



INSTITUT DE FRANCE
Académie des sciences

Comptes Rendus

Chimie

Dominique Bazin, Ivan T. Lucas, Stéphan Rouzière, Erik Elkaim, Cristian Mocuta, Solenn Réguer, David G. Reid, Jérémie Mathurin, Alexandre Dazzi, Ariane Deniset-Besseau, Margaux Petay, Vincent Frochot, Jean-Philippe Haymann, Emmanuel Letavernier, Marie-Christine Verpont, Eddy Foy, Elise Boudierlique, Hester Colboc and Michel Daudon

Profile of an “at cutting edge” pathology laboratory for pathological human deposits: from nanometer to *in vivo* scale analysis on large scale facilities


Volume 25, Special Issue S1 (2022), p. 219-233

Published online: 27 July 2022

<https://doi.org/10.5802/crchim.199>

Part of Special Issue: Microcrystalline pathologies: Clinical issues and nanochemistry

Guest editors: Dominique Bazin (Université Paris-Saclay, CNRS, ICP, France), Michel Daudon, Vincent Frochot, Emmanuel Letavernier and Jean-Philippe Haymann (Sorbonne Université, INSERM, AP-HP, Hôpital Tenon, France)

 This article is licensed under the
CREATIVE COMMONS ATTRIBUTION 4.0 INTERNATIONAL LICENSE.
<http://creativecommons.org/licenses/by/4.0/>



Les Comptes Rendus. Chimie sont membres du
Centre Mersenne pour l'édition scientifique ouverte
www.centre-mersenne.org
e-ISSN : 1878-1543



Microcrystalline pathologies: Clinical issues and nanochemistry / *Pathologies microcristallines : questions cliniques et nanochimie*

Profile of an “at cutting edge” pathology laboratory for pathological human deposits: from nanometer to *in vivo* scale analysis on large scale facilities

Dominique Bazin^{*, a}, Ivan T. Lucas^b, Stéphan Rouzière^c, Erik Elkaim^d, Cristian Mocuta^d, Solenn Réguer^d, David G. Reid^e, Jérémie Mathurin^a, Alexandre Dazzi^a, Ariane Deniset-Besseau^a, Margaux Petay^a, Vincent Frochot^{f, g, h}, Jean-Philippe Haymann^{f, g, h}, Emmanuel Letavernier^{f, g, h}, Marie-Christine Verpont^{f, g, i}, Eddy Foy^j, Elise Boudierlique^{f, g, h}, Hester Colboc^k and Michel Daudon^{f, g, h}

^a Université Paris-Saclay, CNRS, Institut de Chimie Physique, 91405 Orsay cedex, France

^b Laboratoire LISE UMR 8235 CNRS-SU, Sorbonne Université, Paris, France

^c Université Paris-Saclay, CNRS, Laboratoire de Physique des Solides, 91405 Orsay Cedex, France

^d Synchrotron SOLEIL, L'Orme des Merisiers Saint Aubin BP48, 91192 Gif-sur-Yvette Cedex, France

^e Yusuf Hamied Department of Chemistry, University of Cambridge, Lensfield Road, Cambridge CB2 1EW, UK

^f Sorbonne Université, UPMC Univ Paris 6, UMR S 1155, Paris, France

^g INSERM, UMR S 1155, Hôpital Tenon, Paris, France

^h Explorations fonctionnelles multidisciplinaires, AP-HP Hôpital Tenon, Paris, France

ⁱ LUMIC network, F-75020 Paris, France

^j Université Paris-Saclay, CEA, CNRS, NIMBE-LAPA, 91191 Gif-sur-Yvette, France

^k Sorbonne Université, AP-HP, Hôpital Rothschild, Service Plaies et Cicatrisation, Paris, France

E-mails: dominique.bazin@universite-paris-saclay.fr (D. Bazin), ivan.lucas@sorbonne-universite.fr (I. T. Lucas), stephan.rouziere@u-psud.fr (S. Rouzière), erik.elkaim@synchrotron-soleil.fr (E. Elkaim), cristian.mocuta@synchrotron-soleil.fr (C. Mocuta), solenn.reguer@synchrotron-soleil.fr (S. Réguer), dgr30@cam.ac.uk (D. G. Reid), jeremie.mathurin@u-psud.fr (J. Mathurin), alexandre.dazzi@u-psud.fr (A. Dazzi), ariane.deniset@u-psud.fr (A. Deniset-Besseau), margaux.petay@u-psud.fr (M. Petay), vincent.frochot@tnn.aphp.fr (V. Frochot), jean-philippe.haymann@tnn.aphp.fr

* Corresponding author.

(J.-P. Haymann), emmanuel.letavernier@tnn.aphp.fr (E. Letavernier), marie-christine.verpont@upmc.fr (M.-C. Verpont), eddy.foy@cea.fr (E. Foy), elise.bouderlique@inserm.fr (E. Bouderlique), hester.colboc@aphp.fr (H. Colboc), daudonmichel24@gmail.com (M. Daudon)

Abstract. This contribution aims to define an analysis procedure for abnormal deposits in human tissues starting from *in vivo* characterization, down to the nanoscale using major instrumentation. Such an integrated approach is based on recent literature, but particularly on our research over the last twenty years on pathological calcifications. To this end, we begin by describing four successive analytical steps, on the injury site or physician's surgery, at the hospital, at a typical physicochemical laboratory, and finally at a large scale (possibly multinational) facility. For the first step, we present various techniques which can be implemented on portable instruments. For the second step, commercial analytical setups are used. In a physicochemical laboratory, prototype or commercial setups are used and finally on large scale instruments, characterization techniques with better spatial resolution and/or higher sensitivity or techniques specific to synchrotron radiation are employed.

Keywords. Pathological calcifications, Characterization techniques, Experimental approach, Medical diagnostic, Abnormal deposits.

Published online: 27 July 2022

1. Introduction

Pathological deposits are very common in the human body and accurate characterization is of major importance for medical care. The research described here was performed in close collaboration with several hospitals [1–4]. Pathological calcifications and abnormal deposits more generally, such as in tattoos [5] or sarcoidosis [6,7], may be intimately linked with major health problems such as cancer [8–13], infection [14–19], diabetes [20–22], or genetic disorders [23–26]. The various reviews published from either a medical [27–31] or a physicochemical point of view [1–4,22–38] have clearly underlined that characterizing deposits, of endogenous and exogenous origins, in the human body are crucial to not only assist diagnosis but also to optimise and develop specific therapy regimens.

More precisely, recent literature and the results we have gathered on different human tissues, including bone [39–41], skin [5,6,42,43], thyroid [34–46], prostate [8,47], kidney [42–51], cartilage [52–55], breast [9,56], salivary glands [57], and pancreas [58], showed that characterization by staining is often of limited utility. A more comprehensive approach has to encompass a wide range of techniques able to describe pathological calcifications at different scales. They can be classified into different families; techniques able to define elemental compositions such as X-ray fluorescence (XRF) [59,60] or energy dispersive X-ray spectroscopies (EDS) [61,62], those which describe molecular composition such as

Fourier transform infrared (FTIR) [63,64] or Raman spectroscopies [65–68] as well as scattering techniques [69–72]. Imaging techniques such as scanning [61,62] and transmission electron microscopy (SEM and TEM) [73–76] or micro computed tomographic imaging [77–80] are also of prime importance. Finally, it is possible to precisely define the electronic state of some elements through X-ray photoelectron spectroscopy (XPS) [81–83] and/or their local atomic level environment using X-ray absorption spectroscopy (XAS) [84–86], electron energy-loss spectroscopy (EELS) [76,87,88] or nuclear magnetic resonance (NMR) [57,89–91].

The micrometer to nanometer scale information about abnormal deposits that these techniques yield allows an accurate description of the processes underlying the pathology, by defining morphology, elemental composition, and molecular and structural aspects. Some of these techniques can be used *in vivo* on abnormal deposits in human tissues while some can only be used on biopsy material. The application of all of these complementary techniques in the specific characterisation of abnormal human deposits is detailed in the present paper.

2. Defining an analytical procedure

We will consider the analysis procedure in four steps (Figure 1). The first is initial characterization directly on the human body using a portable analytical device at the physician's surgery or at the in-

<p>Step I Physician's office Injury location <i>In vivo</i></p>	<p>Commercial Portable instruments : Imaging : Optical Microscope, X ray radiography Elementary & Chemical Composition FTIR and Raman spectroscopies XRF-XRD,</p>	<p>Millimeter scale Micrometer Scale</p>
<p>Step II Hospital <i>In vivo</i> or Biopsy investigations</p>	<p>Commercial analytical set up Imaging : Optical Microscope, X ray radiography, μCT-Tomography , μSEM-EDX and NanoTEM-EDX, 2D-FTIR and Raman spectroscopies μXRF-μXRD,</p>	<p>Millimeter scale Micrometer Scale Nanometer Scale</p>
<p>Step III Physicochemical Laboratory</p>	<p>Prototype experimental set up μXRF-μXRD, NMR Spectroscopy NanoFTIR and NanoRaman spectroscopies SEM-EDX and TEM-EELS NanoIR (AFMIR & OPTIR)</p>	<p>Micrometer Scale Nanometer Scale</p>
<p>Step IV Large scale facility <i>In vivo</i> or Biopsy investigations</p>	<p>Experiments on Beamlines μFTIR and UV spectroscopies nano – XRD micro or nano - XAS (XANES-EXAFS) nanoCT-Tomography</p>	<p>Micrometer scale Nanometer Scale</p>

Figure 1. Various steps of the experimental approach proposed for structural and chemical investigations of pathological calcifications.

jury location depending on severity. The next would be in a hospital in which commercial analytical apparatus is available. The third step involves characterization of biopsies in a physical laboratory performing fundamental analytical research. We con-

clude by describing experiments requiring large scale instrumentation (often shared with other national and international users) such as synchrotron facilities. Such “photon factories” produce well-collimated and high intensity photon syn-

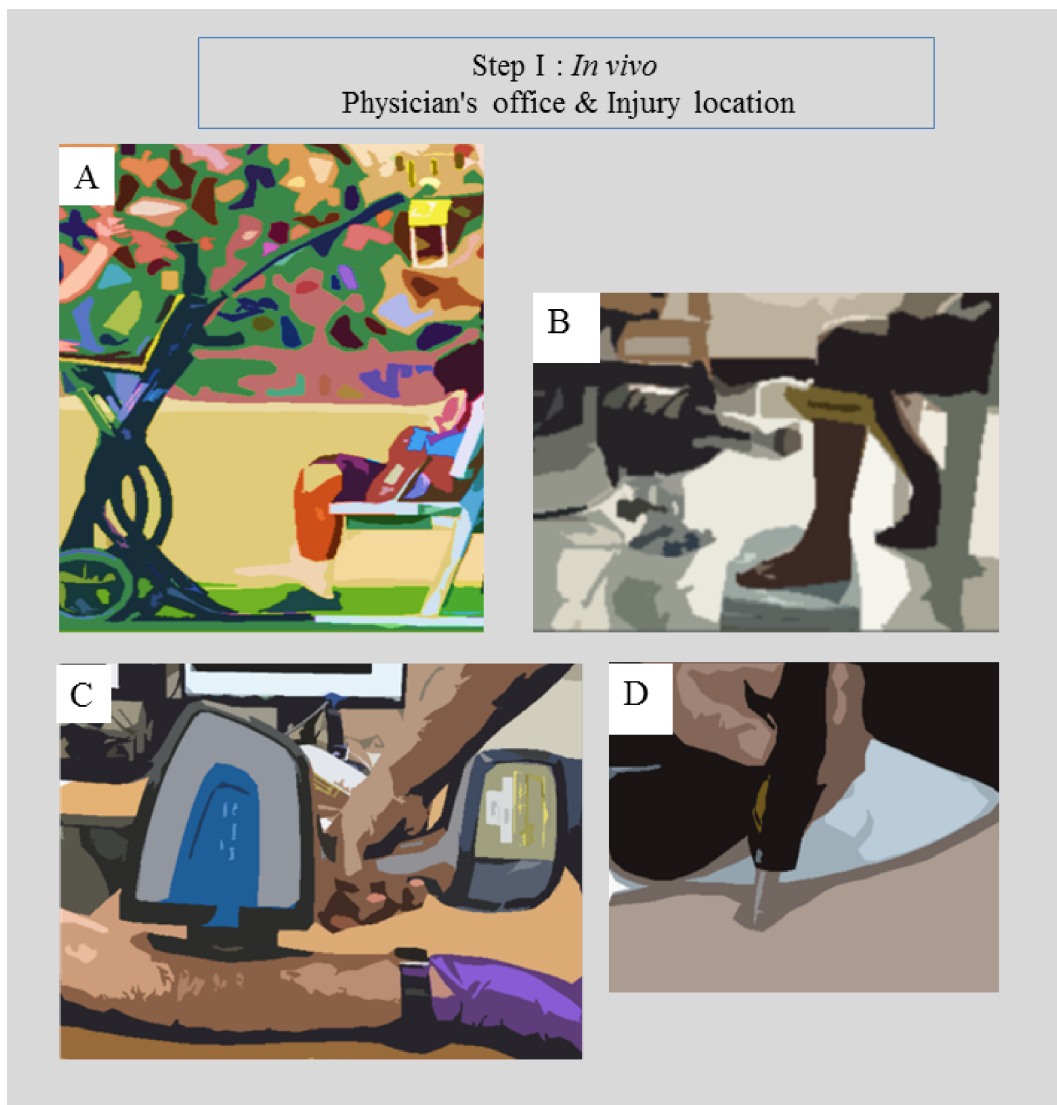


Figure 2. *In vivo* portable devices for (a) Radiography [92], (b) XRF [93], (c) FTIR [94] and (d) Raman [95] spectroscopies.

chrotron radiation beams leading to better spatial resolution and/or higher sensitivity.

2.1. *In vivo* analysis at the physician's surgery or place of injury

The *in vivo* or *in situ* (for forensic medicine) step takes account of the fact that different easy-to-implement experimental setups have been developed over the last decades (Figure 2). These include

portable radiography apparatus [96], which can define an area of interest for extraction of biological samples if possible and necessary for subsequent characterization.

XRF (for elemental analysis and quantification) [97,98], FTIR [99–102] or Raman [103–105] spectroscopies (to determine the nature and amount of chemical compounds present in the sample) constitute at least three characterization techniques which can be applied *in vivo*. It is worth mentioning that biological fluids (or actual tissue in the case of

accident trauma) can also be analysed, in which case, two other techniques namely X-ray Diffraction (XRD, to determine the nature and quantity of any crystalline phases present in the sample) and tabletop SEM (for submicrometer scale surface imaging) can also be applied.

More precisely, XRD can be used to determine the crystallographic structure of unknown deposits [106,107], to identify the different crystalline phases and their composition, concentration, and average crystal dimensions [72,108]. “Tabletop” SEM is also available for observing the micrometer scale surface topology of the sample [109] and ultimately determine particle elemental composition by EDS spectroscopy [72].

2.2. Analysis at the hospital on the patient (biopsy)

The second step in a concerted analysis would be performed in a hospital laboratory where commercial physicochemical setups can be used (Figure 3). Investigations *in vivo* as well as on biopsies are possible. Biopsy procedures are usually performed at the hospital under and/or after radiological examination in the case of breast [110] or kidney [111]. A commercial in-lab setup provides analytical data superior to that produced by portable apparatus (better spatial resolution and sensitivity); the three techniques considered above, namely SEM, XRF and XRD can be performed directly on the biopsy sample (i.e. before paraffin embedding). Note that at Tenon Hospital nanometer scale biopsy characterization by TEM is also possible (Figure 3E).

Embedding the biopsy in a paraffin block allows the sample to be preserved and manipulated simply and safely. It also offers the opportunity to collect a set of data yielding elemental composition (through μ XRF or μ EDX experiments) or chemical mapping (μ FTIR, μ Raman or μ XRD experiments), from the same sample. Note that for μ XRF and μ XRD experiments, it is better to use the paraffin block itself, while for μ FTIR or μ Raman experiments sectioning of the block into several micrometer thick slices should be considered. Microtomography may be necessary to precisely locate the abnormal deposit in the block. The resultant section can be deposited

on different supports for physicochemical characterization, such as conventional glass slides, low-e microscope slides (MirrIR, Kevley Technologies, Tienta Sciences, Indianapolis) or kapton[®] film. It is only possible to collect a Raman spectrum and to observe the sample by SEM using conventional glass slides. Low-e microscope slides enable imaging of surface topology by SEM, or chemical characterization with μ Raman and μ FTIR spectroscopies. Note that other supports have been considered in hospitals in order to perform TEM observations at the nanometer scale.

2.3. Analysis of biological samples in a physics or chemistry laboratory

In a physics or chemistry laboratory (Figures 4 and 5), characterization using prototype experimental set ups can be performed, which may combine optics, sample holders, or detectors, developed in the laboratory. Such experiments are usually not performed in a hospital environment because the instrumentation is expensive, and the techniques are not yet on the mainstream diagnostic analysis path. The sophisticated character of these prototype setups allows significantly improved spatial resolution and/or sensitivity relative to commercial instrumentation.

Special detectors on a TEM microscope (Figure 4B) allow nanometer scale elemental analysis by Nano-EDX as well as chemical analysis by EELS. Also, the experimental set up developed at the Laboratoire de Physiques des Solides combines a multi-layer mirror and a high intensity rotating-anode generator delivering a well-defined beam (100 μ m size) of sufficient intensity. Using a hybrid-pixel detector and a SDD detector, one can measure XRD and XRF signals respectively at the micrometer scale rapidly (time scale between 1 and 10 min depending on the sample) (Figure 4C).

Imaging of the topology of biopsies, deposited on low-e microscope slides and based on SEM observations and chemical characterization with μ Raman and/or μ FTIR spectroscopies, can be complemented by NanoIR [112–122]. NanoIR, for which two experimental configurations are possible, presents novel opportunities. The first configuration is associated with 500 nm spatial resolution and is based on a pump-probe architecture using two laser sources,

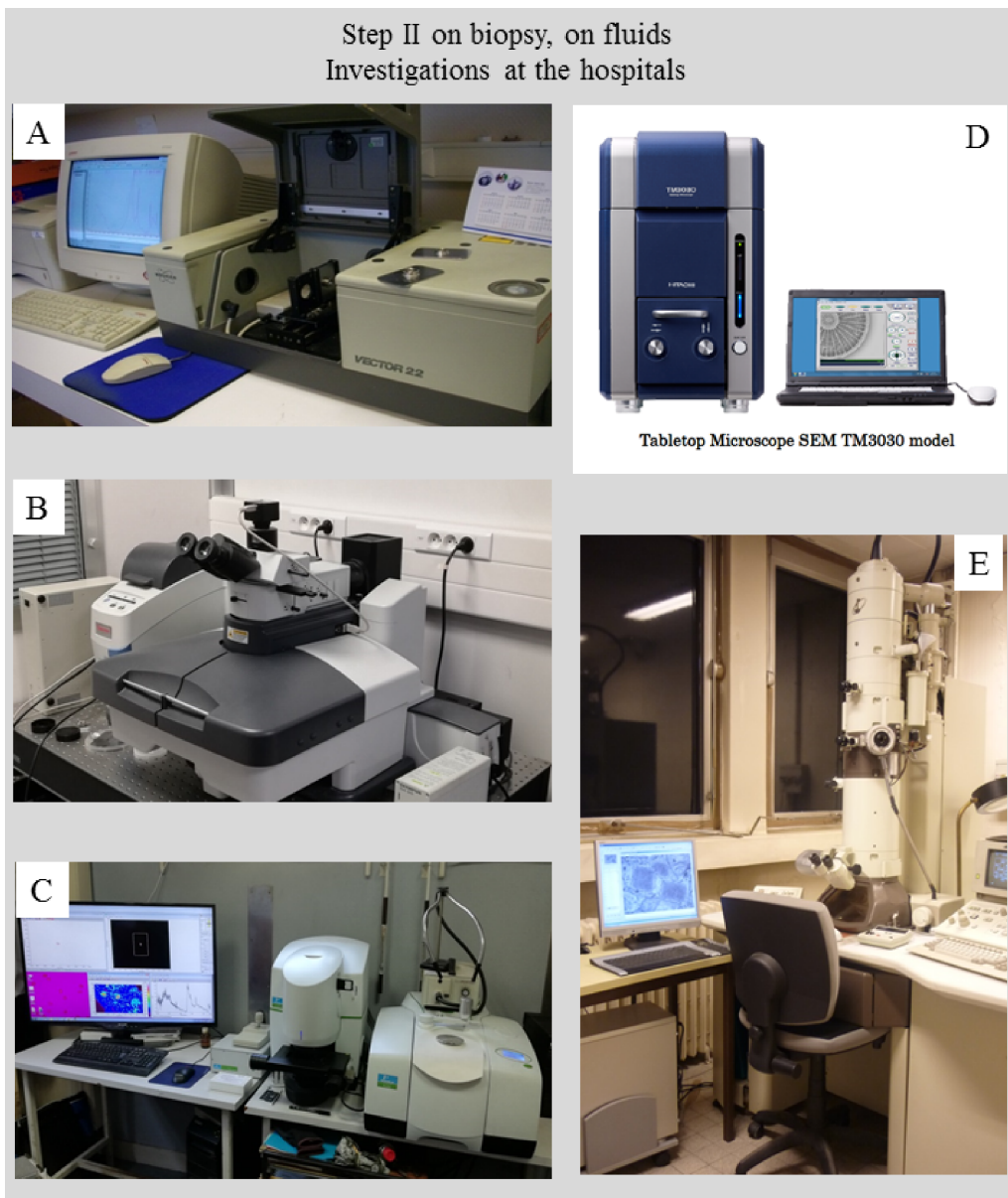


Figure 3. Tabletop commercial instruments (A) Classical FTIR experimental set up, (B) Raman experimental set up (implemented at the synchrotron SOLEIL), (C) μ FTIR microscope implemented at the Service des Explorations Fonctionnelles at the Tenon Hospital, (D) Tabletop SEM (from <https://www.hitachi-hightech>), (E) TEM experimental set up implemented at the Tenon Hospital.

one for mid-infrared excitation (the pump) and the other one for measuring the photothermal effect (the probe) [112]. The second configuration combines an atomic force microscope (AFM) and IR lasers [115, 116] and produces a spatial resolution of 10 nm [122].

2.4. Analysis of biological samples in a large scale facility

The fourth and final step in a comprehensive analysis procedure uses characterization technology

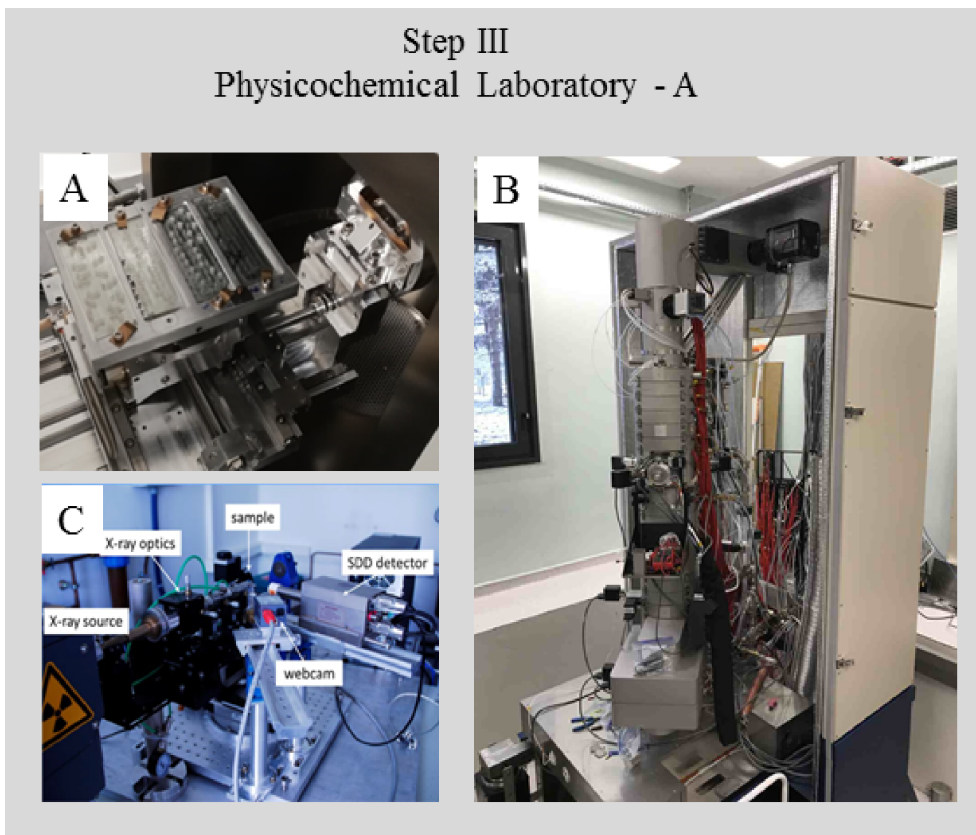


Figure 4. Forefront physics or chemistry laboratory characterization hardware, all located in the Laboratoire de Physique des Solides, Orsay. (A) SEM apparatus sample holder. (B) New transmission electron microscope. (C) An experimental device combining XRD and XRF capabilities.

usually (but not exclusively) specific to large scale instruments, which significantly increases sensitivity and/or in plane spatial resolution.

Synchrotron radiation as a submicrometer probe can be applied in several characterization techniques such as XRF or XRD. Synchrotron radiation is related to the emission of light when charged particles (electrons or positrons) moving with relativistic velocity undergo radial acceleration [123–126]. Among the important advantages of a synchrotron over a laboratory X-ray source are its large spectral range, and its brilliance (a physical characteristic encompassing the photon flux, bandwidth and the angular and lateral spread of the beam) which is more than a million times higher than that of the X-ray tube. Basically, the different beamlines on a synchrotron use a selected (but tunable) part of its energy spectrum, from terahertz to hard X-ray frequencies. This con-

fers the advantage of being able to choose the energy of the incident beam during measurements. Two facilities are implemented in France namely SOLEIL (French national source) [127] and ESRF (European Source) [128].

van der Ent *et al.* [129] have compared different XRF approaches using either X-ray tubes, electron, proton, and synchrotron radiation as probes. While the spatial resolution of an experimental set up using a laboratory source is around 30–100 μm , it is possible to perform similar experiments on synchrotrons with a spatial resolution well below 1 μm , and down to about 50 or even 10 nm [130,131]. Also, there is a significant difference between the limits of detection which is >50 ppm in the case of a laboratory source and 0.1 ppm on synchrotron facilities. These detection limits depend intimately on the sample/matrix studied as well; the detection of heavy elements in

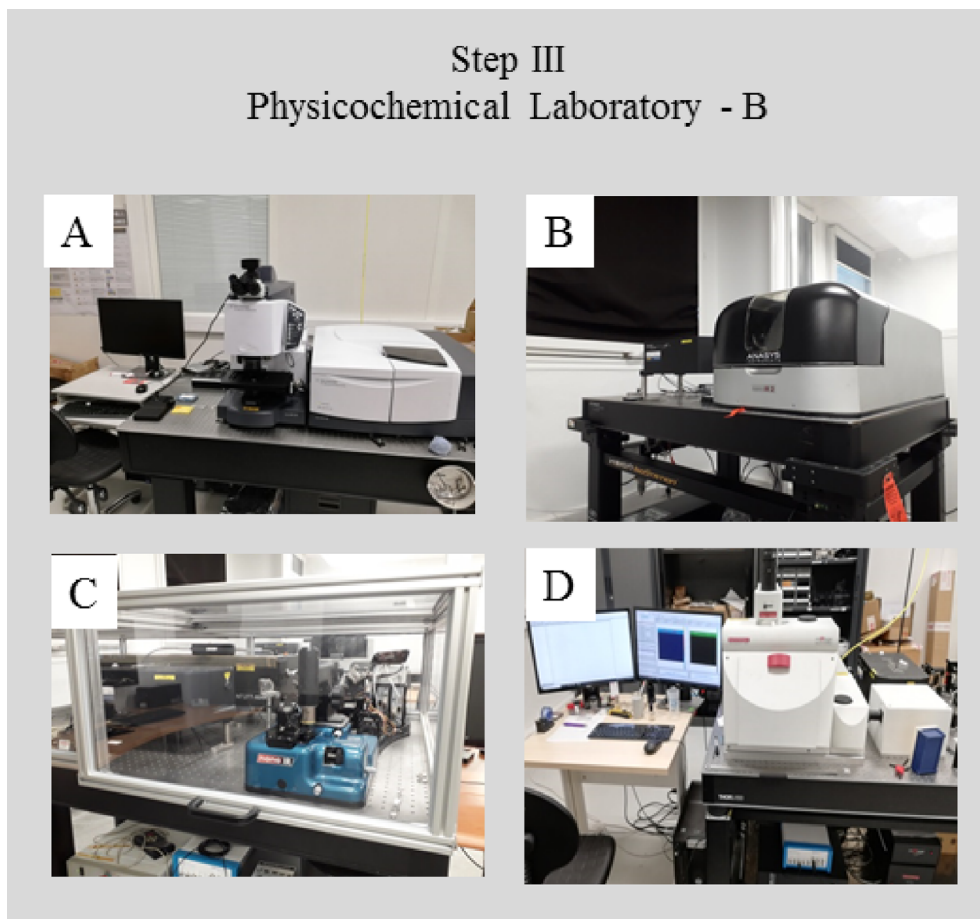


Figure 5. Characterization techniques available in a physics or chemistry laboratory (A) μ FTIR spectrophotometer implemented at soleil (SMIS beamline); (B) and (C) AFMIR experimental set up implemented on the MUSIICS (MUlti Scale Infrared Imaging platform for Complex Systems, located at the institute of physics and chemistry) platform, (D) OPTIR (Optical PhotoThermal IR) experimental set up (SMIS beamline).

tissues is much more favourable than that of light elements in a matrix of heavy elements. Experimental optimization can improve the elemental detection limits to concentrations in the range 10^{-9} to 10^{-12} g/g [132,133].

A similar significant improvement pertains to X-ray scattering experiments. Implemented on synchrotron radiation facilities, these offer the opportunity of measurements on isolated and micrometer sized single crystals. Guo *et al.* [134] have shown that acquisition and assembly of complete datasets from microcrystals may be routinely carried out on synchrotron microdiffraction beamlines.

It is worth underlining that a complete new set of spectroscopies such as XAS [135,136] or UV visible spectroscopy [137–140] is available at a synchrotron radiation centre. XAS is form of spectroscopy able to describe the electronic state and the local environment of trace elements in different kinds of material including those without long range order such as nanoparticles [141–146] and amorphous compounds [147,148]. In a recent review, Ma *et al.* [149] highlighted the emergence of nanoparticles in medicine which offer novel solutions to diagnosis and treatment of chronic kidney disease.

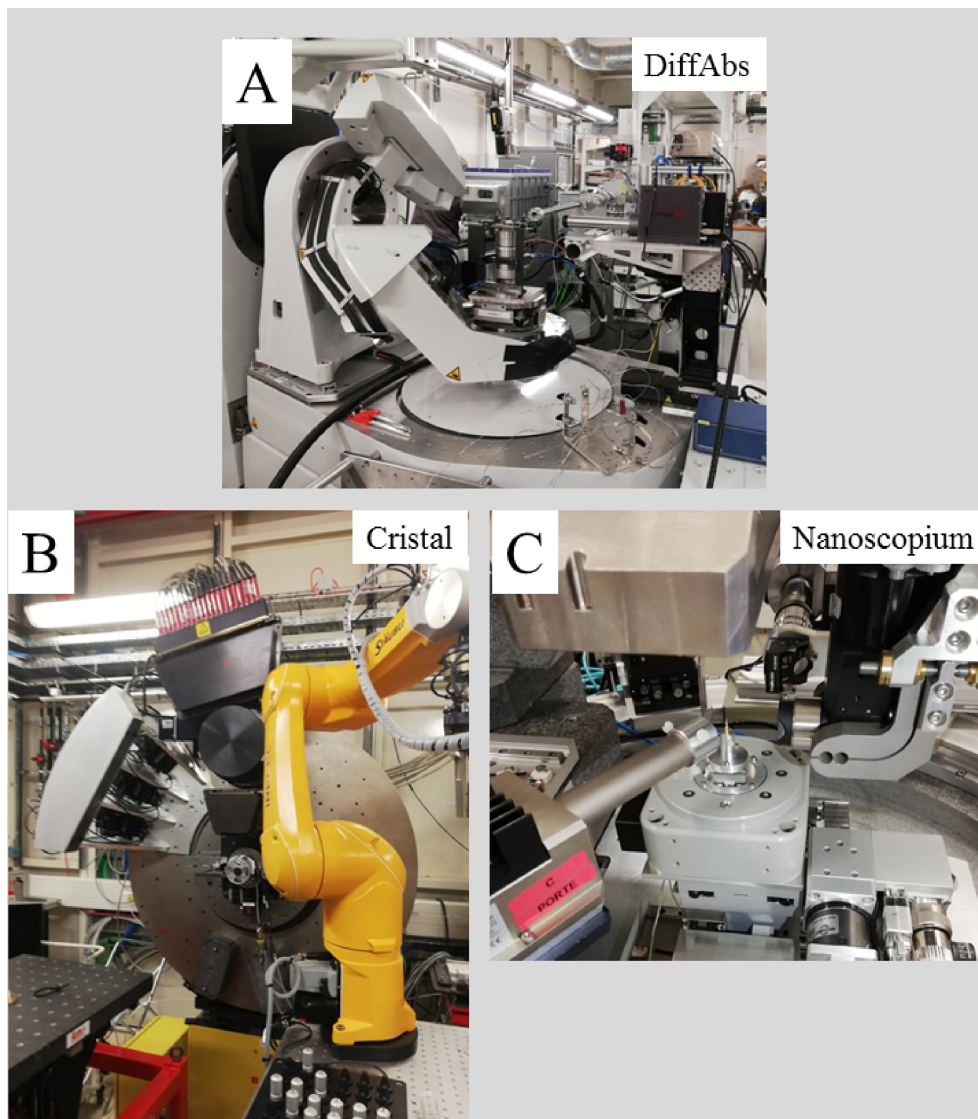


Figure 6. Three beamline experimental stations implemented on the Soleil synchrotron (A) Diffabs (details in Refs [82,83,149]), (B) Cristal (details in Refs [150–152]) and (C) Nanoscopium (details Ref. [153]).

As an example, we show in Figure 6 the different experimental beamlines we have used to characterize abnormal deposits in the human body namely Diffabs [85,86,150], Cristal [19,151] and Nanoscopium [154]. On these beamlines, XRD (Diffabs and Cristal), XAS (Diffabs) or nano-tomography (Nanoscopium) experiments can be performed. Note that other beamlines such as the Disco beamline can be of major importance [140].

3. Sample quantity and availability

Sample quantity and availability can determine the order in which experiments are performed. Large quantities impose no priority but it must be emphasized that several scans may have to be performed several times to take possible heterogeneity into account. Obviously, if the quantity of sample is low, non-destructive characterization techniques have to be implemented first.

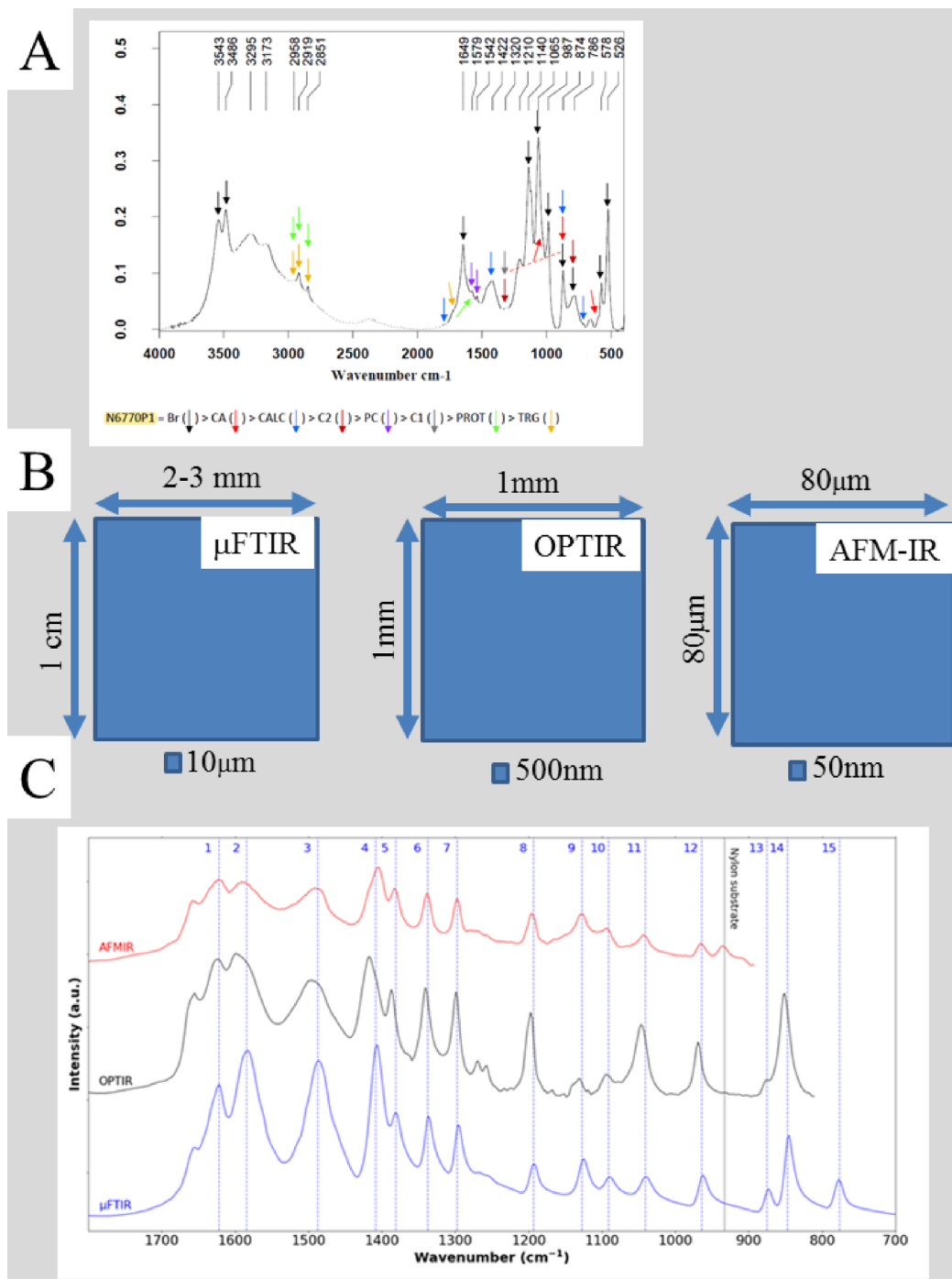


Figure 7. (A) Classical FTIR spectrum collected on part of a kidney stone dispersed in a KBr pellet; IR bands of various compounds present are indicated Br—brushite, CA—calcium phosphate apatite; CALC—calcite, C2—calcium oxalate dihydrate; PC—calcium palmitate; C1—calcium oxalate monohydrate, PROT—protein; TRG—triglycerides. (B) Typical dimensions of the maps which can be collected with μFTIR , OPTIR and AFMIR; the size of the probe for each is indicated. (C) Three IR spectra collected by the three experimental set ups— μFTIR (blue), OPTIR (black) and AFMIR (red).

Sometimes, sample quantity is simply insufficient to perform the experiments. For example, XRD experiments are performed on powder; the sample is typically inserted into a glass capillary ($\varnothing = 0.1\text{--}1\text{ mm}$) and mounted on a spinner rotating at several Hz to improve particle orientational averaging. Typically, for such experiments, less than $1\ \mu\text{L}$ of powder is enclosed in small glass or kapton capillaries.

Another technique needing a relatively large amount of sample is Nuclear Magnetic Resonance (NMR) spectroscopy. In order to gather information regarding local structural changes we have to take into account that the volume of a typical widely used “standard” solid state NMR sample holder (a “magic angle spinning” rotor) in a NMR experiment is around $80\ \mu\text{L}$. Also, it is possible to probe the local environment of several elements namely ^1H , ^{13}C , ^{31}P .

4. Advantages and inconveniences of high spatial resolution

High spatial resolution experiments are not always the most desirable. Lower spatial resolution characterization can resolve many important clinical problems. We will exemplify this by FTIR spectroscopy and the characterization of pathological renal calcification. In the case of kidney stones, several chemical compounds can be present in different proportions. Chemical compounds present at high abundance do not always define a medical diagnosis. Even a small proportion (5%) of struvite informs the clinician of bacterial infection. This factor imposes a requirement for an IR spectrum with a very high signal-to-noise ratio for there to be any possibility of detecting the presence of a small shoulder in the IR absorption bands or a small shift in their wavenumbers, which may help to distinguish up to 9 chemical compounds occurring in kidney stones.

It is very important to obtain a high quality IR spectrum from the core as well as the surface of kidney stones (Figure 7). Currently, only “classical” IR, in which measurements are performed on a small pellet, is capable of sufficiently high signal to noise. Also, high spatial resolution comes with characterization of only a small area of the sample. In the case of

a kidney biopsy the calcification area can be somewhere within a length of typically around one centimetre and width 1 mm; it is clearly impractical to spend a few hours searching the whole sample for ectopic calcification at a μFTIR spatial resolution of $10\ \mu\text{m}$, or by NanoIR for which the area of interest is limited to $80\ \mu\text{m}$ by $80\ \mu\text{m}$. Nevertheless, NanoIR experiments can represent the ultimate opportunity to determine chemical composition for very small deposits which cannot be characterized by μFTIR .

Finally, we must underline that polarization of the lasers used in NanoIR spectroscopy, either in the visible or the IR range, means that the intensity, and position of IR bands may vary, in the latter case by up to a few cm^{-1} . This can impose limitations on the precision of any description of abnormal biopsy deposit chemistry [122].

5. Conclusion

This paper proposes a hierarchical approach to characterize abnormal deposits in biological tissues [155], starting from *in vivo* measurements and progressing to a physicochemical description at the nanometer scale. The approach is based on a selected set of destructive and non-destructive characterization techniques which take into account the complexity of the physicochemistry of abnormal deposits in biological tissues, consistent with their exogenous and endogenous origins. Because some of these techniques need reference compounds such an approach supposes strong interactions with research teams able to generate nanomaterials [156–159]. Finally, it must be remembered that density functional theory [160,161] offers the opportunity to relate nanomaterial atomic structure to IR, Raman or XRD characteristics [162–166]. As emphasized by Khosroshahi *et al.* [167], it's time to use nanotechnology to change the conventional paradigm for analysis and diagnosis in cases of pathological deposits.

Conflicts of interest

Authors have no conflict of interest to declare.

References

- [1] D. Bazin, M. Daudon, C. Combes, C. Rey, *Chem. Rev.*, 2012, **112**, 5092-5120.

- [2] D. Bazin, M. Daudon, *J. Phys. D: Appl. Phys.*, 2012, **45**, article no. 383001.
- [3] M. Daudon, D. Bazin, *J. Phys. Conf. Ser.*, 2013, **425**, article no. 022006.
- [4] D. Bazin, M. Daudon, *J. Spectral Imaging*, 2019, **8**, article no. a16.
- [5] H. Colboc, D. Bazin, P. Moguelet, S. Reguer, R. Amode, C. Jouanneau, I. Lucas, L. Deschamps, V. Descamps, N. Kluger, *J. Eur. Acad. Dermatol. Venereol.*, 2020, **34**, e313-e315.
- [6] H. Colboc, D. Bazin, Ph. Moguelet, V. Frochet, R. Weil, E. Letavernier, Ch. Jouanneau, C. Frances, C. Bachmeyer, J.-F. Bernaudin, M. Daudon, *C. R. Chim.*, 2016, **19**, 1631-1641.
- [7] H. Colboc, P. Moguelet, D. Bazin, C. Bachmeyer, V. Frochet, R. Weil, E. Letavernier, C. Jouanneau, M. Daudon, J. F. Bernaudin, *J. Eur. Acad. Dermatol. Venereol.*, 2019, **33**, 198-203.
- [8] A. Dessombz, P. Méria, D. Bazin, M. Daudon, *PLoS One*, 2012, **7**, article no. e51691.
- [9] A. Ben Lakhdar, M. Daudon, M. C. Matthieu, A. Kellum, C. Balleyguier, D. Bazin, *C. R. Chim.*, 2016, **19**, 1610-1624.
- [10] J. A. M. R. Kunitake, S. Choi, K. X. Nguyen, M. M. Lee, F. He, D. Sudilovsky, P. G. Morris, M. S. Jochelson, C. A. Hudis, D. A. Muller, P. Fratzl, C. Fischbach, A. Masi, L. A. Estroff, *J. Struct. Biol.*, 2018, **202**, 25-34.
- [11] K. S. Shin, M. Laohajaratsang, S. Men, B. Figueroa, S. M. Dintzis, D. Fu, *Theranostics*, 2020, **10**, 5865-5878.
- [12] S. Gosling, D. Calabrese, J. Nallala, C. Greenwood, S. Pinder, L. King, J. Marks, D. Pinto, Th. Lynch, I. D. Lyburn, E. S. Hwang, Grand Challenge PRECISION Consortium, K. Rogers, N. Stone, *Analyst*, 2022, **147**, 1641-1654.
- [13] S. Gosling, R. Scott, C. Greenwood, P. Bouzy, J. Nallala, I. D. Lyburn, N. Stone, K. Rogers, *J. Mammary Gland Biol. Neoplasia*, 2019, **24**, 333-342.
- [14] X. Carpentier, M. Daudon, O. Traxer, P. Jungers, A. Mazouyes, G. Matzen, E. Véron, D. Bazin, *Urology*, 2009, **73**, 968-975.
- [15] D. Bazin, G. André, R. Weil, G. Matzen, E. Véron, X. Carpentier, M. Daudon, *Urology*, 2012, **79**, 786-790.
- [16] R. Flannigan, W. H. Choy, B. Chew, D. Lange, *Nat. Rev. Urol.*, 2014, **11**, 333-341.
- [17] E. J. Espinosa-Ortiz, B. H. Eisner, D. Lange, R. Gerlach, *Nat. Rev. Urol.*, 2019, **16**, 35-53.
- [18] M. Daudon, M. Petay, S. Vimont, A. Denizet, F. Tielens, J.-Ph. Haymann, E. Letavernier, V. Frochet, D. Bazin, *C. R. Chim.*, 2022, **25**, no. S1, 315-334, Online first.
- [19] D. Bazin, R. J. Papoular, E. Elkaim, R. Weil, D. Thiaudière, C. Pisapia, B. Ménez, N. S. Hwang, F. Tielens, M. Livrozet, E. Boudierlique, J.-Ph. Haymann, E. Letavernier, L. Hennet, V. Frochet, M. Daudon, *C. R. Chim.*, 2022, **25**, no. S1, 343-354, Online first.
- [20] E. N. Taylor, M. J. Stampfer, G. C. Curhan, *Kidney Int.*, 2005, **68**, 1230-1235.
- [21] M. Daudon, B. Lacour, P. Jungers, *Nephrol. Dial. Transpl.*, 2005, **20**, 468-469.
- [22] M. Daudon, E. Letavernier, R. Weil, E. Véron, G. Matzen, G. André, D. Bazin, *C. R. Chim.*, 2016, **19**, 1527-1534.
- [23] M. Daudon, P. Jungers, D. Bazin, *New Engl. J. Med.*, 2008, **359**, 100-102.
- [24] M. Daudon, P. Jungers, D. Bazin, *New Engl. J. Med.*, 2009, **360**, 1680.
- [25] A. Dessombz, E. Letavernier, J.-Ph. Haymann, D. Bazin, M. Daudon, *J. Urol.*, 2015, **193**, 1564-1569.
- [26] F. Meiouet, S. El Kabbaj, M. Daudon, *C. R. Chim.*, 2022, **25**, no. S1, 281-293.
- [27] D. Bazin, M. Daudon, P. Chevallier, S. Rouzière, E. Elkaim, D. Thiaudière, B. Fayard, E. Foy, P. A. Albouy, G. André, G. Matzen, E. Véron, *Ann. Biol. Clin. (Paris)*, 2006, **64**, 125-139.
- [28] D. Bazin, J.-Ph. Haymann, E. Letavernier, J. Rode, M. Daudon, *Presse Med.*, 2014, **43**, 135-148.
- [29] D. Bazin, E. Letavernier, J.-P. Haymann, P. Méria, M. Daudon, *Prog. Urol.*, 2016, **26**, 608-618.
- [30] D. Bazin, E. Letavernier, J.-Ph. Haymann, *C. R. Chim.*, 2016, **19**, 1395-1403.
- [31] S. R. Mulay, H.-J. Anders, *N. Engl. J. Med.*, 2016, **374**, 2465-2476.
- [32] D. Bazin, M. Daudon, "Nouvelles Méthodes d'étude des calculs et plaques de Randall", in *Actualités néphrologiques*, Ed. Médecine-Sciences, Flammarion, Paris, 2010, ISBN : 978-2-257-20409-7.
- [33] M. Daudon, D. Bazin, "Application of physical methods to kidney stones and Randall's plaque characterization", in *Urolithiasis: Basic Science and Clinical Practice* (J. J. Talati, H. G. Tiselius, D. M. Albala, Z. Ye, eds.), Springer-Verlag, London, 2012, 683-707.
- [34] L. N. Poloni, M. D. Ward, *Chem. Mater.*, 2014, **26**, 477-495.
- [35] D. Bazin, Ch. Jouanneau, S. Bertazzo, Ch. Sandt, A. Dessombz, M. Réfrégiers, P. Dumas, J. Frederick, J.-Ph. Haymann, E. Letavernier, P. Ronco, M. Daudon, *C. R. Chim.*, 2016, **19**, 1439-1450.
- [36] E. Tsolaki, S. Bertazzo, *Materials*, 2019, **12**, article no. 3126.
- [37] D. Bazin, E. Letavernier, J. P. Haymann, V. Frochet, M. Daudon, *Ann. Biol. Clin.*, 2020, **78**, 349-362.
- [38] N. Vidavsky, J. A. M. R. Kunitake, L. A. Estroff, *Adv. Healthc. Mater.*, 2021, **10**, article no. 2001271.
- [39] D. Bazin, X. Carpentier, I. Brocheriou, P. Dorfmueller, S. Aubert, Ch. Chappard, D. Thiaudière, S. Reguer, G. Waychunas, P. Jungers, M. Daudon, *Biochimie*, 2009, **91**, 1294-1300.
- [40] D. Bazin, C. Chappard, C. Combes, X. Carpentier, S. Rouzière, G. André, G. Matzen, M. Allix, D. Thiaudière, S. Reguer, P. Jungers, M. Daudon, *Osteoporos. Int.*, 2009, **20**, 1065-1075.
- [41] Ch. Chappard, G. André, M. Daudon, D. Bazin, *C. R. Chim.*, 2016, **19**, 1625-1630.
- [42] H. Colboc, J. Fontaine, D. Bazin, V. Frochet, E. Letavernier, M. Daudon, N. Laporte, S. Rouzière, M. Reby, A. Galezowski, Ch. Forasassi, S. Meaume, *J. Gerontol. A Biol. Sci. Med. Sci.*, 2022, **77**, 27-32.
- [43] H. Colboc, Ph. Moguelet, E. Letavernier, V. Frochet, J.-F. Bernaudin, R. Weil, S. Rouzière, P. Senet, C. Bachmeyer, N. Laporte, I. Lucas, V. Descamps, R. Amode, F. Brunet-Possenti, N. Kluger, L. Deschamps, A. Dubois, S. Reguer, A. Somogyi, K. Medjoubi, M. Refregiers, M. Daudon, D. Bazin, *C. R. Chim.*, 2022, **25**, no. S1, 445-476.
- [44] M. MATHONNET, A. Dessombz, D. Bazin, R. Weil, F. Triponez, M. Pusztaszeri, M. Daudon, *C. R. Chim.*, 2016, **19**, 1672-1678.
- [45] J. Guerlain, S. Perie, M. Lefevre, J. Perez, S. Vandermeersch,

- Ch. Jouanneau, L. Huguet, V. Frochot, E. Letavernier, R. Weil, S. Rouzière, D. Bazin, M. Daudon, J. P. Haymann, *PLoS One*, 2019, **14**, article no. e0224138.
- [46] L. Henry, D. Bazin, C. Policar, J.-Ph. Haymann, M. Daudon, V. Frochot, M. Mathonnet, *C. R. Chim.*, 2022, **25**, no. S1, 503-515.
- [47] A. Dessombz, P. Méria, D. Bazin, E. Foy, S. Rouzière, R. Weil, M. Daudon, *Prog. Urol.*, 2011, **21**, 940-945.
- [48] M. Daudon, C. A. Bader, P. Jungers, *Scanning Microsc.*, 1993, **7**, 1081-1106.
- [49] M. Daudon, P. Jungers, D. Bazin, *AIP Conf. Proc.*, 2008, **1049**, 199-215.
- [50] M. Daudon, A. Dessombz, V. Frochot, E. Letavernier, J.-Ph. Haymann, P. Jungers, D. Bazin, *C. R. Chim.*, 2016, **19**, 1470-1491.
- [51] M. Daudon, E. Letavernier, V. Frochot, J.-Ph. Haymann, D. Bazin, P. Jungers, *C. R. Chim.*, 2016, **19**, 1504-1513.
- [52] C. Nguyen, H.-K. Ea, D. Bazin, M. Daudon, F. Lioté, *Arthritis Rheum.*, 2010, **62**, 2829-2830.
- [53] H.-K. Ea, C. Nguyen, D. Bazin, A. Bianchi, J. Guicheux, P. Reboul, M. Daudon, F. Lioté, *Arthritis Rheum.*, 2010, **63**, 10-18.
- [54] Ch. Nguyen, H. Korng Ea, D. Thiaudière, S. Reguer, D. Han-nouche, M. Daudon, F. Lioté, D. Bazin, *J. Synchrotron Rad.*, 2011, **18**, 475-480.
- [55] A. Gauffenic, D. Bazin, Ch. Combes, M. Daudon, H.-K. Ea, *C. R. Chim.*, 2022, **25**, no. S1, 517-534.
- [56] M. Petay, M. Cherfan, A. Deniset-Besseau, E. Boudierlique, S. Reguer, A. Dazzi, J. Mathurin, M. Daudon, V. Frochot, J.-Ph. Haymann, E. Letavernier, D. Bazin, *C. R. Chim.*, 2022, **25**, no. S1, 553-576.
- [57] Y. Li, D. G. Reid, D. Bazin, M. Daudon, M. J. Duer, *C. R. Chim.*, 2016, **19**, 1665-1671.
- [58] C. J. Cros, D. Bazin, A. Kellum, V. Rebours, M. Daudon, *C. R. Chim.*, 2016, **19**, 1642-1664.
- [59] S. Rouzière, D. Bazin, M. Daudon, *C. R. Chim.*, 2016, **19**, 1404-1415.
- [60] D. Bazin, E. Foy, S. Reguer, S. Rouzière, B. Fayard, H. Colboc, J.-Ph. Haymann, M. Daudon, C. Mocuta, *C. R. Chim.*, 2022, **25**, no. S1, 165-188.
- [61] D. Bazin, M. Daudon, *Ann. Biol. Clin.*, 2015, **73**, 517-534.
- [62] D. Bazin, E. Boudierlique, M. Daudon, V. Frochot, J.-Ph. Haymann, E. Letavernier, F. Tielens, R. Weil, *C. R. Chim.*, 2022, **25**, no. S1, 37-60.
- [63] N. Quy Dao, M. Daudon, *Infrared and Raman Spectra of Calculi*, Elsevier, Paris, 1997.
- [64] M. Daudon, D. Bazin, *C. R. Chim.*, 2016, **19**, 1416-1423.
- [65] V. Castiglione, P.-Y. Sacré, E. Cavalier, P. Hubert, R. Gadisseur, E. Ziemons, *PLoS One*, 2018, **13**, article no. e0201460.
- [66] I. T. Lucas, D. Bazin, M. Daudon, *C. R. Chim.*, 2022, **25**, no. S1, 83-103.
- [67] V. Sablinskas, S. Talmosaityte, M. Pucetaite, *C. R. Chim.*, 2022, **25**, no. S1, 73-82.
- [68] V. Vuiblet, T. T. Nguyen, A. Wynckel, M. Fere, L. Van-Gulick, V. Untereiner, P. Birembaut, P. Rieu, O. Piot, *Analyst*, 2015, **140**, 7382-7390.
- [69] A. Guinier, *Théorie et technique de la radiocristallographie*, Dunod, Paris, 1964.
- [70] A. Le Bail, *Adv. X-Ray Anal.*, 2000, **42**, 191-203.
- [71] F. Damay, D. Bazin, M. Daudon, G. André, *C. R. Chim.*, 2016, **19**, 1432-1434.
- [72] D. Bazin, V. Frochot, J.-Ph. Haymann, E. Letavernier, M. Daudon, *C. R. Chim.*, 2022, **25**, no. S1, 133-147.
- [73] Ph. Sciau, *Adv. Imaging Electron Phys.*, 2016, **198**, 43-67.
- [74] S. J. Pennycook, M. F. Chisholm, A. R. Lupini, M. Varela, A. Y. Borisevich, M. P. Oxley, W. D. Luo, K. van Benthem, S.-H. Oh, D. L. Sales, S. I. Molina, J. García-Barriocanal, C. Leon, J. Santamaría, S. N. Rashkeev, S. T. Pantelides, *Phil. Trans. Math. Phys. Eng. Sci.*, 2009, **367**, 3709-3733.
- [75] C. Verrier, D. Bazin, L. Huguet, O. Stéphan, A. Gloter, M.-Ch. Verpont, V. Frochot, J.-Ph. Haymann, I. Brocheriou, O. Traxer, M. Daudon, E. Letavernier, *J. Urol.*, 2016, **196**, 1566-1574.
- [76] C. Gay, E. Letavernier, M.-Ch. Verpont, M. Walls, D. Bazin, M. Daudon, N. Nassif, O. Stephan, M. de Fruto, *ACS Nano*, 2020, **14**, 1823-1836.
- [77] J. D. Boerckel, D. E. Mason, A. M. McDermott, E. Alsberg, *Stem Cell Res. Ther.*, 2014, **5**, article no. N0144.
- [78] D. Bazin, C. Leroy, F. Tielens, Ch. Bonhomme-Courry, F. Damay, D. Le Denmat, J. Sadoine, J. Rode, V. Frochot, E. Letavernier, J.-Ph. Haymann, M. Daudon, *C. R. Chim.*, 2016, **19**, 1492-1503.
- [79] O. L. Katsamenis, M. Olding, J. A. Warner, D. S. Chatelet, M. G. Jones, G. G. Sgalla, B. Smit, O. J. Larkin, I. Haig, L. Richeldi, I. Sinclair, P. M. Lackie, Ph. Schneider, *Am. J. Pathol.*, 2019, **189**, 1608-1620.
- [80] J. C. Williams Jr, J. E. Lingeman, M. Daudon, D. Bazin, *C. R. Chim.*, 2022, **25**, no. S1, 61-72.
- [81] H. Hertz, *Ann. Physik*, 1887, **31**, 983-1000.
- [82] K. Siegbahn *et al.*, *Nova Acta Regiae Soc. Sci. Ser. IV*, 1967, **20**, 1-15.
- [83] F. S. Brigiano, D. Bazin, F. Tielens, *C. R. Chim.*, 2022, **25**, no. S1, 149-163.
- [84] D. E. Sayers, E. A. Stern, F. W. Lytle, *Phys. Rev. Lett.*, 1971, **27**, 1204-1207.
- [85] S. Reguer, C. Mocuta, D. Thiaudière, M. Daudon, D. Bazin, *C. R. Chim.*, 2016, **19**, 1424-1431.
- [86] D. Bazin, S. Reguer, D. Vantelon, J.-Ph. Haymann, E. Letavernier, V. Frochot, M. Daudon, E. Esteve, H. Colboc, *C. R. Chim.*, 2022, **25**, no. S1, 189-208.
- [87] R. F. Egerton, *Electron Energy-Loss Spectroscopy in the Electron Microscope*, Springer US, Boston, 2011.
- [88] R. F. Egerton, *Rep. Prog. Phys.*, 2008, **72**, article no. 016502.
- [89] H. Colas, L. Bonhomme-Courry, C. Diogo-Coelho, F. Tielens, F. Babonneau, C. Gervais, D. Bazin, D. Laurencin, M. Smith, J. V. Hanna, M. Daudon, C. Bonhomme, *Cryst. Eng. Commun.*, 2013, **15**, 8840-8847.
- [90] A. Dessombz, G. Coulibaly, B. Kirakoya, R. W. Ouedraogo, A. Lengani, S. Rouzière, R. Weil, L. Picaud, L. Bonhomme, F. Babonneau, D. Bazin, M. Daudon, *C. R. Chim.*, 2016, **19**, 1573-1579.
- [91] C. Leroy, L. Bonhomme-Courry, Ch. Gervais, F. Tielens, F. Babonneau, M. Daudon, D. Bazin, E. Letavernier, D. Laurencin, D. Iuga, J. Hanna, M. Smith, Ch. Bonhomme, *Magn. Reson.*, 2021, **2**, 1-13.
- [92] AFCA Insider, "Evaluate Injuries With Portable Diagnostic X-ray From MinXray", 2022, <https://insider.afca.com/>

- evaluate-injuries-with-portable-diagnostic-x-ray-from-minxray/.
- [93] X. Zhang, A. J. Specht, E. Wells, M. G. Weisskopf, J. Weuve, L. H. Nie, *Sci. Total Environ.*, 2021, **753**, article no. 142351.
- [94] Sheffield Dermatology Research, "Research".
- [95] E. Guevara, J. C. Torres-Galván, M. G. Ramírez-Eliás, C. Luevano-Contreras, F. J. González, *Biomed. Opt. Express*, 2018, **9**, 4998-5010.
- [96] K. Eklund, R. Klefsgård, B. Ivarsson, M. Geijer, *Gerontology*, 2012, **58**, 107-111.
- [97] J. Börjesson, M. Alpsten, S. Huang, R. Jonson, S. Mattsson, Ch. Thornberg, "In vivo X-ray fluorescence analysis with applications to platinum, gold and mercury in man — experiments, improvements, and patient measurements", in *Human Body Composition* (K. J. Ellis, J. D. Eastman, eds.), Basic Life Sciences, vol. 60, Springer, Boston, MA, 1993.
- [98] R. Jonson, S. Mattsson, B. Unsgaard, *Phys. Med. Biol.*, 1988, **33**, 847-857.
- [99] L. Brancalion, M. P. Bamberg, T. Sakamaki, N. Kollias, *J. Invest. Dermatol.*, 2001, **116**, 380-386.
- [100] A. P. M. Michel, S. Liakat, K. Bors, C. F. Gmach, *Biomed. Opt. Express.*, 2013, **4**, 520-530.
- [101] A. Bauer, O. Hertzberg, A. Küderle, D. Strobel, M. A. Pleitez, W. Mäntele, *J. Biophotonics*, 2018, **11**, article no. e201600261.
- [102] Y. Yarovoy, D. M. Drutis, Th. M. Hancewicz, U. Garczarek, K. P. Ananthapadmanabhan, M. Misra, *Appl. Spectrosc.*, 2019, **73**, 182-194.
- [103] P. J. Caspers, G. W. Lucassen, R. Wolthuis, H. A. Bruining, G. J. Puppels, *Biospectroscopy*, 1998, **4**, S31-S39.
- [104] P. J. Caspers, G. W. Lucassen, G. J. Puppels, *Biophys. J.*, 2003, **85**, 572-580.
- [105] H. Lui, J. Zhao, D. McLean, H. Zeng, *Cancer Res.*, 2012, **72**, 2491-2500.
- [106] A. Le Bail, D. Bazin, M. Daudon, A. Brochot, V. Robbez-Masson, V. Maisonneuve, *Acta Cryst. B*, 2009, **65**, 350-354.
- [107] M. Daudon, D. Bazin, K. Adil, A. Le Bail, *Acta Cryst. E*, 2011, **67**, article no. 1458.
- [108] U. Holzwarth, N. Gibson, *Nat. Nanotechnol.*, 2011, **6**, article no. 534.
- [109] K. Wilkinson, J. Lundkvist, G. Seisenbaeva, V. Kessler, *Environ. Pollut.*, 2011, **159**, 311-318.
- [110] M. C. Mahoney, M. S. Newell, *Radiology*, 2013, **268**, 12-24.
- [111] R. N. Uppot, M. G. Harisinghani, D. A. Gervais, *Am. J. Roentgenol.*, 2010, **194**, 1443-1449.
- [112] D. Fournier, F. Lepoutre, A. Boccara, *J. Phys.*, 1983, **44**, 479-482.
- [113] N. Baden, H. Kobayashi, N. Urayama, *Int. J. Polym. Anal. Charact.*, 2020, **25**, 1-7.
- [114] A. J. Wang, E. P. Dillon, S. Maharjan, K.-S. Liao, B. P. McElhenny, T. Tong, S. Chen, J. Bao, S. A. Curran, *Adv. Mater. Interfaces*, 2021, **8**, article no. 2001720.
- [115] A. Dazzi, F. Glotin, R. Carminati, *J. Appl. Phys.*, 2010, **107**, article no. 124519.
- [116] A. Dazzi, C. B. Prater, *Chem. Rev.*, 2017, **117**, 5146-5173.
- [117] L. Bildstein, A. Deniset-Besseau, I. Pasini, Ch. Mazilier, Y. W. Keoung, A. Dazzi, N. Baghdadli, *Anal. Chem.*, 2020, **92**, 11498-11504.
- [118] G. Latour, L. Robinet, A. Dazzi, F. Portier, A. Deniset-Besseau, M.-C. Schanne-Klein, *Sci. Rep.*, 2016, **6**, article no. 26344.
- [119] E. Esteve, Y. Luque, J. Waeytens, D. Bazin, L. Mesnard, Ch. Jouanneau, P. Ronco, A. Dazzi, M. Daudon, A. Deniset-Besseau, *Anal. Chem.*, 2020, **92**, 7388-7392.
- [120] D. Bazin, M. Rabant, J. Mathurin, M. Petay, A. Deniset-Besseau, A. Dazzi, Y. Su, E. P. Hessouc, F. Tielens, F. Borondic, M. Livrozet, E. Boudelicque, J.-Ph. Haymann, E. Letavernier, V. Frochot, M. Daudon, *C. R. Chim.*, 2022, **25**, no. S1, 489-502.
- [121] D. Bazin, E. Boudierlique, E. Tang, M. Daudon, J.-Ph. Haymann, V. Frochot, E. Letavernier, E. Van de Perre, J. C. Williams Jr., J. E. Lingeman, F. Borondics, *C. R. Chim.*, 2022, **25**, no. S1, 105-131.
- [122] J. Mathurin, A. Deniset-Besseau, D. Bazin, E. Dartois, M. Wagner, A. Dazzi, *J. Appl. Phys.*, 2022, **131**, article no. 010901.
- [123] N. Al Harbi, S. Y. Lee, *Rev. Scientific Instrum.*, 2003, **74**, article no. 2540.
- [124] W. Thomlinson, P. Suortti, D. Chapman, *Nucl. Instrum. Methods Phys. Res. Sect. A*, 2005, **543**, 288-296.
- [125] H. Elleaume, S. Fiedler, B. Bertrand, T. Brochard, F. Estève, J. F. Le Bas, G. Le Duc, C. Nemoz, M. Renier, P. Suortti, W. Thomlinson, *Phys. Med. Biol.*, 2000, **45**, L39-L43.
- [126] S. Bayat, G. Le Duc, L. Porra, G. Berruyer, C. Nemoz, S. Monfraix, S. Fiedler, W. Thomlinson, P. Suortti, C. G. Standertskjold Nordenstam, A. R. A. Sovijarvi, *Phys. Med. Biol.*, 2001, **46**, 3287-3299.
- [127] Synchrotron-Soleil, <https://www.synchrotron-soleil.fr/fr>.
- [128] A. Laissue, H. Blattmann, M. Di Michiel, D. N. Slatkin, N. Lyubimova, R. Guzman, W. Zimmermann, S. Birrer, T. Bley, P. Kircher, R. Stettler, R. Fatzer, A. Jaggy, H. M. Smilowitz, E. Brauer, A. Bravin, G. Le Duc, C. Nemoz, M. Renier, W. Thomlinson, J. Stepanek, H.-P. Wagner, *Proc. SPIE*, 2001, **4508**, 65-73.
- [129] A. van der Ent, W. J. Przybyłowicz, M. D. de Jonge, H. H. Harris, Ch. G. Ryan, G. Tytko, D. J. Paterson, A. D. Barnabas, P. M. Kopittke, J. Mesjasz-Przybyłowicz, *New Phytol.*, 2018, **218**, 432-452.
- [130] Y. S. Chu, J. M. Yi, F. De Carlo, Q. Shen, W. K. Lee, H. J. Wu, C. L. Wang, J. Y. Wang, C. J. Liu, C. H. Wang, S. R. Wu, C. C. Chien, Y. Hwu, A. Tkachuk, W. Yun, M. Feser, K. S. Liang, C. S. Yang, J. H. Je, G. Margaritondo, *Appl. Phys. Lett.*, 2008, **92**, article no. 103119.
- [131] X. Huang, H. Yan, E. Nazaretski, R. Conley, N. Bouet, J. Zhou, K. Lauer, L. Li, D. Eom, D. Legnini, R. Harder, I. K. Robinson, Y. S. Chu, *Sci. Rep.*, 2013, **3**, article no. 3562.
- [132] M. R. Gherase, D. E. B. Fleming, *Crystals*, 2020, **10**, article no. 12.
- [133] M. J. Pushie, I. J. Pickering, M. Korbas, M. J. Hackett, G. N. George, *Chem. Rev.*, 2014, **114**, 8499-8541.
- [134] G. Guo, M. R. Fuchs, W. Shi, J. Skinner, E. Berman, C. M. Ogata, W. A. Hendrickson, S. McSweeney, Q. Liu, *IUCr*, 2018, **5**, 238-246.
- [135] D. E. Sayers, F. W. Lytle, E. A. Stern, *Adv. X-Ray Anal.*, 1970, **13**, 248-271.
- [136] E. A. Stern, D. E. Sayers, F. W. Lytle, *Phys. Rev. B*, 1975, **11**, 4836-4946.
- [137] F. Jamme, S. Kascakova, S. Villette, F. Allouche, S. Pallu, V. Rouam, M. Réfrégiers, *Biol. Cell.*, 2013, **105**, 277-288.
- [138] F. Jamme, S. Villette, A. Giuliani, V. Rouam, F. Wien, B. Lagarde, M. Réfrégiers, *Microsc. Microanal.*, 2010, **16**, 507-514.

- [139] E. Esteve, C. Jouanneau, E. Letavernier, P. Ronco, M. Daudon, D. Bazin, M. Refregiers, *Néphrol. Théor.*, 2015, **11**, 279-280.
- [140] E. Estève, D. Buob, F. Jamme, Ch. Jouanneau, S. Kascakova, J. Ph. Haymann, E. Letavernier, L. Galmiche, P. Ronco, M. Daudon, D. Bazin, M. Réfrégiers, *J. Synchrotron Radiat.*, 2022, **29**, 214-223.
- [141] J. Moonen, J. Slot, L. Lefferts, D. Bazin, H. Dexpert, *Phys. B*, 1995, **208**, 689-690.
- [142] D. Bazin, D. Sayers, J. Rehr, *J. Phys. Chem. B*, 1997, **101**, 11040-11050.
- [143] D. Bazin, D. Sayers, J. J. Rehr, C. Mottet, *J. Phys. Chem.*, 1997, **101**, 5332-5336.
- [144] D. Bazin, J. Rehr, *J. Phys. Chem. C*, 2003, **107**, 12398-12402.
- [145] D. Bazin, J. Rehr, *J. Phys. Chem. C*, 2011, **115**, 23233-23236.
- [146] D. Bazin, J. Lynch, M. Ramos-Fernandez, *Oil Gas Sci. Technol. - Rev. IFP*, 2003, **58**, 667-683.
- [147] D. Sayers, M. Paesler, *J. Phys.*, 1986, **47**, 349-356.
- [148] R. Bellissent, A. Chenevas, P. Lagarde, D. Bazin, D. Raoux, *J. Non-Cryst. Solids*, 1983, **59**, 237-240.
- [149] Y. Ma, F. Cai, Y. Li, J. Chen, F. Hana, W. Lin, *Bioact. Mater.*, 2020, **5**, 732-743.
- [150] E. Esteve, S. Reguer, C. Boissière, C. Chanéac, G. Lugo, Ch. Jouanneau, C. Mocuta, D. Thiaudière, N. Leclercq, B. Leyh, J.-F. Greisch, J. Berthault, M. Daudon, P. Ronco, D. Bazin, *J. Synchrotron Radiat.*, 2017, **24**, 991-999.
- [151] D. Bazin, M. Daudon, E. Elkaim, A. Le Bail, L. Smrcok, *C. R. Chim.*, 2016, **19**, 1535-1541.
- [152] P. Gras, C. Rey, G. André, C. Charvillat, S. Sarda, C. Combes, *Acta Crystallogr. B*, 2016, **72**, 96-101.
- [153] A. Somogyi, K. Medjoubi, G. Baranton, V. Le Roux, M. Ribbens, F. Polack, P. Philippot, J. P. Samama, *J. Synchrotron Radiat.*, 2015, **22**, 1118-1129.
- [154] F. Brunet-Possenti, L. Deschamps, H. Colboc, A. Somogyi, K. Medjoubi, D. Bazin, V. Descamps, *J. Eur. Acad. Dermatol. Venereol.*, 2018, **32**, e442-e443.
- [155] D. Bazin, *C. R. Chim.*, 2022, **25**, no. S3, Forthcoming.
- [156] C. Rey, C. Combes, C. Drouet, A. Lebugle, H. Sfihi, A. Barroug, *Mater. Sci. Eng. Technol.*, 2007, **38**, 996-1002.
- [157] C. Drouet, C. Rey, in *Nanostructured Biomaterials for Regenerative Medicine* (V. Guarino, M. Iafisco, S. Srpiano, eds.), Woodhead Publishing Series in Biomaterials, Elsevier, Duxford, UK, 2020, 223-254.
- [158] C. Rey, C. Combes, C. Drouet, H. Sfihi, A. Barroug, *Mater. Sci. Eng. C*, 2007, **27**, 198-205.
- [159] C. Combes, C. Rey, *Minerals*, 2016, **6**, article no. 34.
- [160] F. Tielens, J. Vekeman, D. Bazin, M. Daudon, *C. R. Chim.*, 2022, **25**, no. S1, 209-218.
- [161] D. Bazin, J. Vekeman, Q. Wang, X. Deraet, F. De Proft, H. Guesmi, F. Tielens, *C. R. Chim.*, 2022, **25**, no. S3, Online first.
- [162] I. Petit, G. D. Belletti, Th. Debroise, M. J. Llansola-Portoles, I. T. Lucas, C. Leroy, Ch. Bonhomme, L. Bonhomme-Coury, D. Bazin, M. Daudon, E. Letavernier, J. Ph. Haymann, V. Frochot, F. Babonneau, P. Quaino, F. Tielens, *ChemistrySelect*, 2018, **3**, 8801-8812.
- [163] Th. Debroise, E. Colombo, G. Belletti, J. Vekeman, Y. Su, R. Papoular, N. S. Hwang, D. Bazin, M. Daudon, P. Quaino, F. Tielens, *Cryst. Growth Des.*, 2020, **20**, 2553-2561.
- [164] D. Bazin, F. Tielens, *Appl. Catal.*, 2015, **504**, 631-641.
- [165] F. Tielens, D. Bazin, *C. R. Chim.*, 2018, **21**, 174-181.
- [166] I. C. Oğuz, H. Guesmi, D. Bazin, F. Tielens, *J. Phys. Chem. C*, 2019, **123**, 20314-20318.
- [167] H. T. Khosroshahi, B. Abedi, S. Daneshvar, Y. Sarbaz, A. S. Babil, *Int. J. Biomed. Imaging*, 2017, **2017**, article no. 6141734.