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Fe₃O₄@SiO₂/collagen: An efficient magnetic nanocatalyst for the synthesis of benzimidazole and benzothiazole derivatives



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ABSTRACT

In this project, Fe₃O₄@SiO₂ was synthesized and combined with collagen for the preparation of Fe₃O₄@SiO₂/collagen. It was characterized by FT-IR, ¹H NMR, VSM, XRD, EDX, SEM and TEM. This nanocatalyst has some interesting advantages such as facile synthetic procedure, high catalytic activity, easy separation and acceptable reusability. It was applied as an efficient nanocatalyst in the synthesis of benzimidazole and benzothiazole derivatives. This method offers several advantages including high yields, short reaction times, easy workup process and environmentally benign reaction conditions.

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1. Introduction

Benzimidazole and benzothiazole derivatives possess a variety of biotic activities. These heterocyclic compounds have shown different pharmacological activities such as antibacterial, antiulcer, antihypertensive, antiviral, antifungal, anticancer, and antihistamine activities [2–7].

Benzimidazole derivatives exhibit significant activity against several viruses such as HIV [3,8], influenza and human cytomegalovirus (HCMV) [8]; they also act as topoisomerase inhibitors [3], selective neuropeptide YY1 receptor antagonists [11], angiotensin II inhibitors [13], and smooth muscle cell proliferation inhibitors [14] and have much more importance in organic synthesis.

Collagen is the most common protein family in the body of living creatures [17]. Collagen has an important role in the formation of tissues and organs and is involved in various functional expressions of cells. Skin, bone, tendon, teeth and blood vessels are some of the living organs where collagen can be found. The main sources of commercial collagen are limited to those of land-based creatures, such as bovine or porcine skins and bones [19].

Collagen has usefulness in a wide variety of biomedical biological applications. As the primary reason, collagen has superior biocompatibility compared with other natural polymers, such as albumin and gelatine. In addition, it can form fibres with extra strength and stability. These features are derived from its enriched functional groups, cross-linkage of its fibres and its supramolecular structure [21].

Until now, a number of methods have been developed for the synthesis of benzimidazoles [22–25]. Generally, 2-substituted benzimidazoles are synthesized using 1,2-phenylendiamine with aldehydes and/or acyl chlorides, carboxylic acids, and orthoesters. However, some synthesis conditions comprise the use microwave irradiation in an acidic medium [26] and/or refluxing in the presence of homo/heterogeneous catalysts such as acetic acid [27], silica supported sulfuric acid [28], Zinc-proline [29], Yb(OTf)₃ [30], etc.

In addition, a lot of methods have been reported for the synthesis of benzothiazoles by condensation of benzene,

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Full paper/Mémoire

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2-aminothiophenol with acyl chlorides or aldehydes. In this case, synthesis procedures include the use of ionic liquid [31] and/or microwave irradiation with a SiO₂ catalyst [32] and/or refluxing in the presence of homo/heterogeneous catalysts such as acetic acid [33], active carbon in toxic solvents [34] and Chitosan-supported Fe₃O₄ [35].

To the best of our knowledge, most of the methods for the synthesis of benzimidazoles and benzothiazoles suffer from one or some disadvantages such as low yields, harsh reaction conditions, time consuming process, use of expensive catalysts and tedious workups. Thus presently, the development of environmentally benign, high-yielding and fast synthesis of benzimidazole and benzothiazole derivatives remains a desired goal in organic synthesis. In this work, we report a highly efficient procedure for the preparation of benzimidazole and benzothiazole derivatives in ethanol (EtOH) media using $Fe_3O_4@SiO_2/$ collagen as an efficient magnetic reusable nanocatalyst.

2. Experimental

2.1. Materials and methods

Hydrolysed collagen was of industrial grade (Parvar Novin-e Tehran Co., Mw ¼ 2000–20,000 Da), which is available in the market, and it has approximately 20% insoluble inorganic salts [36,37]. All chemicals were purchased from Merck, Fluka and Sigma–Aldrich companies and were used without further purification.

All reactions and the purity of benzimidazole and benzothiazole derivatives were monitored by thin-layer chromatography (TLC) using aluminium plates coated with silica gel F254 plates (Merck) using ethyl acetate, *n*-hexane and methanol as eluents. Melting points were determined in open capillaries using an Electrothermal 9100 instrument.

Fourier transform infrared (FT-IR) spectra were recorded on a Shimadzu FT-IR 8400s using KBr plates of samples. Proton nuclear magnetic resonance (¹H NMR) spectra were recorded on a Bruker 400 ultrashield and DMSO-d₆ was used as the solvent. A transmission electron microscope (TEM) from day-petronic company of Iran was used. Scanning electron microscopy (SEM) and energy dispersive Xray (EDX) analysis were performed on a VEGA II TESCAN using 30 KV in high vacuum and Au spin coating for SEM sample preparation. Wide-angle powder X-ray diffraction (XRD) patterns of the solids were obtained in a JEOL with a Cu K α (λ = 0.15420 nm) X-ray irradiation source in a 2 θ range between 5° and 80° . Magnetic properties were recorded by the vibrational sampling magnetometry (VSM) technique in 1.5 T external magnetic fields at room temperature, by using a MDK-6 instrument.

2.2. Preparation of Fe₃O₄ magnetic nanoparticles

Fe₃O₄ nanoparticles were synthesised via a coprecipitation method by using ferric chloride (FeCl₃·6H₂O) and ferrous chloride (FeCl₂·4H₂O) that was introduced in our previous paper [38]. Briefly, Ferric chloride and ferrous chloride dissolved in degassed water and ammonia solution were added to this mixture under vigorous stirring. When pH increases, black colloids of Fe₃O₄ nanoparticles were formed. Fe_3O_4 nanoparticles were separated by using an external magnet in isoelectric point (pH ~ 8). Then, the collected precipitate was poured in EtOH 50% and separated by using a magnet (rinsed using magnet separation) three times. Finally, Fe_3O_4 nanoparticles were dried in a vacuum oven.

2.3. Preparation of the Fe₃O₄@SiO₂/collagen nanocatalyst

First, 2 g of Fe₃O₄ nanoparticles and 5 mL of tetraethyl orthosilicate (TEOS) were added in a 500-mL round bottom flask that contains 150 mL EtOH 50%. In the second vessel, 10 g of collagen was dissolved in 100 mL of a dilute acetic acid solution (5%) under vigorous stirring. A homogeneous supernatant of the second mixture was added to the Fe₃O₄ mixture. The flask was poured in an ultrasonic bath for 30 min. Then, NH₄OH (10%) was added dropwise to the mixture until the pH reached about eight. In the next step, the residue was collected by using an external magnet; it was poured in a Teflon autoclave that contains EtOH (50%) and heated at 90 °C for 6 h. The resulting product was separated by external magnetic fields, dispersed in EtOH 50% and rinsed three times. Finally, it was dried in a vacuum oven at 60 °C for 12 h.

2.4. Synthesis of the non-magnetic nanocatalyst (SiO₂/ collagen)

The nonmagnetic nanocatalyst (SiO₂/collagen) was synthesised by the same procedure (mentioned in 2.3.) without using Fe_3O_4 and tripling the amount of TEOS.

2.5. Synthesis of the non-silicate magnetic nanocatalyst (Fe₃O₄@collagen)

The non-silicate magnetic nanocatalyst ($Fe_3O_4@$ collagen) was synthesised by the same procedure (mentioned in 2.3) without using TEOS in the synthesis procedure.

Table 1

0	ptimization	of the catal	vsts and	solvents f	or synt	hesis of b	enzimidazole.

Entry	Type & amount of catalyst ^b	Solvent	Time (min)	Yield (%)
1	Non-additive	EtOH	1080	25
2	50 (Collagen)	H ₂ O ^c	_	_
3	50 (Collagen)	EtOH ^d	50	85
4	50 (Fe ₃ O ₄)	EtOH	240	30
5	50 (Fe ₃ O ₄ @collagen)	EtOH	20	96
6	50 (SiO ₂ @collagen)	EtOH	25	95
7	50 (Fe ₃ O ₄ @SiO ₂)	EtOH	150	53
8	10 (Fe ₃ O ₄ @SiO ₂ /collagen)	EtOH	20	68
9	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	EtOH	20	97
10	80 (Fe ₃ O ₄ @SiO ₂ /collagen)	EtOH	20	56
11	160 (Fe ₃ O ₄ @SiO ₂ /collagen)	EtOH	20	30
12	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	EtOH	15	97
13	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	H ₂ O	15	Trace
14	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	CH_2Cl_2	15	80
15	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	CHCl ₃	15	49
16	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	CH ₃ CN	15	_
17	50 (Fe ₃ O ₄ @SiO ₂ /collagen)	Solvent free	15	_

^a Benzimidazoles synthesis conditions: 1 mmol 1,2 phenylendiamine, 1 mmol 3-nitrobenzaldehyde, 4 mL solvent at rt.

^b Catalyst (mg).

Table 2
Synthesis of benzimidazole derivatives in the presence of Fe ₃ O ₄ @SiO ₂ /collagen. ^a

MP (°C)		Yield (%)	Time (min)	Product	Di-amine	Aldehyde	Entry
Found	Reported						
315–317 [39]	314-318	78	12		OPD	4-NO ₂ -C ₆ H ₄ CHO	1
185–187 [40]	186-188	97	15		OPD	3-NO ₂ -C ₆ H ₄ CHO	2
261–263 [41]	260	78	15	$\stackrel{\text{H}}{\longrightarrow}$ $\stackrel{\text{O}_2\text{N}}{\longrightarrow}$	OPD	2-NO ₂ -C ₆ H ₄ CHO	3
297–298 [42]	296.5–298	85	15	$ \begin{array}{c} H & \mathbf{1c} \\ \\ \swarrow & \\ H \\ H \\ H \\ H \\ H \\ \mathbf{1d} \end{array} $	OPD	4-Br—C ₆ H ₄ CHO	4
262 [43]	261	90	20		OPD	4-CN-C ₆ H ₄ CHO	5
281–283 [40]	281-282	60	20		OPD	4-Cl—C ₆ H ₄ CHO	6
227 [44]	225	50	20		OPD	2-Cl-C ₆ H ₄ CHO	7
220 [45]	218–220	87	18	$ \begin{array}{c} & & & \mathbf{r} \mathbf{g} \\ & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & & & \mathbf{r} \mathbf{g} \\ & & & & & & & & & & & & \\ & & & & & $	OPD	2,4-Cl ₂ C ₆ H ₃ CHO	8
220 [46]	218–222	73	25		OPD	2,6-Cl ₂ C ₆ H ₃ CHO	9
180–182 [18]	181	55	30		OPD	OCH ₃ -C ₆ H ₄ CHO	10
456–458 [47]	>400	83	20	$ \begin{array}{c} \vdots \\ H \\ \downarrow \\ \downarrow$	OPD	CHO–C ₆ H ₄ CHO	11
342-343 [48]	345	97	25		OPD	2-Thenaldehyde	12
234–235 [49]	234–238	65	60	$\bigcup_{N}^{H} \bigcup_{n}^{U} 1m$	OPD	Furfural	13
183–185 [42]	185	70	30	O_2N N CN N In	m-COOH–(OPD)	4-CN-C ₆ H ₄ CHO	14
191–192 [42]	189	73	25	O_2N N CN N O_2N O	m-NO ₂ -(OPD)	4-CN-C ₆ H ₄ CHO	15

^a Reaction conditions: **1** mmol 1,2 phenylendiamine(OPD), **1** mmol aldehyde, catalytic Fe₃O₄@SiO₂/collagen (50 mg), 4 mL solvent at rt.

2.6. Synthesis of the collagen-less magnetic nanocatalyst $(Fe_3O_4@SiO_2)$

The collagen-less magnetic nanocatalyst (Fe₃O₄@collagen) was synthesised by the same procedure (mentioned in 2.3) without the addition of the second mixture (collagen) in the synthesis procedure.

2.7. General procedure for the synthesis of benzimidazole derivatives

Aromatic aldehyde (1.0 mmol) and 1,2-phenylendiamine (1.0 mmol) were mixed with a catalytic amount of Fe₃O₄@SiO₂/collagen (50 mg) in 4 mL of EtOH and stirred for indicated times (Table 2) at room temperature. Thin-layer chromatography (TLC) was used to monitor the progress of the reaction. After completion of the reaction, the catalyst was separated by external magnetic fields and reused four times in other fresh reactions without a considerable loss of activity. Then the products were crystallized using EtOH 96%. The products were characterized by melting point, FT-IR and ¹H NMR. After each run, the catalyst was separated by using a magnet, rinsed with EtOH and dried. Then, it was used in the next reaction without impressive loss in catalytic reactivity.

Table 3 Synthesis of benzothiazole derivatives the presence of Fe₃O₄@SiO₂/collagen.^a O ADT

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2.8. General procedure for the synthesis of benzothiazole derivatives

Aromatic aldehyde (1.0 mmol) and 2-amino benzothiazole (1.0 mmol) were mixed with a catalytic amount of Fe₃O₄@SiO₂/collagen (50 mg) in 4 mL of EtOH and stirred for indicated times (Table 3) under refluxing conditions. The progress of the reaction was monitored by TLC. After completion of the reaction, the catalyst was separated by external magnetic fields, and reused four times in other fresh reactions without a considerable loss of activity. Then the products were crystallized using EtOH 96%. The products were characterized by melting point, FT-IR and ¹H NMR. After each run, the catalyst was separated by using a magnet, rinsed with EtOH and dried. Then, it was used in the next reaction without impressive loss in catalytic reactivity.

2.9. Selected spectral data

2.9.1. 4-(1H-Benzo[d]imidazol-2-yl) benzonitrile (Table 2, entry 5)

Yellow powder (90%): mp 261 °C. IR (KBr) (v_{max} , cm⁻¹): 3533, 2231, 1658, 1315. ¹H NMR (400 MHz, DMSO- d_6): δ_H (ppm) 13 (s, 1H, NH), 8.3-8.4 (d, 2H, Ar-H), 8 (d, 2H, Ar-H), 7.7 (d, 1H, Ar–H), 7.6 (d, 1H, Ar–H), 7.2 (m, 2H, Ar–H).

V: 11 (0/)

Entry	Aldenyde	Z-AB1	Product	mie (min)	rield (%)	MP (°C)	
						Found	Reported
1	4-NO ₂ -C ₆ H ₄ CHO	2-ABT	$N \rightarrow NO_2$	90	70	223–226	224–225 [1]
2	3-NO ₂ -C ₆ H ₄ CHO	2-ABT		60	83	180–182	180–181 [9]
3	4-CN-C ₆ H ₄ CHO	2-ABT		35	78	161-162	161–162 [10]
4	4-Br—C ₆ H ₄ CHO	2-ABT	\mathbb{A}^{N} \mathbb{A}^{Pr} \mathbb{A}^{Pr}	50	75	120.9–122	120–121 [12]
5	4-Cl ₂ -C ₆ H ₄ CHO	2-ABT		60	73	116	115–117 [10]
6	2,4-Cl ₂ —C ₆ H ₃ CHO	2-ABT	$rac{1}{1}$	65	80	140–143	144–145 [15]
7	2,6-Cl ₂ —C ₆ H ₃ CHO	2-ABT		100	65	89–91	87–90 [16]
8	C ₆ H ₄ CHO	2-ABT		100	70	112-114	110–112 [18]
9	CHO-C ₆ H ₄ CHO	2-ABT		140	63	198	196 [20]

^a Reaction conditions: Aromatic aldehyde (1.0 mmol), 2-Aminobenzothiazol (2-ABT) (1.0 mmol), catalytic amount of Fe₃O₄@SiO₂/collagen (50 mg) and 4 mL of EtOH at reflux.

2.9.2. 2-Phenylbenzothiazole (Table 3, entry 8)

Mp: 112–114 °C. IR (KBr): (v_{max} , cm⁻¹) 3064, 1588, 1555, 1509, 1478, 1433, 1244, 962, 766. ¹H NMR (400 MHz, DMSO-*d*₆): $\delta_{\rm H}$ (ppm) 7.41–8.08 (m, 9H, Ar–H).

3. Results and discussions

3.1. Characterization of Fe₃O₄@SiO₂/collagen

The EDX spectrum of $Fe_3O_4@SiO_2/collagen$ as an exact elemental analysis is depicted in Fig. 1a. As could be seen, existence of Fe, Si, O, C and some of the other atoms in the structure of $Fe_3O_4@SiO_2/collagen$ is proved. In addition, it shows that the molar ratios of Fe:Si:C are 23:21:6. The FT-IR spectrum of the as-synthesized $Fe_3O_4@SiO_2/collagen$ in comparison to that of raw Fe_3O_4 , $Fe_3O_4@collagen$ and $SiO_2/$ collagen is shown in Fig. 1b and these spectra can be compared to the purified collagen spectrum [35,36]. Referring to FT-IR spectra, the presence of the strong Fe–O and Si–O stretching bonds is confirmed. These bonds appear at around 530 and 1050 cm⁻¹, respectively. Moreover, strong and wide peaks appear at approximately 3300 and 3500 cm⁻¹ and a weak peak at around 2900 cm⁻¹ in FT-IR spectra of the catalyst. These wave numbers prove the existence of N–H, O–H and C–H aromatics. Regarding further hydrogen interaction of the end tails of collagen fibres with O–H and N–H moieties, their appeared peaks are broader and stronger than weak peaks of C–H bonds.

XRD patterns were used to determine the crystalline structure of nanocatalysts. The XRD pattern of the asprepared nanocatalyst in comparison to that of pure Fe₃O₄ nanoparticles is shown in Fig. 1c. As can be seen in the XRD patterns, the position and relative intensities of appeared peaks in the catalyst are similar to those of bare Fe₃O₄ nanoparticles. This result confirms the presence of the Fe₃O₄ crystalline structure in the final product.

Fig. 2a and b show the internal and surface morphology of the as-synthesised catalyst that was studied by TEM and SEM images. The TEM image proves the homogeneous spherical core@-shell morphology of Fe₃O₄@SiO₂/collagen.



Fig. 1. a) EDX, b) FT-IR and c) XRD patterns of Fe₃O₄@SiO₂/collagen.



Fig. 2. a) TEM (a1 and a2), b) and SEM images of Fe3O4@SiO2/collagen. c) Diagram of distribution particle size by SEM (red) and TEM (blue) images.

In addition, the distribution diagram of the nanocatalyst diameter is shown in Fig. 2c. These diagrams were obtained by measuring 150 separated particles and show that the average particle size is 39 nm, while about 90% of particles are in the size range of 26–53 nm. Also, this evaluation shows that Fe_3O_4 cores were about 16–25 nm. According to these results, approximately 9–14 nm homogeneous shell covers the surface of the Fe_3O_4 internal core.

One of the important advantages of Fe_3O_4 nanoparticles is superparamagnetic properties which can be separated by external magnetic fields. These unique magnetic properties are due to the configuration of valance electrons in subatomic orbital clouds and arrangement of unique crystalline domains in Fe_3O_4 nanoparticles. Magnetic properties of the as-synthesised Fe_3O_4 were evaluated by VSM analysis. In VSM analysis, high magnetization saturation and low magnetic fields were coercively ascribed to superparamagnetic behaviour of samples. As shown in Fig. 3, magnetization saturation of Fe_3O_4 nanoparticles, Fe_3O_4 @collagen and Fe_3O_4 @SiO₂/collagen reduced respectively. Probably, these reductions were affected by establishment of deterrent shells on the surface of Fe_3O_4 nanoparticles.

These results confirmed that Fe₃O₄@SiO₂/collagen with a core@shell structure and high superparamagnetic properties has been successfully synthesized.



Fig. 3. VSM analysis of 1) Fe_3O_4, 2) Fe_3O_4@collagen and 3) Fe_3O_4@SiO_2/ collagen

3.2. Catalytic application of $Fe_3O_4@SiO_2/collagen$ in organic synthesis

To investigate the catalytic activity, the synthesized $Fe_3O_4@SiO_2/collagen$ was used for the preparation of benzimidazole and benzothiazole compounds as model reactions. Initially, $Fe_3O_4@SiO_2/collagen$ as the nanocatalyst was used for the synthesis of benzimidazole and its



Scheme 1. General procedure for the synthesis of benzimidazole (X:N) and benzothiazole (X:S).

derivatives in the presence of 1,2-phenylendiamine and 3nitrobenzaldehyde (1:1 molar ratio) (Scheme 1).

Table 1 shows optimization conditions with different solvents and catalysts of this reaction. Some suggested solvents such as EtOH, CH_2Cl_2 , H_2O in other literature studies were tested in the model synthesis. The results revealed that EtOH is the best solvent for the reaction. Consequently, the effect of the catalyst on the time and yield of reaction was optimized and the best result was obtained with 50 mg/mmol of $Fe_3O_4@SiO_2/collagen$. In the next step, structural properties of $Fe_3O_4@SiO_2/collagen$ were compared with those of similar nanostructures (Scheme 2).

Collagen and hydrolysed collagen have high intumescent affinity and get swollen when they were used in aprotic and aqueous media [36,37]. Then, they cannot be used directly as catalysts and should be loaded and/or stabilized on a nanostructure substrate.

Collagen inflates in H_2O and the reaction cannot be surveyed. So, it was loaded on the surface of Fe_3O_4 .

In the surveyed synthesis methods, Fe₃O₄@collagen has shown efficient catalytic and magnetic properties. In this case, collagen was loaded on the surface of Fe₃O₄ nanoparticles but it cannot be properly stabilized on them. Thus, some of the unconsolidated collagen was removed, regardless of the loaded collagen amount on the catalyst, after each run, collagen content was reduced and reaction vields decreased. Therein, it does not have good reusability in comparison to Fe₃O₄@SiO₂/collagen (see Fig. 4a). SiO₂/ collagen is a stable nanocatalyst and shows the same reactivity in synthesis method, but it does not have convenient magnetic properties for use as a magnetic recoverable nanocatalyst. In the next experiment, the catalytic reactivity of Fe₃O₄ and Fe₃O₄/SiO₂ was compared with that of Fe₃O₄@SiO₂/collagen as the main catalyst. The results show that collagen-less magnetic nanocatalysts do not have remarkable catalytic reactivity in comparison to collagen containing catalysts. Fe₃O₄@SiO₂/collagen has both stability and superparamagnetic properties that facilitate magnetic recovery after each run. To expand this



Scheme 2. Type of catalyst that was tested in organic synthesis.



Fig. 4. a) Nanocatalyst reactivity for the synthesis of entry 2 in Table 2; 1) $Fe_3O_4@Collagen and 2) Fe_3O_4@SiO_2/Collagen;$ **b** $) FT-IR spectra of 1) asprepared <math>Fe_3O_4@SiO_2/Collagen and 2)$ after four catalytic runs.

method Fe₃O₄@SiO₂/collagen was used in the synthesis of benzimidazole and benzothiazole derivatives and the results are summarized in Tables 2 and 3, respectively.

One of the important advantages of the Fe₃O₄@SiO₂/ collagen catalyst is its easy and reliable separation from the reaction mixture. Then, the reusability of this nanocatalyst was investigated in the benzimidazole model reaction. The heterogeneous nature and magnetic properties of the catalyst allowed its facile recovery via a simple separation process by an external magnet. The collected catalyst was rinsed with EtOH and dried at room temperature for reuse in other reactions. The separated nanocatalyst was reused in the mentioned reaction for the synthesis of entry 2 in Table 2 four times without considerable loss of its catalytic reactivity. The catalytic reactivity of reused Fe₃O₄@SiO₂/ collagen in comparison to that of Fe₃O₄@collagen is depicted in Fig. 4a. In addition, FT-IR spectra of the recovered catalyst after four runs in comparison with those of the as-synthesized Fe₃O₄@SiO₂/collagen are depicted in Fig. 4b. As could be seen, there are no obvious differences in the FT-IR spectra of the as-prepared and recovered nanocatalyst. These appearances confirm the stability of the nanocatalyst under the reaction conditions.

Due to the mentioned benefits and as could be compared by the literature, $Fe_3O_4@SiO_2/collagen$ is one of the best heterogeneous nanocatalysts for the synthesis of benzimidazole and benzothiazole derivatives.

In order to indicate the efficiency of our method it was compared with that of some other reported methods and the results are summarized in Table 4. The results clearly show the high efficiency of Fe₃O₄@SiO₂/collagen in the synthesis of benzimidazole. Also, Table 5, shows the efficiency of our synthesized nanocatalyst in the preparation of benzothiazole compared to that of other reported methods.

The suggested mechanism for the synthesis of benzimidazole and benzothiazole derivatives is proposed in Scheme 3. The surface of collagen in Fe₃O₄@SiO₂/collagen

Table 4

A comparison of the efficiency of Fe $_3O_4@SiO_2/collagen$ with that of other catalysts in the synthesis of benzimidazole. a

Year	Catalyst	Solvent	Temperature (°C)	Time (min)	Yield (%)
2006	Zeolite	Dioxane	100-102	300	69 [50]
2010	Scolecite	EtOH	70	45	91 [41]
2013	Nano-ZnO	EtOH	Reflux	95	68 [18]
2013	Nano-alumina	EtOH	Reflux	90	74 [18]
This	Fe ₃ O ₄ @SiO ₂ /	EtOH	rt	12	78
Work	collagen				

^a All data compared for entry 2 in Table 2.

Table 5

A comparison of the efficiency of Fe $_3O_4@SiO_2/collagen$ with that of other catalysts in the synthesis of benzothiazole.^a

Year	Catalyst	Solvent	Temperature (°C)	Time (min)	Yield (%)
2006 2013 2014	CH ₃ COOH Nano-ZnO FeCl ₃ ·H ₂ O	CH ₃ COOH EtOH DMSO/H ₂ O	Reflux Reflux 100 (°C)	300 80 800	65 [33] 72 [18] 41 [22]
This Work	K ₂ S ₂ U ₈ Fe ₃ O ₄ @SiO ₂ / collagen	EtOH	Reflux	50	75

^a All data compared for entry 3 in Table 3.

collagen is a new nanocatalyst with some interesting advantages such as high magnetic properties, good selectivity (Scheme 4), easy separation and high yield in the synthesis of heterocyclic compounds.

4. Conclusions

In this work, after synthesis of Fe₃O₄@SiO₂, it was combined with collagen for the preparation of Fe₃O₄@-SiO₂/collagen and characterization was done by various instrumental methods. Remarkable advantages of this nanocatalyst are the facile synthesis procedure, high catalytic activity, easy separation and acceptable reusability. This nanocomposite as an efficient nanocatalyst was used for the synthesis of benzimidazole and benzothiazole derivatives due to its excellent properties and high selectivity. Also, this procedure has some benefits such as the use of harmless solvents, facile conditions, easy isolation of the products with excellent purity, increasing reaction yields, reducing reaction time and waste-less chemical process.



Scheme 3. The proposed mechanism for the formation of benzimidazole and benzothiazole derivatives.



Scheme 4. Selectivity of Fe₃O₄@SiO₂/collagen in the synthesis of benzimidazole derivatives.

has a variety of functional groups which can form hydrogen bonds with protic moieties. In addition, collagen fibres with organic chains can interact with aprotic moieties of organic compounds. This quality has the main role in the reaction times and reaction yields in reactions with collagen containing nanocatalysts [51–53].

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References

- C. Praveen, A. Nandakumar, P. Dheenkumar, D. Muralidharan, P.T. Perumal, J. Chem. Sci. 124 (2012) 609–624.
- [2] A.A. Spasov, I.N. Yozhitsa, L.I. Bugaeva, V.A. Anisimova, Pharm. Chem. J. 33 (1999) 232-243.
- [3] J.S. Kim, B. Gatto, C. Yu, A. Liu, L.F. Liu, E.J. LaVoie, J. Med. Chem. 39 (1996) 992–998.
- [4] T. Roth, M.L. Morningstar, P.L. Boyer, S.H. Hughes, R.W. Buckheit Jr., C.J. Michejda, J. Med. Chem. 40 (1997) 4199–4207.
- [5] I. Hutchinson, T.D. Bradshaw, C.S. Matthews, M.F.G. Stevens, A.D. Westwell, Bioorg. Med. Chem. Lett. 13 (2003) 471–474.
- [6] S.T. Huang, I.J. Hsei, C. Chen, Bioorg. Med. Chem. 14 (2006) 6106–6119.
- [7] R.J. Alaimo, S.S. Pelosi, R. Freedman, J. Pharm. Sci. 67 (1978) 281–282.
- [8] A.R. Porcari, R.V. Devivar, L.S. Kucera, J.C. Drach, L.B. Townsend, J. Med. Chem. 41 (1998) 1252–1262.
- [9] K.U. Sadek, R.A. Mekheimer, A.M. Hameed, F. Elnahas, M.H. Elnagdi, Molecules 17 (2012) 6011–6019.
- [10] B. Maleki, H. Salehabadi, M. Khodaverdian Moghaddam, Acta Chim. Slov. 57 (2010) 741–745.
- [11] H. Zarrinmayeh, D.M. Zimmerman, B.E. Cantrell, D.A. Schober, R.E. Bruns, S.L. Gackenheimer, P.L. Ornstein, P.A. Hipskind, T.C. Britton, D.R. Gehlert, Bioorg. Med. Chem. Lett. 9 (1999) 647–652.
- [12] L. Chen, C. Yang, S. Li, J. Qin, Spectrochim. Acta, Part A 68 (2007) 317–322.
- [13] Y. Kohara, K. Kubo, E. Imamiya, T. Wada, Y. Inada, T. Naka, J. Med. Chem. 39 (1996) 5228–5235.
- [14] Elokdah, H. M.; Chai, S. Y.; Sulkowski; T. S. in: U.S. Patent, Google Patents, USA, 1998.
- [15] V.Z. Shirinian, S.Y. Melkova, L.I. Belen'kii, M.M. Krayushkin, Russ. Chem. Bull. 49 (2000) 1859–1862.
- [16] Gurian, S. D.; Schwartz, H. A. in: U.S. Patent, Google Patents, U.S., 1964.
- [17] P. Semal, R. Orban, J. Archaeol. Sci. 22 (1995) 463-467.
- [18] A. Teimouri, A.N. Chermahini, H. Salavati, L. Ghorbanian, J. Mol. Catal. A Chem. 373 (2013) 38–45.
- [19] W. Suphatharaprateep, B. Cheirsilp, A. Jongjareonrak, New Biotechnol. 28 (2011) 649–655.
- [20] S. Samanta, S. Das, P. Biswas, J. Org. Chem. 78 (2013) 11184–11193.
- [21] C.H. Lee, A. Singla, Y. Lee, Int. J. Pharm. 221 (2001) 1–22.
- [22] G. Bai, X. Lan, X. Liu, C. Liu, L. Shi, Q. Chen, G. Chen, Green Chem. 16 (2014) 3160.
- [23] L. Hao, Y. Zhao, B. Yu, H. Zhang, H. Xu, Z. Liu, Green Chem. 16 (2014) 3039.
- [24] H. Sharma, N. Singh, D.O. Jang, Green Chem. 16 (2014) 4922–4930.

- [25] J.-P. Wan, S.-F. Gan, J.-M. Wu, Y. Pan, Green Chem. 11 (2009) 1633.
- [26] A.J. Sandee, D.G. Petra, J.N. Reek, P.C. Kamer, P.W. van Leeuwen, Chemistry 7 (2001) 1202-1208.
- [27] A. Stein, Adv. Mater. 15 (2003) 763-775.
- [28] A. Corma, Chem. Rev. 97 (1997) 2373–2420.
- [29] J.A. Melero, R. van Grieken, G. Morales, Chem. Rev. 106 (2006) 3790–3812.
- [30] Y. Izumi, Catal. Today 33 (1997) 371-409.
- [31] B.C. Ranu, R. Jana, S.S. Dey, Chem. Lett. 33 (2004) 274–275.
- [32] M. Kodomari, Y. Tamaru, T. Aoyama, Synth. Commun. 34 (2004) 3029–3036.
- [33] R.S. Kenny, U.C. Mashelkar, J. Heterocycl. Chem. 43 (2006) 1367–1369.
- [34] M. Hayashi, Y. Kawashita, Lett. Org. Chem. 3 (2006) 571–578.
- [35] A. Maleki, N. Ghamari, M. Kamalzare, RSC Adv. 4 (2014) 9416–9423.
- [36] A. Pourjavadi, M. Kurdtabar, Eur. Polym. J. 43 (2007) 877–889.
- [37] A. Pourjavadi, M. Kurdtabar, H. Ghasemzadeh, Polym. J. 40 (2007) 94-103.
- [38] H. Ghafuri, A. Rashidizadeh, B. Ghorbani, M. Talebi, New J. Chem. 39 (2015) 4821–4829.
- [39] M. Kalhor, A. Mobinikhaledi, J. Jamshidi, Res. Chem. Intermed. 39 (2012) 3127–3133.
- [40] H. Eshghi, M. Rahimizadeh, A. Shiri, P. Sedaghat, Bull. Korean Chem. Soc. 33 (2012) 515–518.
- [41] L.S. Gadekar, B.R. Arbad, M.K. Lande, Chin. Chem. Lett. 21 (2010) 1053–1056.
- [42] G.H. Mahdavinia, S. Rostamizadeh, A.M. Amani, H. Sepehrian, Heterocycl. Commun. 18 (2012) 33–37.
- [43] H. Sharghi, O. Asemani, S.M.H. Tabaei, J. Heterocycl. Chem. 45 (2008) 1293–1298.
- [44] G. Brasche, S.L. Buchwald, Angew. Chemie 47 (2008) 1932–1934.
- [45] M.A. Chari, D. Shobha, E.-R. Kenawy, S.S. Al-Deyab, B.V.S. Reddy, A. Vinu, Tetrahedron Lett. 51 (2010) 5195–5199.
- [46] S.M. Inamdar, V.K. More, S.K. Mandal, Tetrahedron Lett. 54 (2013) 579–583.
- [47] L. Krbechek, H. Takimoto, J. Org. Chem. 29 (1964) 3630-3632.
- [48] Y. Kim, M.R. Kumar, N. Park, Y. Heo, S. Lee, J. Org. Chem. 76 (2011) 9577–9583.
- [49] C.A. Ramsden, H.L. Rose, J. Chem. Soc., Perkin Trans. 1 (1997) 2319–2328.
- [50] M.M. Heravi, M. Tajbakhsh, A.N. Ahmadi, B. Mohajerani, Monatsh. Chem. 137 (2005) 175–179.
- [51] M. Talebi, H. Ghafuri, Ind. Eng. Chem. Res. 55 (2016) 2970–2982.
- [52] Y.-S. Lee, Y.-H. Cho, S. Lee, J.-K. Bin, J. Yang, G. Chae, C.-H. Cheon, Tetrahedron 71 (2015) 532–538.
- [53] K. Bahrami, M.M. Khodaei, F. Naali, J. Org. Chem. 73 (2008) 6835–6837.