



ELSEVIER

Contents lists available at ScienceDirect

Comptes Rendus Geoscience

www.sciencedirect.com



Hydrology, environment (Surface geochemistry)

Lithium isotopes in foraminifera shells as a novel proxy for the ocean dissolved inorganic carbon (DIC)

Nathalie Vigier^{a,*,c}, Claire Rollion-Bard^{a,1}, Yaël Levenson^b, Jonathan Erez^b^a CRPG-CNRS, université de Lorraine, 15, rue Notre-Dame-des-Pauvres, 54501 Vandœuvre-les-Nancy cedex, France^b Institute of Earth Sciences, The Hebrew University of Jerusalem, Jerusalem 91904, Israel^c CNRS, UPMC, laboratoire d'océanographie de Villefranche, 06230 Villefranche-sur-Mer, France

ARTICLE INFO

Article history:

Received 2 December 2014

Accepted after revision 4 December 2014

Available online 13 January 2015

Keywords:

Lithium isotope fractionation

Foraminifera

DIC proxy

 $\delta^7\text{Li}$

pH

Temperature

Calcite

ABSTRACT

Past ocean pH and pCO_2 are critical parameters for establishing relationships between Earth's climate and the carbon cycle. Previous pCO_2 estimates are associated with large uncertainties and are debated. In this study, laboratory cultures of the foraminiferan genus *Amphistegina* were performed in order to examine the possible factors that control the Li isotope composition ($\delta^7\text{Li}$) of their shells. $\delta^7\text{Li}$ is insensitive to temperature and pH variations but correlates positively with the Dissolved Inorganic Carbon (DIC) of seawater. Li/Ca ratio in the shells shows negative correlation with $\delta^7\text{Li}$, consistent with published data for planktonic foraminifera from core tops and from short periods during the Cenozoic. We propose that the sensitivity of $\delta^7\text{Li}$ and Li/Ca ratio to DIC is a biological phenomenon and is related to biomineralization mechanisms in foraminifera. We used the published foraminiferal $\delta^7\text{Li}$ records, and our experimental results, to determine the paleo-ocean DIC and pH for the last glacial–interglacial cycle. The results are consistent with published estimates of pH and pCO_2 based on boron isotopes and ice cores. We suggest Li and its isotopes may serve as a new complementary proxy for the paleo-ocean carbonate chemistry.

© 2014 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

1. Introduction

Atmospheric CO_2 is a greenhouse gas likely responsible for major climate changes during the history of the Earth. Rapid CO_2 increase is also one of the main factors responsible for the recent global warming (IPCC, 2013). In order to quantify and calibrate the sensitivity of global warming to changes in atmospheric CO_2 concentrations, past pCO_2 variations need to be determined and compared

with contemporaneous temperature estimates. For the last 800,000 years, this has been possible using direct atmospheric pCO_2 measurements and temperature estimates from ice cores (e.g., Siegenthaler et al., 2005). For longer timescales, proxies of CO_2 sources (i.e. volcanic activity) and sinks (continental weathering) may be used for modeling these variations (e.g., Godderis and Veizer, 2000). Biological proxies such as stomatal indices and carbon isotope compositions ($\delta^{13}\text{C}$) of marine organic matter (notably of alkenones, which represent fossil coccolithophores or organic material from diatom frustules) can also provide estimates of atmospheric CO_2 (Kürschner et al., 2008; Pagani et al., 2005). However, stomata fossils are rare and phytoplankton $\delta^{13}\text{C}$ can be influenced by unrelated

* Corresponding author.

E-mail address: nathalie.vigier@obs-vlfr.fr (N. Vigier).

¹ Institut de physique du globe de Paris (IPGP), université Paris–Diderot, UMR CNRS 7154, 1, rue Jussieu, 75238 Paris cedex 05, France.

factors such as cell size, paleoproductivity and CO₂ uptake mechanisms (Laws et al., 2001). Other promising approaches for reconstructing surface-ocean carbonate chemistry over the last ~100 Ma use proxies for paleo-pH or for paleo-carbonate ion (CO₃²⁻) concentrations. Boron isotope compositions ($\delta^{11}\text{B}$) (Rollion-Bard et al., 2011) of foraminifera shells have been shown to be a proxy of ocean paleo-pH (e.g., Foster, 2008; Pearson and Palmer, 2000; Pearson et al., 2009; Sanyal et al., 1995; Seki et al., 2010). However, pH alone is not sufficient to calculate the complete ocean carbonate chemistry since two parameters (typically pH and alkalinity or DIC) are needed in order to calculate the CO_{2(aq)} from which atmospheric pCO₂ can be estimated. Previous studies generally assumed a conservative ratio between alkalinity and salinity at the million-year scale. However, this approach may result in significant uncertainties in the final pCO₂ estimates because the alkalinity–salinity relationship of the past ocean could have varied. More recently it was suggested that B/Ca ratio in benthic foraminifera may be used to estimate ΔCO_3^{2-} (i.e. the deviation of seawater from calcite saturation; e.g., Rae et al., 2011; Yu and Elderfield, 2007; Yu et al., 2010), but both $\delta^{11}\text{B}$ and B/Ca are associated with the complex variability of boron speciation and its incorporation into calcite crystals (Klochko et al., 2009). In this study, we describe a new foraminiferal proxy for DIC in the past ocean: the ratio between ⁷Li and ⁶Li isotopes, expressed as $\delta^7\text{Li}$ ($\delta^7\text{Li} = 1000 \times [{}^7\text{Li}/{}^6\text{Li}] / [{}^7\text{Li}/{}^6\text{Li}]_{\text{LSEVC}} - 1$), in their calcite shells, which can provide additional constraints for the quantification of the past carbonate system in the ocean and past atmospheric CO₂.

Present-day seawater is homogenous with respect to Li concentration (175 $\mu\text{g/L}$) and isotopic composition ($\delta^7\text{Li} = 31.2 \text{ ‰}$, Millot et al., 2004). This is explained by the long residence time of lithium in the ocean relative to the oceanic water mixing time (see review in Burton and Vigier, 2011; Tomascak, 2004). Li isotopes fractionate significantly during low and high temperature geochemical processes, in particular during weathering of silicate rocks (Schmitt et al., 2012). Waters are systematically enriched in the heavy isotope (⁷Li) relative to fresh rocks and secondary phases (e.g., Huh et al., 2001; Vigier et al., 2008). The inverse trend between river $\delta^7\text{Li}$ and silicate weathering rates, highlighted by most watershed studies (Kisakurek et al., 2004; Pogge von Strandmann et al., 2006; Vigier et al., 2009), could make Li isotope composition of past oceans a proxy of variation of continental weathering through time, and therefore of corresponding atmospheric CO₂ consumption. Thus, Hathorne and James (2006) and Misra and Froelich (2012) have interpreted foraminifera $\delta^7\text{Li}$ in terms of variation of continental weathering for the period 0–18 Ma and 0–70 Ma, respectively. In addition, von Strandmann et al. (2013) interpreted $\delta^7\text{Li}$ variation measured in OAE2 (Ocean Anoxic Event, 93.5 Ma ago) carbonates by an increased weathering rate. Carbonates are only a minor sink of oceanic lithium. However, depending on how foraminifera record the Li isotope composition of the ocean at the time of their shell formation, they could provide precious information either:

- on variations of the main ocean Li sources (hydro-thermal activity and continental flux) and sinks

(low temperature oceanic crust weathering and clay formation) or;

- on environmental conditions at the time of their formation.

The latter can be envisioned if the dependence of Li isotope fractionation on environmental parameters such as ocean temperature (T), pH or DIC during foraminiferal calcification is known. For periods longer than 1 Ma (the oceanic Li residence time), both the variations of oceanic $\delta^7\text{Li}$ and of environmental conditions may influence the foraminifera Li/Ca and Li isotope compositions. In order to determine separately the role of each of these parameters on Li isotope fractionation, benthic foraminifera of the genus *Amphistegina* were grown at various temperatures (18 °C, 21 °C, 24 °C, 33 °C), various pH (7.90, 8.10, 8.20, 8.45) with constant DIC, and various DIC concentrations (1.0, 1.5, 2.0, 2.5, 3.0 mmol·kg⁻¹) at constant pH and T .

2. Methods

Amphistegina is a large epibenthic symbiont-bearing, calcitic–radial perforate genus, which is abundant in coral reef environments. Before the experiments, live *Amphistegina* specimens were collected from *Halophila* leaves at a depth of ~8 m or from turf algae on stones in the northern Gulf of Eilat (Red Sea) in February 2005. Two different species were used for the following experiments: *A. lessonii* for temperature experiments and *A. lobifera* for pH and DIC experiments. These two species present the same physical characteristics (knob and keel areas) and seem to have the same geochemical behavior (see, for example, Segev and Erez, 2006). Live specimens were then placed in sealed Erlenmeyer bottles filled with natural seawater modified in order to reach the target values of pH and DIC (see below and Rollion-Bard et al., 2008). After the experiments, Li isotopes and Li/Ca ratios were measured in the foraminifera shells using the ims 1270 ion microprobe, following a technique previously developed (Rollion-Bard et al., 2009; Vigier et al., 2007).

2.1. Cultures performed at various temperatures

The temperature experiments were carried out with live *Amphistegina lessonii*, collected from turf algae on stones in the coral reef environment in the Gulf of Eilat, Israel. Uniform populations of ~400 μm of diameter were placed in sealed 60-mL glass stoppered Erlenmeyer bottles filled with natural seawater. The natural seawater was surface-nutrient depleted-water from the Gulf of Eilat. This seawater has an initial salinity of 40.7‰ that was adjusted to 35‰ (by adding distilled water). The Erlenmeyer bottles were immersed for 2–3 months in temperature-controlled baths (at 18, 21, 24, 27, 30, and 33 °C respectively), keeping the assigned temperatures to $\pm 0.1 \text{ °C}$. Water in each bottle was replaced twice a week with fresh seawater. The pH in these experiments ranged between 8.1 and 8.2 with lower pH values at the higher temperatures due to faster calcification. All bottles received the same amount of natural light (~10–15 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$) and were examined daily for the development of algae. The foraminifera in these experiments

increased their weight from 50 μg /individual to between 95 and 135 μg /individual, with optimal growth at 27 °C.

2.2. Cultures performed at various pH

Amphistegina lobifera, collected similarly as above, were used for the cultures undertaken at various pH. The open Gulf of Eilat water used in the experiments has an average DIC of $2070 \pm 10 \mu\text{mol}\cdot\text{kg}^{-1}$ (Silverman et al., 2007), and the original alkalinity is roughly $2500 \mu\text{mol}\cdot\text{kg}^{-1}$. These waters were diluted to a salinity of 35 ‰ with double-distilled water (DDW), and were boiled to remove CO_2 . The DIC and the original alkalinity were thus diluted by the salinity factor of 35/40.7 (as explained above); hence the DIC was roughly $1780 \mu\text{mol}\cdot\text{kg}^{-1}$. These experiments were done at 24 ± 0.1 °C and the carbonate chemistry was modified by keeping the DIC constant, while the pH was lowered with HCl or raised with freshly made NaOH. The seawater reservoirs were kept out of contact with the atmosphere in plastic collapsible containers. Water was pumped with a peristaltic pump from the containers into tightly capped 60-mL Erlenmeyer flasks containing the foraminifera at flow rates between 160 and 180 mL/day. The cumulative alkalinity of the out-flowing water was measured every three days and the calcification of the foraminifera was estimated from the alkalinity difference between the reservoir and the outflow. In addition, the weight increase of the foraminifera was estimated from the final weight of the individuals compared to that of the initial control group. The weight increased on average by a factor of 5 (from 18 μg /individual to about 100 μg /individual). The different pH values for this experiment were 7.90, 8.10, 8.20 and 8.45 ± 0.10 (NBS scale).

2.3. Cultures performed at various Dissolved Inorganic Carbon contents

Amphistegina lobifera were used for the cultures undertaken at various DIC. The diluted Gulf of Eilat seawater was acidified with HCl to pH 2.8 and air bubbled for 12 h to get rid of most of the dissolved inorganic carbon. Then, NaHCO_3 was added to get final concentrations of DIC of 1.0, 1.5, 2.0, 2.5, and 3.0 $\text{mmol}\cdot\text{kg}^{-1}$, respectively. For comparison, the DIC range in the ocean is 1.8–2.4 $\text{mmol}\cdot\text{kg}^{-1}$. The pH was adjusted with freshly prepared 1 N NaOH and 1 N HCl in order to obtain values of 8.15 for all of these waters with different DIC. The pH values ended up with a small variability of ≈ 0.01 pH units. The reagents used were the same as those for varying the pH in the previous experiments except for the NaHCO_3 (section 2.2). The water reservoirs were isolated from the atmosphere inside collapsible plastic reservoirs connected to a peristaltic pump that pumped the diluted modified seawater solutions through each Erlenmeyer flask where the foraminifera were growing. The water coming out of the Erlenmeyer was collected and weighted (every 2 or 3 days) and measured for its alkalinity. Occasionally, we also measured oxygen and pH in the water coming out of the Erlenmeyer flasks. The alkalinities were close to the original ones, but always slightly lower (because of the foraminiferal calcification). The pH of the outcoming waters was slightly different from that in

the reservoir because of the metabolism of the foraminifera.

2.4. Li isotope and Li/Ca measurements in foraminifera tests

After the experiment, the cultured foraminifera were rinsed with double-distilled water (DDW) and dried overnight at 50 °C. Specimens were then placed in dilute sodium hypochlorite (1:3) for 7 h followed by several DDW rinses and overnight drying at 50 °C. Before analyses, samples were embedded in epoxy sections, polished using a diamond paste down to 1 μm and then coated with gold. Samples were put horizontal and were polished until the appearance of the top part of the knob (Rollion-Bard et al., 2008). Thus, only the sample part grown under laboratory conditions was analyzed with the ion probe.

Li isotope analyses and Li/Ca ratios were performed in situ with the ims 1270 ion probe at CRPG (Nancy, France). The analytical settings in this study are the same as those described in Vigier et al. (2007) and in Rollion-Bard et al. (2009). Briefly, the lithium isotopic compositions and Li/Ca ratios were measured using a 20- to 30- μm ^{16}O primary beam of ~ 50 nA. The mass resolving power (MRP) was set at ~ 3000 , with the energy slit well centered and fully opened. A single Li isotope analysis consists of peak switching on masses 6 and 7, which are successively counted on an electron multiplier. After 2 min of pre-sputtering, each run lasts about 20 min (40 to 60 cycles of 12 s and 10 s, respectively). The background is measured every cycle for 2 s on mass 5.5. The instrumental mass fractionation (IMF) and the reproducibility of the isotopic analyses were calculated from repeated measurements of the CAL-HTP calcite reference material that was also analyzed by MC-ICPMS for Li isotopes ($\delta^7\text{Li} = 13 \pm 0.5\%$, 2σ). The isotopic homogeneity of this sample was confirmed by multiple ion microprobe measurements along profiles (Vigier et al., 2007), and the reproducibility is better than 1% (1σ , $n = 10$).

For Li/Ca ratios, the analyses consist of peak switching on masses 7 ($^7\text{Li}^+$) and 44 ($^{44}\text{Ca}^+$), which are successively measured on an electron multiplier and on a Faraday cup. The Li content was also measured in a glass standard (384 ppm Li, and 0.57 W.% CaO). No difference in the yield was observed between carbonate and glass standards. Backgrounds are measured every cycle for 2 s on masses 5.5 and 43.6 to ensure magnet stability. Li/Ca was analyzed in the same spot as $\delta^7\text{Li}$. External (spot-to-spot) reproducibility for Li/Ca ratios was determined from repeated measurements of the glass reference material and was better than 1% (1σ). The total precision for each Li/Ca analysis includes external reproducibility and internal precision.

3. Results

3.1. Assessing T and pH controls

Li/Ca and $\delta^7\text{Li}$ data are reported in Tables 1, 2 and 3. $\delta^7\text{Li}$ measured in the *Amphistegina* shells display no significant variation as a function of temperature

Table 1

Results for the series of cultures performed at constant temperatures (24 °C), constant DIC (1.8 mmol·kg⁻¹) and different pH values.

pH	Li/Ca (μmol·mol ⁻¹)	δ ⁷ Li (‰)
7.9		30.2
7.9		29.0
7.9		30.2
7.9	14.1	29.4
7.9	12.7	31.1
7.9	14.8	29.1
7.9	13.3	29.5
8.1		29.8
8.2	15.5	29.8
8.2		29.9
8.45		29.8
8.45	13.5	29.8
8.45	12.9	29.4

and pH (Fig. 1a and b). For both sets of experiments, mean δ⁷Li values are close to the δ⁷Li of seawater (30.0 ± 0.9‰ and 29.8 ± 0.6‰ respectively, Fig. 1). This strongly suggests little isotope fractionation relative to the seawater – within uncertainty – for the given set of conditions. However, this is significantly different from the fractionation of Li isotopes observed during experimental growth of inorganic calcite, and strongly suggests a vital effect in favor of the heavy ⁷Li isotope (Δ⁷Li calcite–solution = –3‰ to –9‰, see Marriott et al., 2004a, 2004b). These results are also consistent with part of the published core top foraminifera data, which exhibit the same isotope compositions as seawater. However, pH and *T* cannot explain the significant range of δ⁷Li values measured in core top planktonic species (e.g., Hathorne and James, 2006; Misra and Froelich, 2012; Vigier et al., 2007).

Similarly, Li/Ca measured in the same tests exhibit no dependency on *T* and pH, with mean values of 16.4 ± 1.3 μmol·mol⁻¹ and 13.8 ± 2 μmol·mol⁻¹, respectively (Tables 1 and 2, Fig. 2). For comparison, the Li/Ca ratio of seawater is currently 2365 μmol·mol⁻¹. The lack of correlation with temperature is in agreement with studies of core top planktonic species (Hathorne and James, 2006). However, Lear and Rosenthal (2006), Lear et al. (2010), and Bryan and Marchitto (2008) have suggested that concentration in CO₃²⁻ could control the Li/Ca ratio of benthic foraminifera. Here, no significant influence of the pH is seen on both Li/Ca and δ⁷Li values. In this experiment performed at various pHs, but constant DIC and *T*, the CO₃²⁻ ion

Table 2

Results for the series of cultures performed at constant pH (8.15), constant DIC (1.8 mmol·kg⁻¹) and different temperatures.

Temperature (°C)	Li/Ca (μmol·mol ⁻¹)	δ ⁷ Li (‰)
18	16.3	29.8
18	17.4	30.0
18	15.8	30.7
21	18.3	30.9
24	17.6	28.7
24	16.8	29.6
24	14.5	30.3
33	14.4	28.3
33	15.9	30.4
33	17.6	31.1
33	15.9	29.8

Table 3

Results for the series of cultures performed at constant temperature (24 °C) and different DIC.

DIC mmol·kg ⁻¹	Li/Ca (μmol·mol ⁻¹)	δ ⁷ Li (‰)
1.0	19.7	21.6
1.5	15.3	23.2
1.5	14.3	26.3
1.5	15.7	27.2
2.0	14.4	32.7
2.0	16.5	31.2
2.0	14.4	31.2
2.0		32.1
2.5	3.6	41.9
2.5	3.4	40.7
3.0	5.9	37.7
3.0	6.0	37.7

concentration changed by 0.4 mmol·kg⁻¹ and this did not affect the Li/Ca ratio nor the δ⁷Li. We can therefore reasonably rule out a key role of this parameter, at least for *Amphistegina* species. Similarly, Hathorne et al. (2009) show no significant influence of the carbonate ion concentration on Li/Ca measured in situ by LA ICPMS in the calcitic tests of two different planktonic species.

3.2. Assessing DIC control

In contrast to temperature and pH (or CO₃²⁻), both δ⁷Li and Li/Ca ratio exhibit large variability and good linear correlations with the solution DIC (Figs. 1 and 2). *Amphistegina* δ⁷Li values positively and linearly correlate with DIC. There is a strong dependency on DIC, with 20‰ differences between foraminifera tests cultured at 1.0 mmol·kg⁻¹ DIC and at 2.5 mmol·kg⁻¹ DIC. In parallel, Li/Ca ratios increase from ~4 to ~20 μmol·mol⁻¹ as DIC decreases from 2.5 to 1.0 mmol·kg⁻¹ (Fig. 2, Table 3). For both δ⁷Li and Li/Ca the experimental data corresponding to the highest DIC value (3.0 mmol·kg⁻¹) falls off of these linear trends. We can rule out a potential contamination from the reagents used in the DIC experiments, for several reasons. First, we used the same reagents as for pH experiments, except HCO₃⁻. We added HCO₃⁻ in order to obtain the required DIC value, but if this procedure would have resulted in an increase of solution Li content, then the Li/Ca ratio in the shells should be positively correlated with DIC. The actual trend is the opposite (see Fig. 3). Moreover, since the same HCO₃⁻ reagent was added for both low- and high-DIC experiments, the variation of δ⁷Li going from values below the seawater Li isotope composition to values higher than seawater δ⁷Li could not be explained by the addition of this reagent. Thus, the increase in the δ⁷Li and decrease in the Li/Ca ratios must be caused by the DIC increase and related mechanism of calcite precipitation.

4. Discussion

4.1. Biological control on foraminifera δ⁷Li

Our cultures indicate the key role of the concentration of dissolved inorganic carbon on the Li isotope composition and on the Li/Ca ratio of *Amphistegina* tests. Figs. 1c and 2

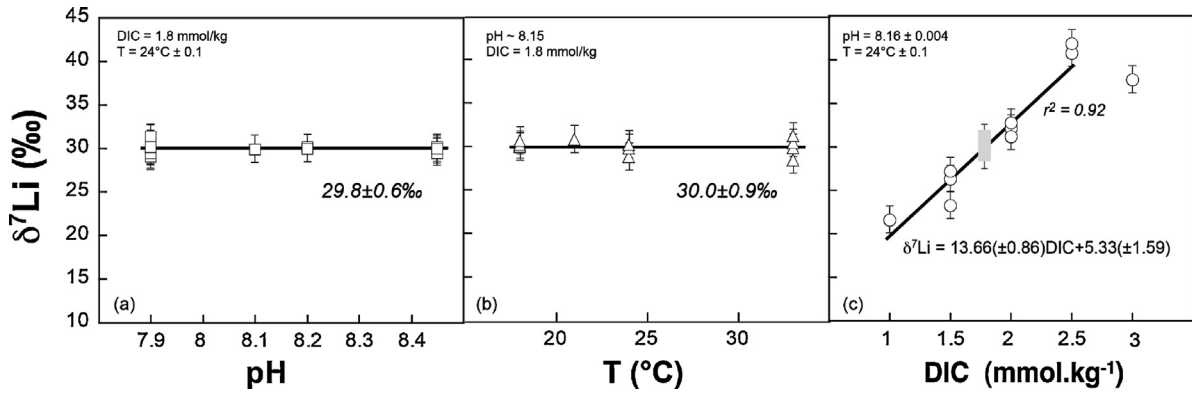


Fig. 1. $\delta^7\text{Li}$ measured in *Amphistegina* tests as a function of culture pH (a), culture temperature (b) and culture dissolved inorganic carbon (c) (see Tables 1–3). The linear regression and corresponding errors on the slope and intercept shown in Fig. 1c were obtained using Axum50 software. The grey rectangle corresponds to the range given by *Amphistegina* grown at various T and pH. The linear regression does not include the point obtained at high DIC ($3 \text{ mmol}\cdot\text{kg}^{-1}$), and is therefore valid for DIC ranging between 1 and $2.5 \text{ mmol}\cdot\text{kg}^{-1}$.

show that, for given T and pH values, at low DIC, the calcite Li/Ca ratio is high and the $\delta^7\text{Li}$ value is low. As described previously, we first rule out a potential influence of the carbonate ion (CO_3^{2-}) concentration. The foraminifera grown at various pH with constant DIC experienced significant CO_3^{2-} variability, yet their Li/Ca and Li isotope compositions remained constant (Fig. 1a). The same argument can be made with respect to alkalinity: the range of alkalinities in the pH experiments was significant ($0.4 \text{ mmol}\cdot\text{kg}^{-1}$) and again the $\delta^7\text{Li}$ and Li/Ca were similar, within uncertainties. One possibility to explain the dependence of *Amphistegina* Li/Ca and $\delta^7\text{Li}$ on DIC concentrations is that it may be related to the physiology of calcification in perforated foraminifera (Bentov et al., 2009, Erez, 2003). In previous studies, it has been shown that seawater vacuoles provide the fluid from which the shell is precipitated (e.g., Erez, 2003). In order to precipitate CaCO_3 from these seawater vacuoles, the foraminifera have to concentrate DIC until the CO_3^{2-} content matches (at least partially) the Ca^{2+} concentration in the seawater vacuoles, which is between 10 and 11 mM. This internal carbon pool has been thoroughly described using ^{14}C pulse-chase experiments with *A. lobifera* (ter Kuile and Erez, 1987, 1988; ter Kuile et al., 1989). In order to concentrate DIC in the vacuoles, the foraminifera elevate their pH and the DIC is increased by diffusion of $\text{CO}_{2(\text{aq})}$ from the cytosol into the vacuolated seawater (Bentov et al., 2009, Erez, 2003). Elevation of pH in the vacuoles and

the calcifying fluid may be achieved through several ways. For example, Ca-ATPase has often been suggested as one potential candidate in corals (e.g., Al-Horani et al., 2003; McConnaughey, 1989; Venn et al., 2013). However, there are other possibilities for such proton transporters, which are protein complexes that transport protons against the concentration gradient (note that the cytosol in which the vacuoles are residing has a low pH of about 7.2–7.4). Usually such processes require ATP, as an energy source, or utilize the cellular electrochemical gradient to achieve the needed result. We suggest that in foraminifera, this proton transport is achieved by an involvement of a Na/proton exchanger (antiporter) or by another pathway in which Na (and Li) ions are transported into the vacuole while protons are transported out into the cytosol. We propose a coupling between Na and Li because Li can often replace Na during such trans-membrane processes, due to similar chemical behavior and charge (Parker, 1986). It is interesting to note the non-dependency of Li and $\delta^7\text{Li}$ on the experimental temperature, suggesting that Li and Mg behaviors are decoupled, despite similar ionic radii (assuming that Mg/Ca and T are correlated). This indicates that the ion charge is an important parameter in biological activity (in contrast to abiotic reactions where Li^+ and Mg^{2+} are often coupled; e.g., Huh et al., 2001; Vigier et al., 2008). Our observations may therefore be explained as follows: when the solution DIC is low, the activity of the Na/proton exchanger is intensive in order to elevate the pH, the alkalinity, and

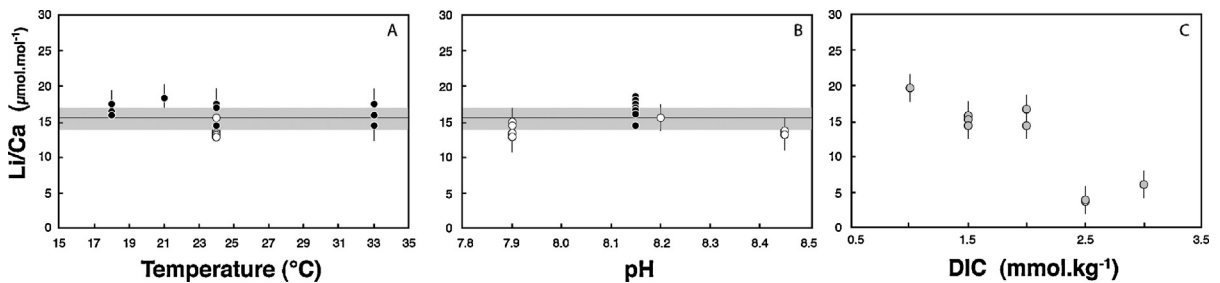


Fig. 2. Li/Ca measured in *Amphistegina* tests as a function of culture pH (open symbols) (B), culture temperature (black symbols) (A) and DIC (in grey circles) (C). The average of all values measured at various pH and T is $15.4 \pm 1.7 \mu\text{mol}\cdot\text{mol}^{-1}$ (grey line on A and B).

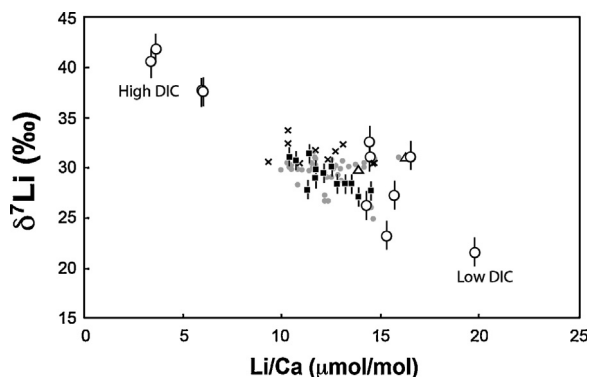


Fig. 3. $\delta^7\text{Li}$ versus Li/Ca measured in *Amphistegina* cultured at various DIC (white circles). The white triangles correspond to the average values given by the *Amphistegina* cultured at various T and pH (see Fig. 1a and b). On this plot, are also reported the published data for various species of planktonic foraminifera: squares, core top from Hathorne and James (2006); grey circles, core top from Misra and Froelich, (2012); crosses, Glacial/Interglacial from Hall et al. (2005).

therefore to accumulate DIC in the vacuoles. While this is happening, Li follows Na and accumulates in the vacuoles. Under these conditions, the lighter isotope (^6Li) may be preferentially transported into the vacuoles, which would explain the high Li/Ca ratio associated with the low $\delta^7\text{Li}$ values (Fig. 3). When CaCO_3 is precipitated from modified seawater, it records its Li concentration and its $\delta^7\text{Li}$ value. Further speculations on this mechanism are perhaps too early, but it explains well our experimental observations and is in good agreement with the models suggested for foraminiferal calcification (Bentov and Erez, 2005; Bentov et al., 2009; Erez, 2003).

When DIC is high, calcite $\delta^7\text{Li}$ is also higher (higher than seawater value, 31‰) and Li/Ca is lower (Fig. 3). Published $\delta^7\text{Li}$ values for core-top foraminifera tests higher than 31‰

(seawater) are rare, but exist (see, e.g., Hall et al., 2005; Kosler et al., 2001). Actually, these values are still consistent with a vital effect in favor of the heavy ^7Li isotope, as underlined in section 3.1 and shown by T and pH experiments (Fig. 1a and b), compared to inorganic calcite (see section 3.1). Under high DIC conditions, we can speculate that there is no need to increase the DIC in order to calcify, and therefore, the high $\delta^7\text{Li}$ observed in the *Amphistegina* tests cultured in these conditions likely reflects the isotope composition of the solution at the calcifying site. The reaction responsible for this increase may also be either kinetic or associated with a Rayleigh-type isotope fractionation occurring in the cytosol with preferential release of the light ^6Li isotope, and decrease in Li/Ca ratio. A more refined biological study of the mechanisms occurring at high DIC is underway.

4.2. Applications to natural conditions

Amphistegina are symbiont-bearing foraminifera, belonging to the calcitic-radial perforate group, and thus are thought to precipitate their calcite using the same mechanisms as most planktonic foraminifera (Erez, 2003). This was illustrated by boron isotope ratios measured on cultured *Amphistegina* tests, using SIMS, which can be explained by a pH range similar to the one measured by pH microsensors in living planktonic *Globigerinoides sacculifer* (Jorgensen et al., 1985; Rollion-Bard and Erez, 2010). In order to test this for Li isotopes, we reported in Fig. 3 all the Li isotopes and concentration data published for various species of recent and core top planktonic foraminifera. The data plot along the trend defined by the *Amphistegina* cultured in this study. As a consequence, we consider that the trend between *Amphistegina* $\delta^7\text{Li}$ and DIC may also be valid for the various planktonic foraminifera species studied thus far. It should therefore be possible to calculate

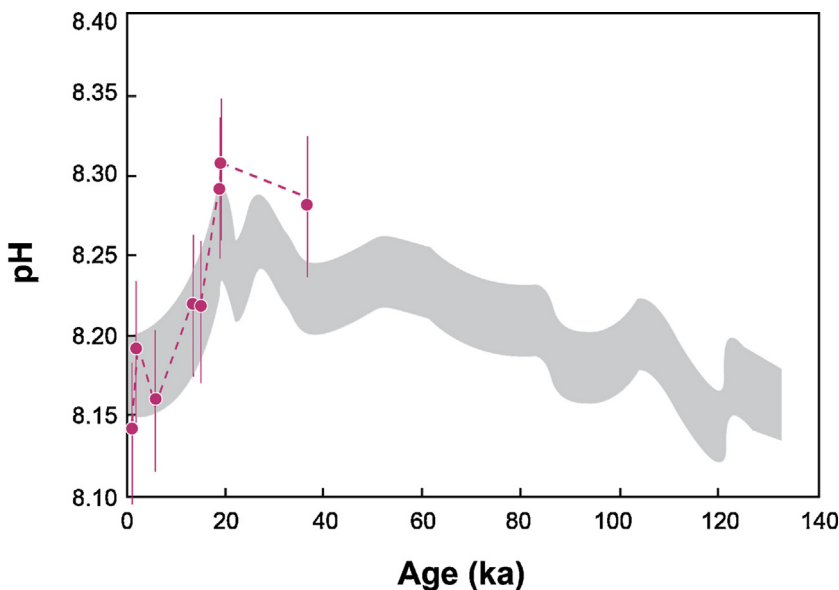


Fig. 4. (Color online.) pH values (in pink) calculated from DIC using the $\delta^7\text{Li}$ measured in foraminifera for the 0.4–35.8-ka period (Hall et al., 2005) and the pCO_2 values taken from ice core (Monnin et al., 2001). The calculated pHs are compared to the pH variation determined by boron isotopes and B/Ca, shown in grey (Foster, 2008).

the DIC at the time of the foraminifera test formation, using the linear regression of the experimental data points of *Amphistegina* (Fig. 1c). In addition, the DIC control evidenced in this study may partly explain the large published range of $\delta^7\text{Li}$ measured for recent and sub-recent foraminifera (21.1–33.7‰, Hall et al., 2005; Hathorne and James, 2006; Marriott et al., 2004b; Misra and Froelich, 2012).

For short periods of time, such as for the last glacial/interglacial period, the ocean $\delta^7\text{Li}$ has remained constant since the ocean Li residence time is larger than 1 Ma. However, foraminifera exhibit $\delta^7\text{Li}$ variations during the last ~40 ka (Hall et al., 2005). From 400 yr to 35.8 ka, the foraminifera $\delta^7\text{Li}$ range is 30.4–33.7‰, which corresponds to a narrow DIC range (1.82–2.08 mmol·kg⁻¹), using the empirical relationship determined experimentally. For this

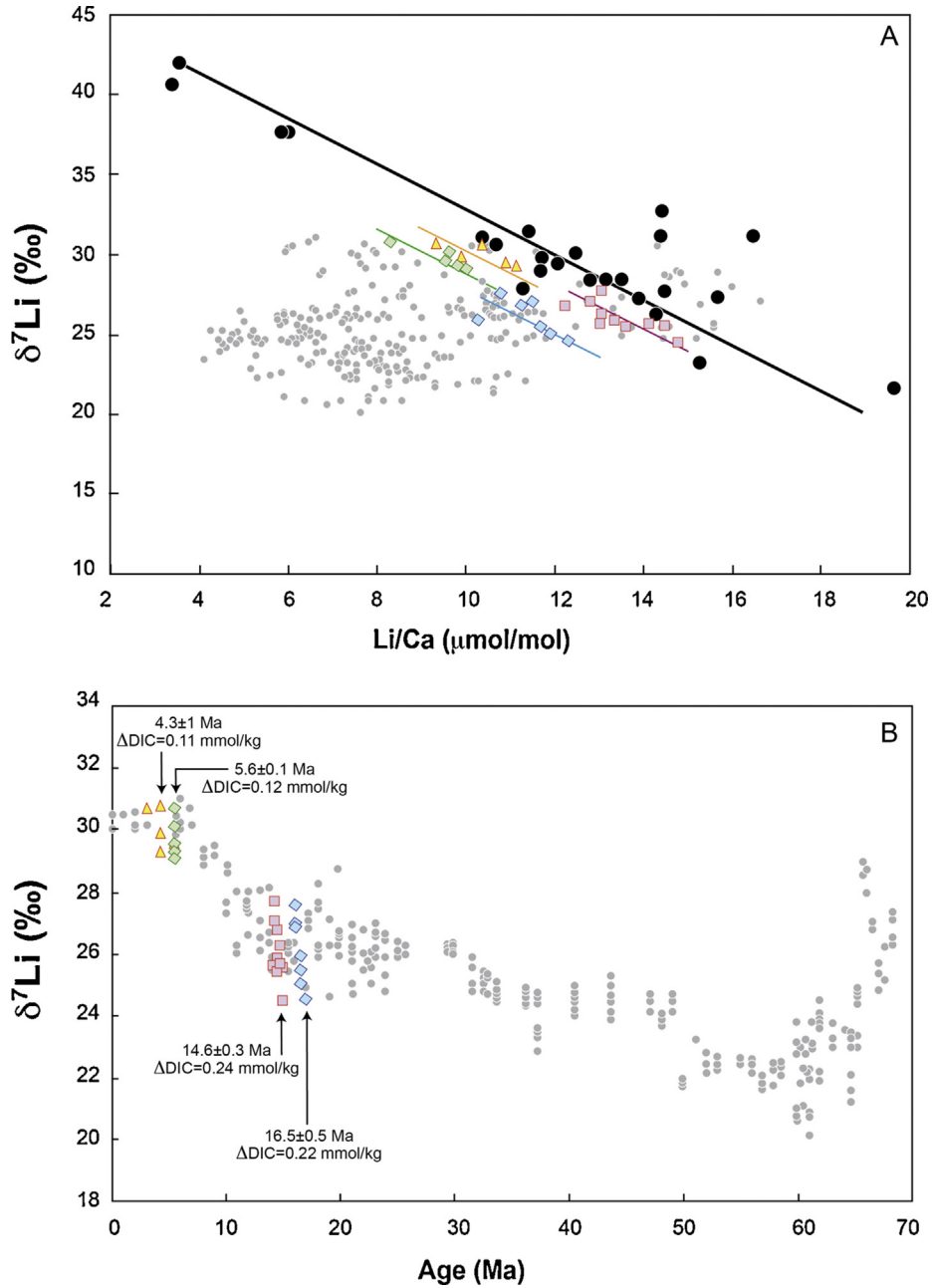


Fig. 5. (Color online.) A. $\delta^7\text{Li}$ versus Li/Ca for *Amphistegina* cultured with various DIC, and for core-top planktonic foraminifera (black circles: this study, Hathorne and James, 2006), and for 0–70 Ma foraminifera (grey circles and colored symbols, Misra and Froelich, 2012). Yellow triangles highlight all values displayed for the 3.15–5.5 Ma period, green diamonds for the 5.5–5.61 Ma period, blue diamonds for the 14.25–15 Ma period, and pink squares for the 16.1–17 Ma period; B. The 0–70 Ma foraminifera record of Misra and Froelich (2012) (grey circles) as a function of time. The periods highlighted in color in Fig. 5A are reported here using the same symbols (see text for more details). The corresponding maximum DIC variations (ΔDIC) have been calculated using the equation given in Fig. 1c, and assuming no variation of ocean $\delta^7\text{Li}$.

period of time, the pCO₂ range is well known, since it is given by direct measurement of CO₂ trapped in air inclusions analyzed in the Vostok ice cores (Monnin et al., 2001). Using the calculated DIC and the following equation:

$$p\text{CO}_2 = \frac{(\text{H}^+)^2}{[(\text{H}^+)^2 + K_1(\text{H}^+) + K_1K_2] \cdot \text{DIC}} / K_H \quad (1)$$

with K_1 , K_2 the acidity constants and K_H the Henry's law constant, it is then possible to calculate the ocean pH in equilibrium with the pCO₂ measured in Vostok ice at a given time (Table S1). The pH calculated using this method ranges between 8.32 (±0.04) at the LGM (Last Glacial Maximum) and 8.17 ± 0.04 for the present interglacial. This range is similar to the range published by Foster (2008), which was determined using B isotopes and B/Ca measured in *G. ruber* for the same period (Fig. 4). This similarity suggests that the empirical relationship linking δ⁷Li and DIC is reliable. It is also in agreement with elevated pH at the LGM (Sanyal et al., 1995), and with lower atmospheric pCO₂ during glacial time (Broecker and Peng, 1992).

The 0–70 Ma record of δ⁷Li in foraminifera (Misra and Froelich, 2012) is also characterized by several periods of significant short-term variability (see Fig. 5b). This is particularly visible during the Miocene, between 11 Ma and 24 Ma (with, e.g., a 3.2‰ range of the δ⁷Li values between 14.25 Ma and 15 Ma). Since the Li residence time in the ocean is roughly 1 Ma, such rapid fluctuations cannot be explained by ocean variations due to changes in source or sink. Instead, our experimental results suggest that they can be related to changes of the Li isotope fractionation during foraminiferal growth, due to rapid variations of the DIC in seawater. We show that, for four critical periods, ranging between 16.5 Ma and 4.3 Ma, the foraminifera δ⁷Li and Li/Ca are linearly correlated and can be fitted by a similar slope as the one defined by the cultured *Amphistegina* and the core-top foraminifera (Fig. 3 and 5a). This strongly supports a DIC control on foraminiferal δ⁷Li at these times, which also correspond to critical periods of the Earth's history, i.e. the beginning of the Paleocene and the Mid-Miocene climatic optimum. The corresponding DIC variations range between 0.10 mmol·kg⁻¹ in the Pliocene up to 0.22 mmol·kg⁻¹ at the Miocene.

5. Conclusion

Using foraminifera cultures, we determined the influence of T , pH and DIC concentration on their calcite-seawater Li isotope fractionation and Li/Ca ratio. No significant influence of T and pH were observed, and the apparent lack of significant isotope fractionation suggests a biological control. A highly significant linear DIC-δ⁷Li relationship was observed as well as with Li/Ca ratio. These findings were used to re-interpret parts of published Cenozoic records in terms of DIC changes in the past ocean. The precision of this method still needs to be improved, but the first results are consistent – within uncertainties – with published past estimates of pCO₂ and pH. Over short timescales, shorter than the ocean Li residence time (1 Ma), the Li isotope composition of foraminifera provides a new proxy of the ocean DIC, which, combined with other

paleo-proxies of the carbonate system such as boron isotopes, may represent a great potential for reconstructing large and rapid variations of the past oceanic carbon cycle.

Acknowledgements

The authors thank the Editor François Chabaux, and the two reviewers, Franck Bassinot and Paul Tomaschak for their constructive comments. Marc Chaussidon (CRPG, Nancy), Christian France-Lanord (CRPG, Nancy), Yehoshua Kolodny (University of Jerusalem), and Greg Ravizza (University of Hawaii) for fruitful discussions on an earlier version of the manuscript. Pete Burnard (CRPG) is thanked for English corrections. This project was funded by a regional grant (CNRS/Lorraine Region), by CNRS-INSU programs (SYSTER, INTERRVIE and LEFE-CYBER) and by the Israel Science Foundation (grants 551/10 and 870/05).

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.crte.2014.12.001>.

References

- Al-Horani, F.A., Al-Moghrabi, S.M., De Beer, D., 2003. The mechanism of calcification and its relation to photosynthesis and respiration in the scleractinian coral *Galaxea fascicularis*. *Mar. Biol.* 142, 419–426.
- Bentov, S., Erez, J., 2005. Novel observations on biomineralization processes in foraminifera and implications for Mg/Ca ratio in the shells. *Geology* 33, 841–844.
- Bentov, S., Brownleeb, C., Erez, J., 2009. The role of seawater endocytosis in the biomineralization process in calcareous foraminifera. *Proc. Natl. Acad. Sci. USA* 106, 21500–21504.
- Broecker, W.S., Peng, T.H., 1992. Interhemispheric transport of carbon dioxide by ocean circulation. *Nature* 356, 587–589.
- Bryan, S.P., Marchitto, T., 2008. The Mg/Ca-temperature proxy in benthic foraminifera: new calibrations from the Florida Straits and a hypothesis regarding Mg/Li. *Paleoceanography* 23, PA220, <http://dx.doi.org/10.1029/2007PA001553>.
- Burton, K.W., Vigier, N., 2011. Lithium isotopes as tracers in marine and terrestrial environments. In: *Handbook of Environmental Isotope Geochemistry*. Springer-Verlag, [315 p].
- Erez, J., 2003. The source of ions for biomineralization in foraminifera and their implications for paleoceanographic proxies. In: Dove, P.M., De Yoreo, J.J., Weiner, S. (Eds.), *Biomineralization*, *Rev. Mineral. Geochem.*, 54, pp. 115–149.
- Foster, G.L., 2008. Seawater pH, pCO₂ and [CO₃²⁻] variations in the Caribbean Sea over the last 130 kyr: a boron isotope and B/Ca study of planktic foraminifera. *Earth Planet. Sci. Lett.* 271, 254–266.
- Godderis, Y., Veizer, J., 2000. Tectonic control of chemical and isotopic composition of ancient oceans: the impact of continental growth. *Am. J. Sci.* 300, 434–461.
- Hall, J.M., Chan, L.-H., McDonough, W.F., Turekian, K.K., 2005. Determination of the lithium isotopic composition of planktic foraminifera and its application as a paleo-seawater proxy. *Mar. Geol.* 217, 255–265.
- Hathorne, E.C., James, R.H., 2006. Temporal record of lithium in seawater: a tracer for silicate weathering? *Earth Planet. Sci. Lett.* 246, 393–406.
- Hathorne, E.C., James, R.H., Lampitt, R.S., 2009. Environmental versus biomineralization controls on the intrateat variation in the trace element composition of the planktonic foraminifera *G. inflata* and *G. scitula*. *Paleoceanography* 24, <http://dx.doi.org/10.1029/2009PA001742>.
- Huh, Y., Chan, L.-H., Edmond, J.M., 2001. Lithium isotopes as a probe of weathering processes: Orinoco River. *Earth Planet. Sci. Lett.* 194, 189–199.
- IPCC, 2013. *Climate Change 2013: the physical science basis*. In: Stocker, T.F., Qin, D., Plattner, G.-K., Tignor, M., Allen, S.K., Boschung, J., Nauels, A., Xia, Y., Bex, V., Midgley, P.M. (Eds.), *Contribution of Working*

- Group I to the fifth assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, [1535 p].
- Jorgensen, B.B., Erez, J., Revsbech, N.P., Cohen, Y., 1985. Symbiotic photosynthesis in a planktonic foraminiferan, *Globigerinoides sacculifer* (Brady), studied with microelectrodes. *Limnology Oceanography* 30, 1253–1267.
- Kisakurek, B., Widdowson, M., James, R.H., 2004. Behaviour of Li isotopes during continental weathering: the Bidar laterite profile, India. *Chem. Geol.* 212, 27–44.
- Klochko, K., Cody, G.D., Tossell, J.A., Dera, P., Kaufman, A.J., 2009. Re-evaluating boron speciation in biogenic calcite and aragonite using ^{11}B MAS NMR. *Geochim. Cosmochim. Acta* 73, 1890–1900.
- Kosler, J., Kucera, M., Sylvester, P., 2001. Precise measurement of Li isotopes in planktonic foraminifera tests by quadrupole ICPMS. *Chem. Geol.* 181, 169–179.
- ter Kuile, B., Erez, J., 1987. Uptake of inorganic carbon and internal carbon cycling in symbiont-bearing benthonic foraminifera. *Mar. Biol.* 94, 499–509.
- ter Kuile, B., Erez, J., 1988. The size and function of the internal inorganic carbon pool of the foraminifer *Amphistegina lobifera*. *Mar. Biol.* 99, 481–487.
- ter Kuile, B., Erez, J., Padan, E., 1989. Competition for inorganic carbon between photosynthesis and calcification in the symbiont-bearing foraminifer *Amphistegina lobifera*. *Marine Biol.* 103, 253–259.
- Kürschner, W.M., Kvacek, Z., Dilcher, D.L., 2008. The impact of Miocene atmospheric carbon dioxide fluctuations on climate and the evolution of terrestrial ecosystems. *Proc. Natl. Acad. Sci. USA* 105, 449–453.
- Laws, E.A., Popp, B.N., Bidigare, R.R., Riebesell, U., Burkhardt, S., Wakeham, S.G., 2001. Controls on the molecular distribution and carbon isotopic composition of alkenones in certain haptophyte. *Geochim. Geophys. Geosyst.* 2, 1006. <http://dx.doi.org/10.1029/2000GC000057>.
- Lear, C.H., Rosenthal, Y., 2006. Benthic foraminiferal Li/Ca: insights into Cenozoic seawater carbonate saturation state. *Geol. Soc. Am.* 34 (11), 985–988.
- Lear, C.H., Mawbey, E.M., Rosenthal, Y., 2010. Cenozoic benthic foraminiferal Mg/Ca and Li/Ca records: toward unlocking temperatures and saturation states. *Paleoceanography* 25, PA4215.
- Marriott, C.S., Henderson, G.M., Crompton, R., Staubwasser, M., Shaw, S., 2004a. Effect of mineralogy, salinity, and temperature on Li/Ca and Li isotope composition of calcium carbonate. *Chem. Geol.* 212, 5–15.
- Marriott, C.S., Henderson, G.M., Belshaw, N.S., Tudhope, A.W., 2004b. Temperature dependence of $\delta^7\text{Li}$, $\delta^{44}\text{Ca}$ and Li/Ca during growth of calcium carbonate. *Earth Planet. Sci. Lett.* 222, 615–624.
- McConnaughey, T., 1989. ^{13}C and ^{18}O isotopic disequilibrium in biological carbonates: II in vitro simulation of kinetic isotope effects. *Geochim. Cosmochim. Acta* 53, 163–171.
- Millot, R., Guerrot, C., Vigier, N., 2004. Accurate and high-precision measurement of lithium isotopes in two reference materials by MC-ICP-MS. *Geostand. Geoanal. Res.* 28 (1), 153–159.
- Misra, S., Froelich, P.N., 2012. Lithium isotope history of Cenozoic seawater: changes in silicate weathering and reverse weathering. *Science* 335, 818–823.
- Monnin, E., Indermühle, A., Dällenbach, A., Flückiger, J., Stauffer, B., Stocker, T.F., Raynaud, D., Barnola, J.-M., 2001. Atmospheric CO_2 concentrations over the last glacial termination. *Science* 291, 112–114.
- Pagani, M., Zachos, J.C., Freeman, K.H., Tiplle, B., Bohaty, S.M., 2005. Marked decline in atmospheric carbon dioxide concentrations during the Paleogene. *Science* 309, 600–603.
- Parker, J.C., 1986. Interactions of Lithium and Protons with the Sodium-Proton exchanger of dog red blood cells. *J. Gen. Physiol.* 87, 189–200.
- Pearson, P.N., Palmer, M.R., 2000. Atmospheric carbon dioxide concentrations over the past 60 million years. *Nature* 406, 695–699.
- Pearson, P.N., Foster, G.L., Wade, B.S., 2009. Atmospheric carbon dioxide through the Eocene-Oligocene climate transition. *Nature* 461, 110–1113.
- Pogge von Strandmann, P.A.E., Burton, K.W., James, R.H., van Calsteren, P., Gislason, S.R., Mokadem, F., 2006. Riverine behaviour of uranium and lithium isotopes in an actively glaciated basaltic terrain. *Earth Planet. Sci. Lett.* 251, 134–147.
- Rae, J.W.B., Foster, G.L., Schmidt, D.N., Elliott, T., 2011. Boron isotopes and B/Ca in benthic foraminifera: proxies for the deep ocean carbonate system. *Earth Planet. Sci. Lett.* 302, 403–413.
- Rollion-Bard, C., Erez, J., 2010. Intra-shell boron isotope ratios in the symbiont-bearing benthic foraminiferan *Amphistegina lobifera*: implications for $\delta^{11}\text{B}$ vital effects and paleo-pH reconstructions. *Geochim. Cosmochim. Acta* 74, 1530–1536.
- Rollion-Bard, C., Erez, J., Zilberman, T., 2008. Intra-shell oxygen isotope ratios in the benthic foraminifera genus *Amphistegina* and the influence of seawater carbonate chemistry and temperature on this ratio. *Geochim. Cosmochim. Acta* 72, 6006–6014.
- Rollion-Bard, C., Vigier, N., Meibom, A., Blamart, D., Reynaud, S., et al., 2009. Effect of environmental conditions and skeletal ultrastructure on the Li isotopic composition of scleractinian corals. *Earth Planet. Sci. Lett.* 286, 63–70.
- Rollion-Bard, C., Chausson, M., France-Lanord, C., 2011. Biological control of internal pH in scleractinian corals: implications on paleo-pH and paleo-temperature. *C. R. Geoscience* 343, 397–405.
- Sanyal, A., Hemming, N.G., Hansona, G.N., Broecker, W.S., 1995. Evidence for a higher pH in the glacial ocean from boron isotopes in foraminifera. *Nature* 373, 234–236.
- Schmitt, A.-D., Vigier, N., Lemarchand, D., Millot, R., Stille, P., Chabaux, F., 2012. Processes controlling the stable isotope compositions of Li, B, Mg and Ca in plants, soils and waters: a review. *C. R. Geoscience* 344, 704–722.
- Segev, E., Erez, J., 2006. Effect of Mg/Ca ratio in seawater on shell composition in shallow benthic foraminifera. *Geochim. Geophys. Geosyst.* 7 (2), <http://dx.doi.org/10.1029/2005GC000969>.
- Seki, O., Foster, G.L., Schmidt, D.N., Mckensen, A., Kawamura, K., Pancost, R.D., 2010. Alkenone and boron-based Pliocene pCO_2 records. *Earth Planet. Sci. Lett.* 292, 201–211.
- Siegenthaler, U., Stocker, T.F., Monnin, E., Lüthi, D., Schwander, J., et al., 2005. Stable carbon cycle-climate relationship during the Late Pleistocene. *Science* 310, 1313–1317.
- Silverman, J., Lazar, B., Erez, J., 2007. Effect of aragonite saturation, temperature, and nutrients on the community calcification rate of a coral reef. *J. Geophys. Res.* 112, C05004. <http://dx.doi.org/10.1029/2006JC003770>.
- von Strandmann, P., Jenkyns, H.C., Woodfine, R.G., 2013. Lithium isotope evidence for enhanced weathering during Oceanic anoxic event 2. *Nature Geoscience* 6, 668–672.
- Tomascak, P.B., 2004. Developments in the understanding and application of lithium isotopes in the Earth and Planetary Sciences. *Rev. Mineral. Geochem.* 55, 153–195.
- Venn, A.A., Tambutté, E., Holcomb, M., Laurent, J., Allemand, D., Tambutté, S., 2013. From the cover: impact of seawater acidification on pH at the tissue-skeleton interface and calcification in reef corals. *PNAS* 110 (5), 1634–1639.
- Vigier, N., Rollion-Bard, C., Spezzaferri, S., Brunet, F., 2007. In situ measurements of Li isotopes in foraminifera. *Geochim. Geophys. Geosyst.* 8, Q01003. <http://dx.doi.org/10.1029/2006GC01432>.
- Vigier, N., Decarreau, A., Millot, R., Carignan, J., Petit, S., France-Lanord, C., 2008. Quantifying Li isotope fractionation during smectite formation and implications for the Li cycle. *Geochim. Cosmochim. Acta* 72, 780–792.
- Vigier, N., Gislason, S.R., Burton, K.W., Millot, R., Mokadem, F., 2009. The relationship between riverine lithium isotope composition and silicate weathering rates in Iceland. *Earth Planet. Sci. Lett.* 287, 434–441.
- Yu, J., Elderfield, H., 2007. Benthic foraminiferal B/Ca ratios reflect deep-water carbonate saturation state. *Earth Planet. Sci. Lett.* 258, 73–86.
- Yu, J.M., Foster, G.L., Elderfield, H., et al., 2010. An evaluation of benthic foraminiferal B/Ca and $\delta^{11}\text{B}$ for deep-ocean carbonate ion and pH reconstruction. *Earth Planet. Sci. Lett.* 293, 114–120.