8$ Hz, $J = 8.3$ Hz, $J = 5.3$ Hz); 3.65 (dd, 1 H, 2-H, $J = 7.2$ Hz, $J = 3.7$ Hz); 3.84–3.95 (m, 2 H, 3-H + 4-H); 3.98 (dd, 1 H, 1-H, $J = 9.0$ Hz, $J = 4.1$ Hz); 6.59 (ddd, 1 H, 13-H, $J = 7.9$ Hz, $J = 7.4$ Hz, $J = 1.5$ Hz); 7.00 (dd, 1 H, 15-H, $J = 8.3$ Hz, $J = 1.5$ Hz); 7.05–7.10 (m, 1 H, 14-H); 7.20 (dd, 1 H, 12-H, $J = 7.9$ Hz, $J = 1.5$ Hz). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-4.7 + -4.5 + -4.2 + -4.1 + -4.0 + -3.9$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 18.2 + 18.47 + 18.52 (7-C + 7'-C + 7''-C); 26.1 + 26.2 + 26.3 (8-C + 8'-C + 8''-C); 41.0 (5-C); 66.5 (3-C); 72.8 (1-C); 77.4 (4-C); 78.7 (2-C); 113.6 (11-C); 117.3 (13-C); 119.0 (15-C); 127.9 (12-C); 129.0 (14-C); 144.5 (10-C). MS (ESI⁺): 586 [M + H]⁺.



5.31. (\pm)-2-chloro-N-((1R,2S,3R,5S)-2,3,5-tris(tert-butyl dimethylsilyloxy)cyclopentyl)aniline **20**

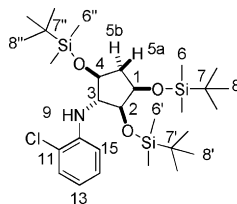
A solution of **19** (363 mg, 0.619 mmol) in THF (2 mL) was slowly added to a stirred solution of Chlorosulfonyl isocyanate (0.081 mL, 0.928 mmol) in THF (1 mL) at -10°C . The reaction mixture was stirred 10 min at -10°C , then quenched with water and stirred for 30 min, then NaOH (3 M) was added. The organic layer was diluted with AcOEt and washed 2 times with brine, dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The residue was purified with Biotage SP1TM (n-hexane/AcOEt from 100/0 to 60/40, SNAP 10 g column) to give pure **20** (362 mg, 93%) as a white solid. $R_f = 0.5$ (Hexane/EtOAc 7/3). $^1\text{H-NMR}$ (400 MHz, C_6D_6): 0.09 + 0.11 + 0.12 + 0.14 + 0.17 + 0.18 + 0.21 + 0.22 + 0.26 + 0.27 (10 s for 2 rotamers, $6 \times 3 \text{ H}$, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.93 + 0.95 + 0.99 + 1.02 + 1.11 (5 s for 2 rotamers, $3 \times 9 \text{ H}$, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.70–1.81 (m, 0.6 H for 1 rotamer, 5a-H); 2.12–2.37 (m, 1.4H, $1 \times 5b\text{-H} + 0.4 \times 5a\text{-H}$); 3.92 (d, 0.4H, 3-H, $J = 3.9 \text{ Hz}$); 4.07 (bs, 2 H, NH_2); 4.14 (dd, 0.6 H, 3-H, $J = 8.5 \text{ Hz}$, $J = 5.7 \text{ Hz}$); 4.76–4.89 (bs, 1 H, 1-H); 4.94–5.05 (m, 1 H, 2-H); 5.12–5.24 (m, 1 H, 4-H); 6.56–6.71 (m, 1 H, Ar-H); 6.78–6.93 (m, 1 H, Ar-H); 7.05 (d, 1 H, Ar-H, $J = 8.0 \text{ Hz}$); 7.65 (d, 0.4H, Ar-H, $J = 7.8 \text{ Hz}$); 7.70 (d, 0.6H, Ar-H, $J = 7.8 \text{ Hz}$). $^{13}\text{C-NMR}$ (100.61 MHz, C_6D_6): $-4.6 + -4.4 + -4.3 + -4.0 + -3.9 + -3.8 + -3.3$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$ for the 2 rotamers); 18.1 + 18.4 + 18.5 ($7\text{-C} + 7'\text{-C} + 7''\text{-C}$); 26.1 + 26.2 + 26.3 + 26.5 + 26.6 ($8\text{-C} + 8'\text{-C} + 8''\text{-C}$ for the 2 rotamers); 40.5 + 40.6 (5-C for the 2 rotamers); 71.5 + 72.0 + 72.1 + 72.6 + 73.7 + 78.3 + 78.4 + 80.9 ($3\text{-C} + 4\text{-C} + 1\text{-C} + 2\text{-C}$ for the 2 rotamers); 128.2 (Ar-C); 128.4 (Ar-C); 130.5 (Ar-C); 131.0 (Ar-C); 133.1 (Ar-C); 142.8 (Ar-C); 156.4 (15-C). MS (ESI⁺): 629 [M + H]⁺.



5.32. (\pm)-1-((1R,2S,3R,5S)-2,3,5-tris(tert-butyl dimethylsilyloxy)cyclopentyl)1H-benzo[d]imidazol-2(3H)-one **22**

DIPEA (0.195 mL, 1.122 mmol), palladium(II) acetate (8 mg, 0.036 mmol) and 2-dicyclohexylphosphino-2',4',6'-triisopropylbiphenyl (178 mg, 0.373 mmol) were added to a stirred solution of **20** (235 mg, 0.373 mmol) in 2-propanol (3 mL) under N_2 atmosphere at room temperature. The mixture was stirred at reflux overnight, and then

the crude product was taken up in CH_2Cl_2 and filtered on a celite pad. The filtrate was washed twice with water, dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude was purified with Biotage SP1TM (n-hexane/EtOAc from 90/10 to 50/50) to give pure **22** (187 mg, 69%) as a yellow oil. $R_f = 0.5$ (Hexane/EtOAc 8/2). $^1\text{H-NMR}$ (400 MHz, CDCl_3): $-0.34 + -0.14 + -0.09 + 0.11 + 0.12$ (6 s , $6 \times 3 \text{ H}$, $6 \times 6\text{-H} + 6 \times 6'\text{-H} + 6 \times 6''\text{-H}$); 0.65 + 0.73 + 0.98 (3 s , $3 \times 9 \text{ H}$, $9 \times 8\text{-H} + 9 \times 8'\text{-H} + 9 \times 8''\text{-H}$); 1.72 (ddd, $J_{\text{gem}} = 14.2 \text{ Hz}$, $J_{4-5a} = 2.6 \text{ Hz}$, $J_{1-5a} = 2.0 \text{ Hz}$, 1 H, 5a-H); 2.34 (ddd, $J_{\text{gem}} = 14.2 \text{ Hz}$, $J_{4-5b} = 8.6 \text{ Hz}$, $J_{1-5b} = 4.4 \text{ Hz}$, 1 H, 5b-H); 4.08 (bs, 1 H, 1-H); 4.56 (dd, $J_{2-3} = 9.6 \text{ Hz}$, $J_{3-4} = 6.0 \text{ Hz}$, 1 H, 3-H); 4.61 (dd, $J_{2-3} = 9.6 \text{ Hz}$, $J_{1-2} = 4.1 \text{ Hz}$, 1 H, 2-H); 4.90 (ddd, $J_{4-5b} = 8.6 \text{ Hz}$, $J_{3-4} = 6.0 \text{ Hz}$, $J_{4-5a} = 2.6 \text{ Hz}$, 1 H, 4-H); 6.99–7.09 (m, 4H, 10-H + 11-H + 12-H + 13-H); 8.96 (bs, 1 H, 15-H). $^{13}\text{C-NMR}$ (100.61 MHz, CDCl_3): $-5.5 + -5.3 + -5.1 + -4.7 + -4.6 + -4.4$ ($2 \times 6\text{-C} + 2 \times 6'\text{-C} + 2 \times 6''\text{-C}$); 17.7 + 17.8 + 18.1 ($7\text{-C} + 7'\text{-C} + 7''\text{-C}$); 25.5 + 25.6 + 25.9 ($8\text{-C} + 8'\text{-C} + 8''\text{-C}$); 40.5 (5-C); 67.6 (3-C); 69.1 (4-C); 72.3 (1-C); 73.3 (2-C); 108.4 (13-C); 108.9 (12-C); 121.0 (10-C); 121.3 (11-C); 127.2 (14-C); 132.1 (9-C); 154.8 (16-C). MS (ESI⁺): 593 [M + H]⁺; 615 [M + Na]⁺.



5.33. (\pm)-1-((1R,2S,3R,5S)-2,3,5-trihydroxycyclopentyl)1H-benzo[d]imidazol-2(3H)-one **23** (Fig. 2. – S39)

Hydrochloric acid (0.464 mmol, 23 μL , 20 M solution in MeOH) was added to a stirred solution of **22** (83.3 mg, 0.140 mmol) in MeOH (3 mL). The reaction was stirred for 3 h at room temperature, and then the solvent was removed under reduced pressure. The residue was taken up with distilled water and washed three times with CH_2Cl_2 . The aqueous phase was evaporated under reduced pressure and pure **23** (35 mg, quantitative yield) was obtained as a white solid without further purification. $^1\text{H-NMR}$ (400 MHz, CD_3OD): 1.82 (ddd, $J_{\text{gem}} = 14.4 \text{ Hz}$, $J_{4-5a} = 5.4 \text{ Hz}$, $J_{1-5a} = 3.4 \text{ Hz}$, 1 H, 5a-H); 2.55 (ddd, $J_{\text{gem}} = 14.4 \text{ Hz}$, $J_{1-5b} = 8.5 \text{ Hz}$, $J_{4-5b} = 5.4 \text{ Hz}$, 1 H, 5b-H); 4.15–4.20 (m, 1 H, 1-H); 4.56 (dd, $J_{2-3} = 8.9 \text{ Hz}$, $J_{3-4} = 7.7 \text{ Hz}$, 1 H, 3-H); 4.62 (dd, $J_{2-3} = 8.9 \text{ Hz}$, $J_{1-2} = 5.0 \text{ Hz}$, 1 H, 2-H); 4.67–4.77 (m, 1 H, 4-H); 6.32–6.44 (m, 3 H, $3 \times \text{Ar-H}$); 6.98–7.06 (m, 1 H, Ar-H). $^{13}\text{C-NMR}$ (100.61 MHz, CD_3OD): 40.0 (5-C); 67.7 (3-C); 69.8 (4-C); 70.8 (1-C); 73.1 (2-C); 109.8 (13-C); 110.3 (12-C); 122.3 (10-C); 122.4 (11-C); 129.6 (14-C); 132.2 (9-C); 156.8 (16-C). MS (ESI⁺): 273 [M + Na]⁺; 523 [2M + Na]⁺. HRMS: $\text{C}_{12}\text{H}_{14}\text{N}_2\text{O}_4\text{Na}$: calcd. 273.08458; found 273.08455.

6. Conflicts of interest

There is no conflict of interest for all the authors.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.crci.2009.11.004](https://doi.org/10.1016/j.crci.2009.11.004).

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